# Drug Therapy

ALASTAIR J.J. WOOD, M.D., Editor

# INTERACTIONS AMONG DRUGS FOR HIV AND OPPORTUNISTIC INFECTIONS

STEPHEN C. PISCITELLI, PHARM.D., AND KEITH D. GALLICANO, PH.D.

RUG interactions are an important factor in the treatment of patients with human immunodeficiency virus (HIV) infection. The complexity of current drug regimens for such patients requires that clinicians recognize and manage drug interactions. Antiretroviral drug regimens typically consist of three or four antiretroviral drugs but may include even more. In addition, patients may receive other drugs for supportive care, treatment of opportunistic infections, and immunomodulation, as well as alternative drugs obtained from health care providers other than their primary provider. Drug interactions are often unavoidable in HIV-infected patients because of the drug classes involved and the number of drugs prescribed. In this article we review the clinically important interactions among drugs used to treat HIV infection, provide an overview of the primary mechanisms of drug interactions, and discuss ways to prevent or minimize the adverse effects of such interactions on clinical care.

## MECHANISMS OF DRUG INTERACTIONS

Drug interactions can be either pharmacokinetic or pharmacodynamic in nature. Pharmacokinetic interactions alter the absorption, transport, distribution, metabolism, or excretion of a drug. In therapy for HIV infection, pharmacokinetic interactions are often multifactorial. They may involve alterations in drug metabolism mediated by the cytochrome P-450 system, modulation of P-glycoprotein (a cellular transport protein), changes in renal elimination, changes in gastric pH and drug absorption, and fluctuations in intracellular drug concentrations (Table 1). These processes may take place at various sites in the body (Fig. 1). Pharmacodynamic interactions alter the pharmacologic response to a drug. The response can be additive, synergistic, or antagonistic. Pharmacodynamic interactions do not always modify a drug's concentration in tissue fluids.

#### Metabolic Interactions

All HIV-protease inhibitors and non-nucleoside reverse-transcriptase inhibitors that have been approved by the Food and Drug Administration (FDA), as well as those that are investigational drugs, are metabolized by the cytochrome P-450 enzyme system, primarily by the 3A4 isoform (CYP3A4), and each of these drugs may alter the metabolism of other antiretroviral and concomitantly administered drugs.<sup>15</sup> The cytochrome P-450 system consists of at least 11 families of enzymes, classified by number, of which 3 (CYP1, CYP2, and CYP3) are important in humans.<sup>16</sup> The families are further divided into subfamilies, denoted by a capital letter (e.g., CYP3A). Individual proteins within a subfamily, called isozymes or isoenzymes, are identified by a second number (e.g., CYP3A4).<sup>16</sup>

Drugs can be classified as cytochrome P-450 substrates, inhibitors, or inducers. However, some drugs, such as ritonavir, nelfinavir, and efavirenz, may have properties of all three, depending on the specific combination (Table 2). Substrates are drugs metabolized through this enzyme system, and the plasma concentrations of such drugs may be increased or decreased by other drugs. Inhibition of cytochromes is usually reversible and competitive, in that the substrate and inhibitor compete for the same site on the enzyme. Inhibition also occurs by irreversible inactivation of the enzyme, leading to pharmacologic effects that are prolonged until new enzyme can be synthesized.<sup>19</sup> Drugs that inhibit cytochromes cause decreased clearance and increased plasma concentrations of substrate drugs, and the effects may be greater if inhibitory metabolites accumulate during multiple dosing.

Drugs that induce cytochromes increase the rate of hepatic metabolism of other drugs by increasing the transcription of cytochrome messenger RNA (mRNA), which in turn leads to the production of more enzyme and a corresponding decrease in plasma concentrations of drugs metabolized by the induced pathway. When a CYP3A4 inhibitor, such as ritonavir, is added to another protease inhibitor, such as saquinavir, plasma concentrations of the second protease inhibitor increase markedly (Fig. 2A), often allowing for more convenient dosing. Increased concentrations may also overcome viral resistance to the drug.<sup>21</sup> The addition of a CYP3A4 inducer, such as nevirapine, to indinavir or amprenavir results in a decrease in the area under the plasma concentrationtime curve of the protease inhibitor (a measure of total exposure). A substantial decrease could reduce trough plasma concentrations of the protease inhibitor to a level below the in vitro concentration required to inhibit replication of 50 percent of viral strains  $(IC_{50})$ , with the subsequent development of resistance.

# Intestinal Metabolism and P-Glycoprotein

The liver is the primary site of drug metabolism mediated by the cytochrome P-450 system, but CYP3A4

From the Clinical Pharmacokinetics Research Laboratory, Department of Pharmacy, Warren G. Magnuson Clinical Center, National Institutes of Health, Bethesda, Md. (S.C.P.); and the Clinical Investigation Unit and Ottawa Hospital Research Institute, Ottawa Hospital, Ottawa, Ont., Canada (K.D.G.). Address reprint requests to Dr. Piscitelli at Virco Laboratories, Johns Hopkins Bayview Campus, Alpha Ctr., 3rd Fl., 5210 Eastern Ave., Baltimore, MD 21224.

Mechanism	Examples	EFFECT	Consequences
Altered intracellular activation Impairment of phosphorylation	Ribavirin and zidovudine, <sup>1</sup> zido- vudine and stavudine, <sup>2</sup> zalcita- bine and lamivudine <sup>3</sup>	Interference with intracellular phos- phorylation (in vitro)	Potential for decreased effectiveness and treatment failure
Altered drug absorption and tissue distribution			
Chelation	Fluoroquinolones with antacids <sup>4,5</sup>	Marked reduction in quinolone AUC from formation of insoluble com- plexes	Reduced antimicrobial effect
Change in gastric pH	Indinavir and didanosine <sup>6</sup>	Impaired absorption of indinavir due to increased pH	Low plasma indinavir concentrations may lead to viral resistance and treat- ment failure
Induction of efflux transporters Inhibition of efflux transporters	Rifampin and digoxin <sup>7</sup> Ketoconazole with saquinavir and ritonavir <sup>8</sup>	Decrease in digoxin AUC Increased CSF concentrations of saquinavir and ritonavir in relation to unbound plasma concentrations	Reduced therapeutic effect Combination being studied to target drug delivery to CSF; clinical rele- vance unknown
Altered drug metabolism Induction of cytochrome P-450	Rifabutin and saquinavir <sup>9</sup>	Saquinavir AUC reduced by 47 per- cent	Low plasma saquinavir concentrations may lead to viral resistance and treat-
Inhibition of cytochrome P-450 (hepatic and gastrointestinal)	Ritonavir and indinavir <sup>10</sup>	Marked increases in indinavir AUC and trough concentration	Combination under study to optimize therapy and develop more conven- ient regimens for patients
Inhibition of cytochrome P-450 (gastrointestinal only)	Grapefruit juice and saquinavir <sup>11</sup>	Saquinavir AUC increased by 50 to 150 percent	Increased plasma saquinavir concentra- tions, but the effect is highly variable
Increase in glucuronosyltransferase	Rifampin and zidovudine <sup>12</sup>	Zidovudine AUC decreased by 47 percent	Clinical relevance unknown but may lead to reduced antiviral effect if tri- phosphate concentrations are also decreased
Reduced renal excretion	TMP-SMX and lamivudine <sup>13</sup>	Lamivudine AUC increased by 44 percent due to inhibition of tubu- lar secretion	Dosage alteration unnecessary, since increased lamivudine concentrations are unlikely to have toxic effects
Pharmacodynamic interactions Additive or synergistic interactions	Zidovudine and ganciclovir	Additive bone marrow suppression	May require discontinuation or re- duced doses of one or both drugs or addition of G-CSF
	Combination HAART therapy	Sustained viral suppression	Potent therapy associated with long- term clinical and immunologic im- provement
Antagonist or opposing interactions	Indinavir and saquinavir <sup>14</sup>	In vitro antagonism at high doses	Clinical consequences unclear

TABLE 1. COMMON MECHANISMS FOR DRUG INTERACTIONS IN PATIENTS WITH HIV INFECTION.*
---

\*AUC denotes the area under the concentration-time curve, CSF cerebrospinal fluid, TMP-SMX trimethoprim-sulfamethoxazole, G-CSF granulocyte colony-stimulating factor, and HAART highly active antiretroviral therapy.

is also present in the enterocytes of the small intestine.<sup>22</sup> Thus, drugs that inhibit CYP3A4 may alter intestinal or hepatic metabolism of other drugs. The 20-fold increase in plasma concentrations of saquinavir caused by ritonavir is probably produced by inhibition of CYP3A4 at both sites.20 Grapefruit juice contains various substances that inhibit CYP3A4-mediated metabolism only in the wall of the gut, mainly by selective down-regulation of CYP3A4 protein in the small intestine.<sup>23</sup> The area under the curve for plasma saquinavir is increased by 50 to 150 percent during concomitant administration of grapefruit juice.<sup>11</sup> However, grapefruit juice should not be relied on to increase plasma concentrations of protease inhibitors, because the variations in the amounts of flavonoids and other potentially active substances among products can result in inconsistent effects.24

The enterocytes in the intestinal mucosa are also

a major site of expression of P-glycoprotein, one of several membrane-bound proteins that increase the efflux of drugs from cells.25 Several protease inhibitors are substrates for and inhibitors of P-glycoprotein<sup>26,27</sup>; ritonavir is the most potent inhibitor.28,29 Both cytochrome P-450 enzymes and P-glycoprotein can present a barrier to the absorption of orally administered drugs and have a considerable effect on drug interactions. Figure 2B shows the effect of rifampin on plasma digoxin concentrations through the induction of intestinal P-glycoprotein.7 Although the inhibition and induction of intestinal CYP3A enzymes from metabolic processes result in direct changes in drug absorption, the inhibition and induction of P-glycoprotein primarily affect the rate of drug absorption.<sup>30</sup> The overlap of tissue distribution and substrate specificity of CYP3A4 and P-glycoprotein in the gut wall makes it difficult to define the specific mechanisms of some



Figure 1. Various Sites in the Body in Which Drug Interactions Occur.

The inset shows a T lymphocyte in which nucleoside-analogue reverse-transcriptase inhibitors are undergoing intracellular conversion to their active forms. AZT denotes zidovudine, MP monophosphate, DP diphosphate, and TP triphosphate.

SELECTED DRU	JGS USED FOR TH	HE TREATMENT			
OF	OF HIV INFECTION.*				
CYP3A4 substrates	CYP3A4 inhibitors	CYP3A4 inducers			
Astemizole Clarithromycin Cyclosporine Dapsone Efavirenz Erythromycin Estrogens Etoposide Fentanyl Midazolam Nefazodone Prednisone Protease inhibitors Sertraline Testosterone Triazolam	Amprenavir Clarithromycin Delavirdine Efavirenz Erythromycin Fluconazole Fluoxetine Grapefruit juice Indinavir Itraconazole Ketoconazole Lopinavir Nelfinavir Ritonavir Saquinavir	Carbamazepine Efavirenz Nevirapine Phenytoin Phenobarbital Rifampin Rifabutin Rifabutin Ritonavir Troglitazone			
CYP2D6 substrates Codeine Desipramine Fluoxetine Haloperidol Methadone Morphine Paroxetine Risperidone	CYP2D6 inhibitors Fluoxetine Paroxetine Quinidine Ritonavir Sertraline				
<b>CYP2C19</b> substrates Nelfinavir Omeprazole Diazepam	CYP2C19 inhibitors Fluconazole Omeprazole Fluoxetine	CYP2C19 inducers Rifampin			
CYP1A2 substrates Haloperidol Theophylline Zileuton	CYP1A2 inhibitors Ciprofloxacin Clarithromycin Erythromycin Fluvoxamine	CYP1A2 inducers Carbamazepine Phenytoin Phenobarbital Ritonavir			

 TABLE 2. CYTOCHROME P-450 ISOFORMS AND

\*Data are from Flockhart<sup>17</sup> and Michalets.<sup>18</sup>

drug interactions and predict the plasma concentrations of certain drug combinations. Morover, the involvement of CYP3A4 and P-glycoprotein in drug interactions is not always complementary. For example, plasma indinavir concentrations either do not change or are decreased by the ingestion of grapefruit juice, suggesting that the activation of P-glycoprotein may compensate for the inhibition of CYP3A4,6,31,32 but P-glycoprotein has little effect on the absorption of saquinavir.<sup>33</sup> The specific contribution of CYP3A4 inhibition and P-glycoprotein activation to interactions within the gastrointestinal tract remains unclear because most drugs that modulate P-glycoprotein also inhibit CYP3A4.34 In any case, the inhibition of CYP3A4, P-glycoprotein, or both in the gut wall may have a substantial effect on plasma concentrations of many anti-HIV drugs.

P-glycoprotein is present at numerous other sites in the body.<sup>35</sup> Its presence in the renal tubular cells and hepatocytes results in increased drug excretion in urine and bile. P-glycoprotein in the endothelial cells of the blood-brain barrier prevents the entry of certain drugs into the central nervous system.<sup>36</sup> Ketoconazole, an inhibitor of both CYP3A4 and P-glycoprotein, causes a larger increase in cerebrospinal fluid concentrations of saquinavir and ritonavir than in unbound plasma concentrations, suggesting that the inhibition of efflux transporters can be used to target therapy in the central nervous system.

#### **Drug Absorption**

Interactions that alter the absorption of drugs often lead to dramatic changes in plasma drug concentrations. The concomitant administration of a fluoroquinolone with divalent and trivalent cations can reduce the area under the curve for plasma quinolone by more than 90 percent.<sup>4</sup> A didanosine formulation containing an aluminum–magnesium antacid buffer decreases the area under the curve for plasma ciprofloxacin by 80 percent (Fig. 2C).<sup>5</sup> These interactions can easily be avoided by administering the fluoroquinolone at least two hours before or six hours after the antacid, or by using the new, enteric-coated formulation of didanosine.

The absorption of other drugs may be altered by changes in gastric pH. For example, because ketoconazole is best absorbed when the gastric pH is low, concomitant administration of ketoconazole and  $H_2$ -antagonists, antacids, or proton-pump inhibitors results in marked impairment of the absorption of ketoconazole.<sup>37</sup> Itraconazole is also best absorbed when the gastric pH is low, but its administration with food is more important for achieving high plasma concentrations.<sup>38</sup> The absorption of fluconazole is unaffected by variations in gastric pH.<sup>39</sup>

#### **Renal Elimination**

Probenecid and trimethoprim are competitive inhibitors of renal tubular secretion of other drugs that are primarily eliminated through this pathway. Although probenecid increases plasma acyclovir concentrations and trimethoprim–sulfamethoxazole increases plasma lamivudine concentrations (Fig. 2D), these interactions are not clinically important and do not warrant a change in dose, because high concentrations of these drugs are not associated with adverse effects.<sup>13,40</sup> Probenecid also inhibits hepatic glucuronidation of zidovudine and increases plasma zidovudine concentrations by 80 percent.<sup>41</sup> The clinical importance of this increase is unknown.

# PREDICTING DRUG INTERACTIONS

The multiple metabolic pathways of some drugs make it difficult to predict the outcome of drug interactions. Although in vitro systems have been de-

#### Figure 2. Mechanisms of Drug Interactions.

Inhibition of intestinal and hepatic CYP3A4 by ritonavir markedly increases plasma saquinavir concentrations when saquinavir is given at a dose of 400 mg and ritonavir at a dose of 600 mg (Panel A; bars denote standard errors).<sup>20</sup> Induction of intestinal P-gly-coprotein by rifampin decreases plasma digoxin concentrations (Panel B; I bars denote standard deviations).<sup>7</sup> The combination of ciprofloxacin and didanosine results in decreased plasma ciprofloxacin concentrations, because absorption of the drug is decreased by chelation with divalent cations in the antacid buffer of didanosine (Panel C; bars denote standard deviations).<sup>5</sup> When lamivudine is combined with trimethoprim–sulfamethoxazole (TMP-SMX), plasma lamivudine c0z pamUzkn-Z08pc-Z0zFpentr-8:zFpati-0 z8pons -Z:Fz8apr-8/z1ps -Z

veloped to test the effects of certain drugs on the metabolism of other drugs, these systems may not accurately predict the effect in patients receiving drugs with complex metabolism, and induction interactions may not be detected if the system can assess only inhibition.<sup>42</sup> For example, initial in vitro data from human liver microsomes suggested that plasma methadone concentrations would be increased by the inhibitory effects of ritonavir on CYP3A4.<sup>43</sup> In a clinical study, however, plasma methadone concentrations were decreased by the administration of ritonavir.<sup>44</sup> Subsequently, ritonavir was found to displace methadone from plasma protein–binding sites and to increase its metabolism by inducing CYP2B6, which degrades methadone.<sup>45,46</sup> Studies in hepatocytes revealed no effect of rifampin on glucuronidation of zidovudine, but rifampin-induced glucuronidation of zidovudine was demonstrated in vivo.<sup>12,47,48</sup>

Even clinical studies may not accurately predict changes if the interaction is dependent on time. Ritonavir inhibits the metabolism of alprazolam during a short-term regimen of ritonavir but induces the metabolism of alprazolam when it is given for 10 days.<sup>49</sup> Furthermore, most in vivo and in vitro studies of drug interactions evaluate two-drug regimens, and the results may not apply to the multi-drug regimens often used clinically. This is especially true for a regimen consisting of three or more drugs with opposing effects on CYP3A4 metabolism. The lack of multidrug interaction studies provides little assistance to the clinician, who is left to rely on adverse effects or treatment failure to demonstrate whether an interaction occurred.

# NUCLEOSIDE-ANALOGUE REVERSE-TRANSCRIPTASE INHIBITORS

Because nucleoside-analogue reverse-transcriptase inhibitors are primarily eliminated by the kidneys, they do not interact with other drugs through the cytochrome P-450 system. These drugs can be given with protease inhibitors and non-nucleoside reverse-transcriptase inhibitors without dosage adjustments.

Nucleoside reverse-transcriptase inhibitors are prodrugs that require intracellular phosphorylation to the active moiety, and they may therefore interact with drugs that compete for the intracellular activation pathway. Ribavirin decreases the phosphorylation of zidovudine and stavudine in vitro, resulting in decreased concentrations of the active compound.<sup>50,51</sup> Patients who have HIV infection and hepatitis C may be treated with regimens that contain ribavirin, which may reduce the efficacy of zidovudine. Similarly, zidovudine may impair the intracellular phosphorylation of stavudine,<sup>1</sup> and this combination is associated with a less favorable outcome than other regimens containing two nucleoside reverse-transcriptase inhibitors.<sup>2</sup> Also, lamivudine inhibits phosphorylation of zalcitabine.3

Other intracellular interactions may increase the activity of nucleoside reverse-transcriptase inhibitors. Hydroxyurea, an inhibitor of the enzyme ribonucleotide reductase, which is involved in the formation of deoxynucleotides, increases the antiviral action of didanosine.52 One possible mechanism for this effect involves a decrease in the intracellular pool of 2'-deoxvadenosine-5'-triphosphate (dATP), which competes with 2',3'-dideoxyadenosine-5'-triphosphate (ddATP), the active metabolite of didanosine, for incorporation into viral DNA. As a result, the intracellular ratio of ddATP to dATP is increased, improving the antiviral potency of didanosine. However, the long-term clinical benefits of hydroxyurea-containing combinations are unclear, because hydroxyurea blunts the increase in CD4 cells that occurs in response to antiretroviral therapy and has numerous adverse effects, including hepatitis, pancreatitis, and bone marrow toxicity.53

# NON-NUCLEOSIDE REVERSE-TRANSCRIPTASE INHIBITORS

The three non-nucleoside reverse-transcriptase inhibitors that have been approved by the FDA can inhibit or induce cytochrome P-450 activity, depending on the specific drug. Nevirapine and efavirenz are moderate inducers of CYP3A4. Nevirapine decreases plasma concentrations of indinavir and saquinavir (Table 3) but does not have clinically important effects on nelfinavir and ritonavir, because these drugs are not exclusively metabolized by CYP3A4, and they induce their own metabolism, minimizing the effects of further induction.<sup>58,60,63</sup> Efavirenz inhibits or induces cytochrome P-450 activity, depending on the concomitantly administered drug. Efavirenz decreases plasma concentrations of indinavir, lopinavir, saquinavir, and amprenavir<sup>55,57,59,64</sup> but increases plasma concentrations of ritonavir and nelfinavir by approximately 20 percent, possibly through inhibition of the CYP2C9 or CYP2C19 pathway.<sup>65,66</sup> Since efavirenz causes large decreases in plasma saquinavir concentrations, this combination should be avoided, unless given concomitantly with ritonavir.<sup>67</sup>

Of particular concern is the effect of nevirapine or efavirenz on plasma methadone concentrations. Both drugs can reduce plasma methadone concentrations by about 50 percent in patients receiving methadone maintenance therapy,<sup>68,69</sup> and many patients have symptoms consistent with methadone withdrawal, requiring an increase in the dose of methadone. Patients receiving methadone should be monitored closely when given efavirenz or nevirapine, with the expectation that the methadone dose will need to be increased.

Delavirdine is a potent inhibitor of cytochrome P-450. Because of its effect on CYP3A4, serious toxic effects may occur if delavirdine is administered with antiarrhythmic drugs, calcium-channel blockers, sedative or hypnotic drugs, or quinidine.<sup>70</sup> The administration of delavirdine with vasoconstrictor drugs such as ergotamine can lead to peripheral ischemia and can increase the toxicity of certain chemotherapeutic drugs, such as etoposide and paclitaxel. Clinicians need to be aware of and avoid these combinations.

# **HIV-PROTEASE INHIBITORS**

HIV-protease inhibitors are associated with numerous drug interactions, many of which are clinically important (Table 3). These drugs are inhibitors of CYP3A4 enzymes and are contraindicated in combination with certain antiarrhythmic drugs, sedative and hypnotic drugs, ergot derivatives, cisapride, and the 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors lovastatin and simvastatin. Ritonavir is the most potent inhibitor of cytochrome activity and is therefore most likely to interact with other drugs. Indinavir, amprenavir, and nelfinavir have a moderate probability of causing interactions, and saquinavir has the lowest probability. The newer, soft-gel formulation of saquinavir is similar to the original formulation in this respect.9 The combination of lopinavir and ritonavir is likely to have interactions that are similar to those of full-dose ritonavir alone, but the magnitude of the interactions may be smaller.57

In addition to inhibiting enzymes, ritonavir has enzyme-inducing properties, even inducing its own metabolism in a dose-dependent manner during the

Drug	Interacting Drug	Result*	RECOMMENDATION	
Amprenavir	Rifampin	Amprenavir AUC decreased by 81%54		
Indinavir	Rifampin	Indinavir AUC decreased by 92%54	Avoid combinations of rifampin and protease inhibitor except possibly for	
Ritonavir	Rifampin	Ritonavir AUC decreased by 35% <sup>54</sup>		
Saquinavir (hard or soft gel capsules)	Rifampin	Saquinavir AUC decreased by 70-80%9,54	at adjusted dose with nelfinavir, am-	
Nelfinavir	Rifampin	Nelfinavir AUC decreased by 82% <sup>54</sup>	1 , ,	
Amprenavir	Efavirenz	Amprenavir AUC decreased by 36% <sup>55</sup>	Increase amprenavir dose to 1200 mg 3 times a day or add ritonavir (200 mg twice a day)	
HMG CoA reductase inhibitors				
Simvastatin	Ritonavir with saquinavir (soft gel capsules)	Simvastatin AUC increased by a factor of 3256	Do not use simvastatin with ritonavir	
Atorvastatin	Ritonavir with saquinavir (soft gel capsules)	Atorvastatin AUC increased by a factor of 4.5 <sup>56</sup>	Use atorvastatin with slow dose titra- tion and close monitoring	
Pravastatin	Ritonavir with saquinavir (soft gel capsules)	Pravastatin AUC decreased by a factor of 0.5 <sup>56</sup>	Adjustment of pravastatin dose not re- quired	
Atorvastatin	Lopinavir-ritonavir	Atorvastatin AUC increased by a factor of 5.9 <sup>57</sup>	Use atorvastatin with slow dose titra- tion and close monitoring	
Pravastatin	Lopinavir-ritonavir	Pravastatin AUC increased by 30%57	Pravastatin does not require dose ad- justment	
Indinavir	Nevirapine	Indinavir AUC decreased by 28%58	Increase indinavir dose to 1000 mg ev- ery 8 hours	
Indinavir	Efavirenz	Indinavir AUC decreased by 35% <sup>59</sup>	Increase indinavir dose to 1000 mg ev- ery 8 hours	
Lopinavir-ritonavir	Efavirenz	Lopinavir trough concentration decreased by $40\%^{57}$	Consider increasing lopinavir dose to 533 mg and ritonavir dose to 133 mg	
Rifabutin	Amprenavir Indinavir	Rifabutin increased by a factor of 2 to $3^{54}$	Decrease rifabutin dose to 150 mg/day Decrease rifabutin dose to 150 mg/day, increase indinavir dose to 1000 mg 3 times a day	
	Nelfinavir		2	
Rifabutin	Ritonavir	Rifabutin AUC increased by a factor of $4^{54}$	Decrease rifabutin dose to 150 mg ev- ery 2 or 3 days or 2 or 3 times a week	
Rifabutin	Lopinavir-ritonavir	Rifabutin AUC increased by a factor of $3^{57}$	Decrease rifabutin dose to 150 mg ev- ery 2 or 3 days or 2 or 3 times a week	
Saquinavir	Nevirapine	Saquinavir AUC decreased by 62%60	Avoid combination unless ritonavir is used concomitantly	
Saquinavir (hard or soft gel capsules)	Rifabutin	Saquinavir AUC decreased by 45% (hard gel capsules) or 47% (soft gel capsules) <sup>9,54</sup>	Avoid combination unless ritonavir is administered concomitantly	
Sildenafil	Indinavir, saquinavir, or ritonavir	Sildenafil AUC increased by a factor of 2 with indinavir, a factor of 3 with saquinavir, and a factor of 11 with ritonavir <sup>61,62</sup>	Start with 25 mg of sildenafil; with ri- tonavir, do not repeat the sildenafil dose for 48 hours	

#### TABLE 3. SELECTED INTERACTIONS AMONG ANTIRETROVIRAL DRUGS AND BETWEEN ANTIRETROVIRAL AND OTHER DRUGS.\*

\*AUC denotes area under the concentration-time curve, and HMG CoA 3-hydroxy-3-methylglutaryl coenzyme A.

first 14 days of therapy.<sup>71</sup> Ritonavir decreases plasma concentrations of theophylline, probably through the induction of CYP1A2.<sup>72</sup> Ritonavir and nelfinavir also increase glucuronosyltransferase activity, which may partly explain the substantial decreases in plasma ethinyl estradiol concentrations during concurrent therapy with these protease inhibitors.<sup>73,74</sup> Alternative or additional methods of contraception are recommended in women taking ritonavir or nelfinavir.

Not only do HIV-protease inhibitors affect the metabolism of certain drugs, but their own metabolism is also altered by other inducers or inhibitors of cytochrome activity. Potent enzyme-inducing drugs can cause clinically important decreases in plasma concentrations of protease inhibitors. For example, rifampin decreases plasma saquinavir concentrations by 70 to 80 percent.<sup>9,54</sup> The resulting low plasma concentrations may promote viral resistance and result in treatment failure. With the possible exception of ritonavir, protease inhibitors should not be given to patients receiving rifampin.<sup>75</sup> Patients with tuberculosis who are already receiving a protease inhibitor should be treated with a four-drug regimen that includes rifabutin instead of rifampin.<sup>54</sup> For patients receiving indinavir, nelfinavir, or amprenavir, the dose of rifabutin should be reduced from 300 to 150 mg per day to compensate for the inhibition of rifabutin clearance by these drugs. Increasing the dose of indinavir from 800 to 1000 mg every eight hours, in addition to reducing the dose of rifabutin, is also an option.<sup>76</sup> Ritonavir increases the area under the curve for plasma rifabutin by a factor of four, which may lead to clinically important adverse effects.<sup>77,78</sup> However, intermittent administration of rifabutin, either 150 mg every three days or 300 mg every seven days, is safe and tolerable over a two-month period with a combination of ritonavir and saquinavir (400 mg of each drug every 12 hours).<sup>79</sup> Updated guidelines for using rifabutin and rifampin in patients receiving antiretroviral drugs have recently been issued by the Centers for Disease Control and Prevention.<sup>79</sup>

Other potent enzyme inducers, such as phenytoin, phenobarbital, and carbamazepine, can cause similar reductions in plasma concentrations of protease inhibitors.<sup>80</sup> Although standard doses of carbamazepine and phenobarbital may have to be decreased in the presence of protease inhibitors, standard doses of phenytoin may have to be increased in the presence of nelfinavir or ritonavir.81 For example, plasma phenytoin concentrations were decreased by nelfinavir, perhaps through the induction of its CYP2C9-mediated metabolism.82 Ritonavir may either inhibit or induce the metabolism of phenytoin, as it does with alprazolam, depending on the duration of ritonavir therapy.49 Interactions between anticonvulsant drugs and protease inhibitors are complex because of their twoway nature. It is best to avoid these combinations, and close monitoring is required when they must be used.

Patients who are taking protease inhibitors and who require prophylaxis against *Mycobacterium avium* complex infection can be given azithromycin or clarithromycin. The area under the curve for plasma clarithromycin is moderately increased by ritonavir and indinavir, but dosage adjustments are not necessary in patients with normal renal function.<sup>83,84</sup> Azithromycin is excreted primarily by the biliary route and does not interact with protease inhibitors or delavirdine.<sup>85</sup>

The concept of using drug interactions to the patient's benefit has been the focus of much research. The administration of cytochrome P-450 inhibitors with other drugs can reduce the pill burden, increase plasma concentrations, simplify the dosing schedule, and circumvent drug interactions. Table 4 lists combinations that improve the pharmacokinetic profile of protease inhibitors. The bioavailability of saquinavir is less than 20 percent, and up to 18 capsules per day must be given to achieve effective plasma concentrations. When ritonavir is given with saquinavir, however, steady-state plasma concentrations of saquinavir increase by a factor of 20 or more, dramatically improving the oral bioavailability of the drug.<sup>22</sup> With ritonavir 400 mg twice daily, the dose of saquinavir can be reduced from 1200 mg every eight hours to 400 mg twice daily, decreasing the number of saquinavir capsules that must be taken from 18 to 4 per day. Similarly, nelfinavir raises plasma saquinavir concentrations by a factor of about 12, allowing the dose of saquinavir to be reduced to 1000 mg twice daily.87

The combination of ritonavir and indinavir can overcome the unfavorable pharmacokinetic properties of indinavir. Indinavir must be taken every eight hours on an empty stomach or with a meal that is low in fat ( $\leq 2$  g). The trough plasma concentrations of indinavir are highly variable, and in some patients the concentrations at the end of the dosing interval may be below the in vitro concentration needed to inhibit the replication of 90 percent of HIV isolates (IC<sub>90</sub>). Concomitant administration of ritonavir increases the area under the curve by a factor of up to three and increases the trough plasma concentration by a factor of three to seven.<sup>10,86</sup> This allows for a decrease in dosing from three times daily without food to twice daily with food. Twice-daily regimens of indinavir and ritonavir (800 mg of indinavir and 100 mg of ritonavir, 800 mg of indinavir and 200 mg of ritonavir, and 400 mg of each) are being evaluated in patients with HIV infection.88,89 All three regimens result in higher plasma indinavir concentrations, although the lower doses of ritonavir may be better tolerated. In one study, the incidence of renal complications was low among patients who were given 400 mg of each drug twice a day, and there were no cases of nephrolithiasis among the 89 patients who received this regimen for a mean period of 40 weeks.89

Similarly, the reduction in plasma amprenavir or saquinavir concentrations produced by efavirenz can be circumvented by the addition of ritonavir. Efavirenz markedly decreases the area under the curve for amprenavir and saquinavir.<sup>55,64</sup> The addition of ritonavir (200 mg twice daily) to amprenavir and efavirenz not only prevents the efavirenz-induced reduction, but also increases the area under the curve for plasma amprenavir by a factor of two and increases the trough plasma concentration by a factor of four.<sup>90</sup> Plasma saquinavir concentrations are not affected by efavirenz in patients who are also taking 400 mg of ritonavir twice daily.<sup>91</sup>

Lopinavir, a recently approved protease inhibitor, relies on the inhibitory effects of ritonavir to achieve plasma concentrations well above the  $IC_{90}$  value for wild-type HIV. Low doses of ritonavir increase the area under the curve for plasma lopinavir by a factor of 20.<sup>92</sup> The trough plasma concentration of tipranavir, an investigational protease inhibitor, is increased in a dose-dependent fashion by ritonavir.<sup>93</sup>

Delavirdine is also an inhibitor of CYP3A4 and can be given to increase plasma concentrations of protease inhibitors.<sup>94,95</sup> Recent improvement in the formulation (a 200-mg tablet) and studies of twice-daily dosing make it a possible alternative to ritonavir as a means of increasing the concentrations of protease inhibitors.

#### DRUGS FOR OPPORTUNISTIC INFECTIONS

Azole antifungal drugs, macrolide antibiotics, and rifamycins have important interactions with other

AFFECTED DRUG	Interacting Drug	Results*	RECOMMENDATION
Amprenavir	Lopinavir-ritonavir	Amprenavir AUC increased <sup>57</sup>	Consider giving amprenavir at a dose of 750 mg twice a day
Indinavir	Ritonavir	Indinavir AUC increased by a factor of up to 3 and trough concentration in- creased by a factor of 3 to 7 <sup>10,86</sup>	Regimens under evaluation: 800 mg of indinavir and 100 mg of ritonavir twice a day, 800 and 200 mg twice a day, and 400 and 400 mg twice a day
Indinavir	Delavirdine	Indinavir AUC increased by a factor of <b>3</b> 94	Consider decreasing indinavir dose to 600 mg three times a day
Indinavir	Lopinavir-ritonavir	Increased indinavir AUC <sup>57</sup>	Consider giving indinavir at a dose of 600 mg twice a day
Saquinavir	Lopinavir-ritonavir	Increased saquinavir AUC <sup>57</sup>	Consider giving saquinavir at a dose of 800 mg twice a day
Saquinavir	Nelfinavir	Saquinavir AUC increased by a factor of 5 (soft gel cap- sules) or 12 (hard gel cap- sules) <sup>87</sup>	Decrease saquinavir dose to 800 mg three times a day or 1000 mg twice a day
Saquinavir	Ritonavir	Saquinavir C <sub>ss</sub> increased by a factor of 20 or more <sup>22</sup>	Give both drugs at a dose of 400 mg twice a day; regi- mens under evaluation: 200 mg of ritonavir and 800 mg of saquinavir twice a day, 100 and 1000 mg twice a day
Saquinavir	Delavirdine	Saquinavir AUC increased by a factor of 595	Consider decreasing the saquinavir dose to 800 mg three times a day

**TABLE 4.** COMBINATIONS OF DRUGS THAT CAN BE GIVEN TO ACHIEVE OPTIMAL PLASMA CONCENTRATIONS OF ANTIRETROVIRAL DRUGS.

\*AUC denotes area under the concentration–time curve, and  $\mathrm{C}_{\mathrm{ss}}$  plasma concentration in steady state.

drugs, complicating the prophylaxis and treatment of opportunistic infections (Table 3). Ketoconazole and itraconazole, which are potent inhibitors of CYP3A4 and moderate inhibitors of P-glycoprotein, can be given to increase plasma concentrations of protease inhibitors.<sup>34</sup> For example, ketoconazole increases the area under the curve for plasma saquinavir by 150 percent.96 However, this combination is rarely given because of concern about the toxicity of ketoconazole and the development of resistant fungal infections. Fluconazole can also inhibit CYP3A4, although the extent of inhibition and the magnitude of interactions are dose-dependent. Doses of less than 200 mg per day are not associated with important interactions, whereas higher doses can increase plasma concentrations of substrates of CYP3A4,97 as well as glucuronidated drugs such as zidovudine.98 Fluconazole also inhibits CYP2C9, as evidenced by its potential to increase the risk of bleeding in patients also taking warfarin.99

Erythromycin and clarithromycin are substrates of CYP3A4 and inhibitors of both CYP3A4 and P-glycoprotein.<sup>34</sup> Rifampin and rifabutin can substantially decrease plasma clarithromycin concentrations, but the clinical importance of the decrease is unknown because intracellular concentrations of macrolides are much higher than plasma concentrations.<sup>100,101</sup> Decreased effectiveness against pathogens such as *M. avium* complex is a potential concern, but no studies have specifically addressed the clinical effect of the combination.

Fluconazole increases the area under the curve for plasma rifabutin by 75 percent.<sup>102</sup> This combination of drugs is more effective in preventing *M. avium* complex bacteremia than rifabutin alone, although there is an increased risk of rifabutin-associated toxic effects, such as uveitis and arthralgias.<sup>103</sup>

#### SPECIAL ISSUES

#### **Alternative Therapies**

Alternative therapies, including herbal remedies and nutritional supplements, have long been considered harmless. However, certain alternative therapies may interact with drugs used in the treatment of HIV infection. St. John's wort decreases the area under the curve for plasma indinavir by more than 50 percent in normal subjects, an effect that is due to the induction of CYP3A4 or P-glycoprotein.<sup>104-106</sup> Use of this herb should be avoided in patients taking protease inhibitors and non-nucleoside reverse-transcriptase inhibitors, because of the risk of viral resistance to these drugs.

Raw garlic and garlic supplements inhibit the activity of CYP3A4 in vitro and in animals.<sup>107,108</sup> Severe gastrointestinal toxicity was reported in two persons after they ingested garlic supplements with ritonavir.<sup>109</sup> Other herbs with reported in vitro effects on cytochrome P-450–mediated drug metabolism include silymarin (milk thistle), ginseng, and skullcap.<sup>110</sup> Clinicians should be aware of these potential interactions, because alternative therapies are not usually evaluated as a cause of treatment failure or toxicity.

# **Drug-Cytokine Interactions**

Cytochrome P-450 drug metabolism can be altered by certain proinflammatory cytokines such as interleukin-6, interleukin-1, and tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ).<sup>111</sup> These cytokines are released during periods of stress, trauma, or infection. In several in vitro studies, interleukin-6 and TNF- $\alpha$  inhibited cytochrome P-450–mediated metabolism through a metabolic interaction at the level of transcription of cytochrome mRNA.<sup>112</sup>

The administration of immunomodulators such as interleukin-2 results in a profound release of these cytokines.<sup>113,114</sup> In a study of HIV-infected patients who were receiving a five-day infusion of interleukin-2, the area under the curve for plasma indinavir was increased by 75 percent.<sup>115</sup>

# ROLE OF THERAPEUTIC DRUG MONITORING

Several studies have established associations between plasma concentrations of protease inhibitors and their antiviral effects,<sup>116-118</sup> suggesting a role for therapeutic monitoring of these drugs. Although there is considerable debate regarding the value of drug monitoring,119-121 determination of plasma drug concentrations may have a role in the evaluation of drug interactions, provided that the limitations in the use of plasma drug measurements to evaluate individual patients are recognized. These limitations include the large variability in pharmacokinetic characteristics within individual patients, lack of information on specific therapeutic ranges and target concentrations (i.e., data on the concentrations that cause 50 percent inhibition), variations in drug binding to  $\alpha$ 1-acid glycoprotein and albumin, slow viral responses to changes in plasma drug concentrations, and clinical interpretations of measurements.

The clinical utility of therapeutic monitoring of antiretroviral drugs has yet to be proved, but trials are ongoing. Adjustments of doses on the basis of plasma drug measurements in cases of drug interactions should be made with caution pending the outcome of trials examining the correlations between such measurements and virologic and clinical end points. Any decision to adjust a dose, whether because of low plasma drug concentrations or drug toxicity, must take into consideration the wide variability in plasma drug concentrations in an individual patient, both on a single day and from one day to the next due to diurnal and food effects, the stage of the disease, and changes in adherence to the treatment regimen.

## MANAGEMENT OF DRUG INTERACTIONS

New information about drug interactions in patients with HIV infection becomes available almost weekly. The increasing number of documented and theoretical drug interactions can be overwhelming for the practicing clinician. Fortunately, extensive tables and product information are available to aid in the recognition and management of drug interactions (Table 5). A thorough drug history, including overthe-counter drugs and alternative therapies, should be obtained at each clinic visit. Clinicians should have a high index of suspicion for a drug interaction in patients receiving antiretroviral therapy who have an increased viral load or clinical progression, if other factors, including adherence, can be ruled out. Interactions should also be suspected in patients with serious toxic effects of antiretroviral or supportive drugs. Regimens containing many drugs and drugs with a high potential for interactions (rifamycin, protease inhibitors, and antifungal drugs) should be reviewed and assessed for drug interactions. The selection of a drug that is less likely to interact with other drugs should be considered if warranted by the clinical circumstances. For example, azithromycin is not metabolized by cytochrome P-450 and does not have the interactions associated with other macrolide antibiotics. Similarly, fluconazole at low doses is less likely to interact with other drugs than are ketoconazole

 
 TABLE 5. Web Sites with Information About Drug Interactions.

www.dml.georgetown.edu/depts/pharmacology (Department of Pharmacology, Georgetown University Medical Center) www.foodmedinteractions.com (food and drug interactions) www.hivatis.org (HIV/AIDS Treatment Information Service) www.hivdent.org (dental information) hivinsite.ucsf.edu www.hiv.net (in German) www.hopkins-aids.edu (Johns Hopkins AIDS Service) www.iapac.org (International Association of Physicians in AIDS Care) www.hiv-druginteractions.org (Liverpool HIV Pharmacology Group) www.medscape.com and itraconazole. Therapeutic use of pharmacokinetic interactions should be considered to simplify complex regimens and reduce the pill burden.

With so many new drugs in clinical development, drug interactions will continue to be an important aspect of the treatment of patients with HIV infection. It is essential that clinicians understand the main mechanisms and concepts underlying these interactions, so that they can choose regimens for their patients that are potent, safe, and convenient.

Dr. Gallicano has received research grants from Hoffmann-LaRoche Canada and Merck.

#### REFERENCES

**1.** Hoggard PG, Kewn S, Barry MG, Khoo SH, Back DJ. Effects of drugs on 2',3'-dideoxy-2',3'-didehydrothymidine phosphorylation in vitro. Antimicrob Agents Chemother 1997;41:1231-6.

**2.** Havlir DV, Friedland G, Pollard R, et al. Combination zidovudine (ZDV) and stavudine (d4T) therapy versus other nucleosides: report of two randomized trials (ACTG 290 and 298). In: Program and abstracts of the 5th Conference on Retroviruses and Opportunistic Infections, Chicago, February 1–5, 1998:79. abstract.

**3.** Veal GJ, Hoggard PG, Barry MG, Khoo S, Back DJ. Interaction between lamivudine (3TC) and other nucleoside analogues for intracellular phosphorylation. AIDS 1996;10:546-8.

**4.** Lehto P, Kivisto KT. Effect of sucralfate on absorption of norfloxacin and ofloxacin. Antimicrob Agents Chemother 1994;38:248-51.

**5.** Sahai J, Gallicano K, Oliveras L, Khaliq S, Hawley-Foss S, Garber G. Cations in the didanosine tablet reduce ciprofloxacin bioavailability. Clin Pharmacol Ther 1993;53:292-7.

6. Crixivan (indinavir). West Point, Pa.: Merck, 1998 (package insert).

**7.** Greiner B, Eichelbaum M, Fritz P, et al. The role of intestinal P-glycoprotein in the interaction of digoxin and rifampin. J Clin Invest 1999;104: 147-53.

**8.** Khaliq Y, Gallicano K, Venance S, Kravcik S, Cameron DW. Effect of ketoconazole on ritonavir and saquinavir concentrations in plasma and cerebrospinal fluid from patients infected with human immunodeficiency virus. Clin Pharmacol Ther 2000;68:637-46.

**9.** Jorga K, Buss NE. Pharmacokinetic (PK) drug interaction with saquinavir soft gelatin capsule: In: Program and abstracts of the **39**th In-

terscience Conference on Antimicrobial Agents and Chemotherapy, San Francisco, September 26–29, 1999. Washington, D.C.: American Society for Microbiology, 1999:20. abstract.

**10.** Hsu A, Granneman GR, Cao G, et al. Pharmacokinetic interaction between ritonavir and indinavir in healthy volunteers. Antimicrob Agents Chemother 1998;42:2784-91.

**11.** Kupferschmidt HH, Fattinger KE, Ha HR, Follath F, Krahenbuhl S. Grapefruit juice enhances the bioavailability of the HIV protease inhibitor saquinavir in man. Br J Clin Pharmacol 1998;45:355-9.

**12.** Gallicano KD, Sahai J, Shukla VK, et al. Induction of zidovudine glucuronidation and amination pathways by rifampicin in HIV-infected patients. Br J Clin Pharmacol 1999;48:168-79.

**13.** Moore KHP, Yuen GJ, Raasch RH, et al. Pharmacokinetics of lamivudine administered alone and with trimethoprim-sulfamethoxazole. Clin Pharmacol Ther 1996;59:550-8.

**14.** Merrill DP, Manion DJ, Chou TC, Hirsch MS. Antagonism between human immunodeficiency virus type 1 protease inhibitors indinavir and saquinavir in vitro. J Infect Dis 1997;176:265-8.

**15.** Barry M, Mulcahy F, Merry C, Gibbons S, Back D. Pharmacokinetics and potential interactions amongst antiretroviral agents used to treat patients with HIV infection. Clin Pharmacokinet 1999;36:289-304.

**16.** Nelson DR, Kamataki T, Waxman DJ, et al. The P450 superfamily: update on new sequences, gene mapping, accession numbers, early trivial names of enzymes, and nomenclature. DNA Cell Biol 1993;12:1-51.

**17.** Flockhart D. Cytochrome P450 drug interaction table. Washington, D.C.: Georgetown University Medical Center, May 2000. (See http://www.dml.georgetown.edu/depts/pharmacology/davetab.html.) (See

NAPS document no. 05584 for 6 pages, c/o Microfiche Publications, 248 Hempstead Tpke., West Hempstead, NY 11552.) **18.** Michalets EL. Update: clinically significant cytochrome P-450 drug

**18.** Michalets EL. Update: clinically significant cytochrome P-450 drug interactions. Pharmacotherapy 1998;18:84-112.

**19.** Koudriakova T, Latsimirskaia E, Utkin I, et al. Metabolism of the human immunodeficiency virus protease inhibitors indinavir and ritonavir by human intestinal microsomes and expressed cytochrome P4503A4/3A5: mechanism-based inactivation of cytochrome P4503A by ritonavir. Drug Metab Dispos 1998;26:552-61.

**20.** Hsu A, Granneman GR, Cao G, et al. Pharmacokinetic interactions between two human immunodeficiency virus protease inhibitors, ritonavir and saquinavir. Clin Pharmacol Ther 1998;63:453-64.

**21.** Condra JH, Petropoulos CJ, Ziermann R, Schleif WA, Shivaprakash M, Emini EA. Drug resistance and predicted virologic responses to human immunodeficiency virus type 1 protease inhibitor therapy. J Infect Dis 2000;182:758-65.

**22.** Kolars JC, Lown KS, Schmiedlin-Ren P, et al. CYP3A gene expression in human gut epithelium. Pharmacogenetics 1994;4:247-59.

23. Fuhr U. Drug interactions with grapefruit juice: extent, probable

mechanism and clinical relevance. Drug Saf 1998;18:251-72.

**24.** Bailey DG, Malcolm J, Arnold MJ, Spence JD. Grapefruit juice-drug interactions. Br J Clin Pharmacol 1998;46:101-10.

**25.** Fojo AT, Ućda K, Slamon DJ, Poplack DG, Gottesman MM, Pastan I. Expression of a multidrug-resistance gene in human tumors and tissues. Proc Natl Acad Sci U S A 1987;84:265-9.

**26.** Washington CB, Duran GE, Man MC, Sikic BI, Blaschke TF. Interaction of anti-HIV protease inhibitors with the multidrug transporter P-gly-coprotein (P-gp) in human cultured cells. J Acquir Immune Defic Syndr Hum Retrovirol 1998;19:203-9.

**27.** Lee CG, Gottesman MM, Cardarelli CO, et al. HIV-1 protease inhibitors are substrates for the MDR1 multidrug transporter. Biochemistry 1998;37:3594-601.

**28**. Drewe J, Gutmann H, Fricker G, Török M, Beglinger C, Huwyler J. HIV protease inhibitor ritonavir: a more potent inhibitor of P-glycoprotein than the cyclosporine analog SDZ PSC 833. Biochem Pharmacol 1999;57: 1147-52.

**29.** Profit L, Eagling VA, Back DJ. Modulation of P-glycoprotein function in human lymphocytes and Caco-2 cell monolayers by HIV-1 protease inhibitors. AIDS 1999;13:1623-7.

**30.** Benet LZ, Izumi T, Zhang Y, Silverman JA, Wacher VJ. Intestinal MDR transport proteins and P-450 enzymes as barriers to oral delivery. J Control Release 1999;62:25-31.

**31.** Wynn H, Shelton MJ, Bartos L, Difrancesco R, Hewitt R. Grapefruit juice (GJ) increases gastric pH, but does not affect indinavir (IDV) exposure, in HIV patients. In: Program and abstracts of the 39th Interscience Conference on Antimicrobial Agents and Chemotherapy, San Francisco, September 26–29, 1999. Washington, D.C.: American Society for Microbiology, 1999:25. abstract.

**32.** Soldner A, Christians U, Susanto M, Wacher VJ, Silverman JA, Benet LZ. Grapefruit juice activates P-glycoprotein-mediated drug transport. Pharm Res 1999;16:478-85.

**33**. Eagling VA, Profit L, Back DJ. Inhibition of CYP3A4-mediated metabolism and P-glycoprotein-mediated transport of the HIV-1 protease inhibitor saquinavir by grapefruit juice components. Br J Clin Pharmacol 1999;48:543-52.

**34.** Kim RB, Wandel C, Leake B, et al. Interrelationship between substrates and inhibitors of human CYP3A and P-glycoprotein. Pharm Res 1999;16:408-14.

**35.** Wacher VJ, Silverman JA, Zhang Y, Benet LZ. Role of P-glycoprotein and cytochrome P450 3A in limiting oral absorption of peptides and peptidomimetics. J Pharm Sci 1998;87:1322-30.

**36.** Fromm MF. P-glycoprotein: a defense mechanism limiting oral bioavailability and CNS accumulation of drugs. Int J Clin Pharmacol Ther 2000;38:69-74.

**37.** Piscitelli SC, Goss TF, Wilton JH, D'Andrea DT, Goldstein H, Schentag JJ. Effects of ranitidine and sucralfate on ketoconazole bioavailability. Antimicrob Agents Chemother 1991;35:1765-71.

Zimmermann T, Yeates RA, Laufen H, Pfaff G, Wildfeuer A. Influence of concomitant food intake on the oral absorption of two triazole antifungal agents, intraconazole and fluconazole. Eur J Clin Pharmacol 1994;46:147-50.
 Blum RA, D'Andrea DT, Florentino BM, et al. Increased gastric pH and the bioavailability of fluconazole and ketoconazole. Ann Intern Med 1991;114:755-7.

**40**. Laskin OL, de Miranda P, King DH, et al. Effects of probenecid on the pharmacokinetics and elimination of acyclovir in humans. Antimicrob Agents Chemother 1982;21:804-7.

**41.** Kornhauser DM, Petty BG, Hendrix CW, et al. Probenecid and zidovudine metabolism. Lancet 1989;2:473-5.

**42**. Kostrubsky VE, Ramachandran V, Venkataramanan R, et al. The use of human hepatocyte cultures to study the induction of cytochrome P-450. Drug Metab Dispos 1999;27:887-94.

**43.** Guibert A, Furlan V, Martino J, Taburet AM. In vitro effects of HIV protease inhibitors on methadone metabolism. In: Program and abstracts of the 37th Interscience Conference on Antimicrobial Agents and Chemotherapy, Toronto, September 28–October 1, 1997. Washington, D.C.: American Society for Microbiology, 1997:12. abstract.

**44.** Hsu A, Granneman GR, Carothers L, et al. Ritonavir does not increase methadone exposure in healthy volunteers. In: Program and abstracts of the 5th Conference on Retroviruses and Opportunistic Infections, Chicago, February 1–5, 1998:143. abstract.

**45.** Gerber JG, Rhodes RJ. Cytochrome P450 2B6 (CYP2B6) metabolizes methadone (M) preferentially and stereospecifically: an explanation of drug interaction with antiretroviral drugs. In: Final program of the First International Workshop on Clinical Pharmacology of HIV Therapy, Noordwijk, the Netherlands, March 30–31, 2000:14. abstract.

**46.** Gerber JG, Gal J, Rosenkranz S, et al. The effect of ritonavir (RTV)/ saquinavir (SQV) on the stereoselective pharmacokinetics (PK) of methadone (M). In: Abstracts of the XIII International AIDS Conference, Durban, South Africa, July 9–14, 2000:81. abstract.

**47**. Reinach B, de Sousa G, Dostert P, Ings R, Gugenheim J, Rahmani R. Comparative effects of rifabutin and rifampicin on cytochromes P450 and UDP-glucuronosyl-transferases expression in fresh and cryopreserved human hepatocytes. Chem Biol Interact 1999;121:37-48.

**48.** Li AP, Reith MK, Rasmussen A, et al. Primary human hepatocytes as a tool for the evaluation of structure-activity relationship in cytochrome P450 induction potential of xenobiotics: evaluation of rifampin, rifapentine and rifabutin. Chem Biol Interact 1997;107:17-30.

**49.** Greenblatt DJ, von Moltke LL, Harmatz JS, et al. Alprazolam-ritonavir interaction: implications for product labeling. Clin Pharmacol Ther 2000:67:335-41.

**50.** Sim SM, Hoggard PG, Sales SD, Phiboonbanakit D, Hart CA, Back DJ. Effect of ribavirin on zidovudine efficacy and toxicity in vitro: a concentration-dependent interaction. AIDS Res Hum Retroviruses 1998;14: 1661-7.

**51**. Back D, Haworth S, Hoggard P, Khoo S, Barry M. Drug interactions with d4T phosphorylation *in vitro*. In: Abstracts of the XI International Conference on AIDS, Vol. 1, Vancouver B.C., July 7–12, 1996:88. abstract.

**52.** Palmer S, Shafer RW, Merigan TC. Hydroxyurea enhances the activities of didanosine, 9-[2-(phosphonylmethoxy)ethyl]adenine, and 9-[2-(phosphonylmethoxy)propyl]adenine against drug susceptible and drug-resistant human immunodeficiency virus isolates. Antimicrob Agents Chemother 1999:43:2046-50.

**53.** Weissman SB, Sinclair GI, Green CL, Fissell WH. Hydroxyureainduced hepatitis in human immunodeficiency virus-positive patients. Clin Infect Dis 1999;29:223-4.

54. Prevention and treatment of tuberculosis among patients infected with human immunodeficiency virus: principles of therapy and revised recommendations. MMWR Morb Mortal Wkly Rep 1998;47(RR-20):1-58.
55. Falloon J, Piscitelli S, Vogel S, et al. Combination therapy with amprenavir, abacavir, and efavirenz in human immunodeficiency virus (HIV)-infected patients failing a protease-inhibitor regimen: pharmacokinetic

drug interactions and antiviral activity. Clin Infect Dis 2000;30:313-8. **56.** Fichtenbaum C, Gerber J, Rosenkranz S, et al. Pharmacokinetic interactions between protease inhibitors and selected HMG-CoA reductase inhibitors. In: Program and abstracts of the 7th Conference on Retroviruses and Opportunistic Infections, San Francisco, January 30–February 2, 2000:236. abstract.

57. Kaletra (Lopinavir-ritonavir). Abbott Park, Ill.: Abbott Laboratories, September 2000 (package insert).

**58.** Murphy RL, Sommadossi JP, Lamson M, Hall DB, Myers M, Dusek A. Antiviral effect and pharmacokinetic interaction between nevirapine and indinavir in persons infected with human immunodeficiency virus type 1. J Infect Dis 1999;179:1116-23.

**59.** Fiske WD, Mayers D, Wagner K, et al. Pharmacokinetics of DMP 266 and indinavir multiple oral doses in HIV-1 infected individuals. In: Program and abstracts of the 4th Conference on Retroviruses and Opportunitic Infectione Weakington DC. Jonus 22, 26 (1997) 169, abstract

60. Sahai J, Cameron W, Salgo M, et al. Drug interaction study between saquinavir (SQV) and nevirapine (NVP). In: Program and abstracts of the 4th Conference on Retroviruses and Opportunistic Infections, Washington, D.C., January 22–26, 1997:178. abstract.

**61.** Muirhead GJ, Wulff MB, Fielding A, Kleinermans D, Buss N. Pharmacokinetic interactions between sildenafil and saquinavir/ritonavir. Br J Clin Pharmacol 2000;50:99-107.

**62**. Merry C, Barry MG, Ryan M, et al. Interaction of sildenafil and indinavir when co-administered to HIV-positive patients. AIDS 1999;13:F101-F107.

**63.** Skowron G, Leoung G, Dusek A, et al. Stavudine (d4T), nelfinavir (NFV) and nevirapine (NVP): preliminary safety, activity and pharmacokinetic (PK) interactions. In: Program and abstracts of the 5th Conference on Retroviruses and Opportunistic Infections, Chicago, February 1–5, 1998:145. abstract.

**64.** Moyle GJ. Efavirenz: shifting the paradigm in adult HIV-1 infection. Exp Opin Invest Drugs 1999;8:473-86.

**65.** Fiske W, Benedek IH, Joseph JL, et al. Pharmacokinetics of efavirenz (EFV) and ritonavir (RIT) after multiple oral doses in healthy volunteers.

In: Conference record of the 12th World AIDS Conference, Geneva, June 28–July 3, 1998:827. abstract.
66. Fiske WD, Benedek IH, White SJ, Pepperess KA, Joseph JL, Korn-

**66.** Fiske WD, Benedek IH, White SJ, Pepperess KA, Joseph JL, Kornhauser DM. Pharmacokinetic interaction between efavirenz (EFV) and nelfinavir mesylate (NFV) in healthy volunteers. In: Program and abstracts of the 5th Conference on Retroviruses and Opportunistic Infections, Chicago, February 1–5, 1998:144. abstract.

**67.** Piketty C, Race E, Castiel P, et al. Efficacy of a five-drug combination including ritonavir, saquinavir and efavirenz in patients who failed on a conventional triple-drug regimen: phenotypic resistance to protease inhibitors predicts outcome of therapy. AIDS 1999;13:F71-F77.

**68.** Altice FL, Cooney E, Friedland GH. Nevirapine induced methadone withdrawal: implications for antiretroviral treatment of opiate dependent HIV infected patients. In: Program and abstracts of the 6th Conference on Retroviruses and Opportunistic Infections, Chicago, January 31–February 4, 1999:137. abstract.

**69.** Clarke S, Mulcahy F, Back D, Gibbons S, Tija J, Barry M. Managing methadone and non-nucleoside reverse transcriptase inhibitors: guidelines for clinical practice. In: Program and abstracts of the 7th Conference on Retroviruses and Opportunistic Infections, San Francisco, January 30–February 2, 2000:91. abstract.

**70.** Rescriptor (delavirdine). La Jolla, Calif.: Agouron Pharmaceuticals, 2000 (package insert).

**71.** Hsu A, Granneman GR, Witt G, et al. Multiple-dose pharmacokinetics of ritonavir in human immunodeficiency virus-infected subjects. Antimicrob Agents Chemother 1997;41:898-905.

**72.** Hsu Å, Granneman GR, Witt G, Cavanaugh JH, Leonard J. Assessment of multiple doses of ritonavir on the pharmacokinetics of theophylline. In: Abstracts of the XI International Conference on AIDS, Vancouver, B.C., July 7–12, 1996:89. abstract.

**73.** Ouellet D, Hsu A, Qian J, et al. Effect of ritonavir on the pharmacokinetics of ethinyl oestradiol in healthy female volunteers. Br J Clin Pharmacol 1998;46:111-6.

**74.** Pai VB, Nahata MC. Nelfinavir mesylate: a protease inhibitor. Ann Pharmacother 1999;33:325-39.

**75.** Veldkamp AI, Hoetelmans RM, Beijnen JH, Mulder JW, Meenhorst PL. Ritonavir enables combined therapy with rifampin and saquinavir. Clin Infect Dis 1999;29:1586.

**76.** Hamzeh F, Benson C, Gerber J, et al. Steady-state pharmacokinetic interaction of modified-dose indinavir and rifabutin. In: Program and abstracts of the 7th Conference on Retroviruses and Opportunistic Infections, San Francisco, January 30–February 2, 2000:92. abstract.

**77.** Cato A III, Cavanaugh J, Shi H, Hsu A, Leonard J, Granneman R. The effect of multiple doses of ritonavir on the pharmacokinetics of rifabutin. Clin Pharmacol Ther 1998;63:414-21.

**78.** Sun E, Heath-Chiozzi M, Cameron DW, et al. Concurrent ritonavir and rifabutin increases risk of rifabutin-associated adverse effects. In: Abstracts of the XI International Conference on AIDS, Vol. 1, Vancouver, B.C., July 7–12, 1996:18. abstract.

**79.** Updated guidelines for the use of rifabutin or rifampin for the treatment and prevention of tuberculosis among HIV-infected patients taking protease inhibitors or nonnucleoside reverse transcriptase inhibitors. MMWR Morb Mortal Wkly Rep 2000;49:185-9.

**80**. Hugen PWH, Burger DM, Brinkman K, et al. Carbamazepine-indinavir interaction causes antiretroviral failure. Ann Pharmacother 2000;34: 465-70.

**81.** Hsu A, Granneman GR, Bertz RJ. Ritonavir: clinical pharmacokinetics and interactions with other anti-HIV agents. Clin Pharmacokinet 1998;35: 275-91. [Erratum, Clin Pharmacokinet 1998;35:473.]

**82.** Shelton MJ, Cloen D, Becker M, Hsyu PH, Wilton JW, Hewitt RG. Evaluation of the pharmacokinetic (PK) interaction between phenytoin (Phen) and nelfinavir (NFV) in healthy volunteers at steady state. In: Pro-

gram and abstracts of the 40th Interscience Conference on Antimicrobial Agents and Chemotherapy, Toronto, September 17–20, 2000. Washington, D.C.: American Society for Microbiology, 2000:14. abstract.
83. Ouellet D, Hsu A, Granneman GR, et al. Pharmacokinetic interaction

**83**. Ouellet D, Hsu A, Granneman GR, et al. Pharmacokinetic interaction between ritonavir and clarithromycin. Clin Pharmacol Ther 1998;64:355-62.

**84**. Boruchoff SW, Sturgill MG, Grasing KW, et al. The steady-state disposition of indinavir is not altered by the concomitant administration of clarithromycin. Clin Pharmacol Ther 2000;67:351-9.

**85.** Harris S, Hilligoss DM, Colangelo PM, Eller M, Okerholm R. Azithromycin and terfenadine: lack of drug interaction. Clin Pharmacol Ther 1995;58:310-5.

**86.** Saah AJ, Winchell G, Seniuk M, Deutsch P. Multiple-dose pharmacokinetics (PK) and tolerability of indinavir (IDV) ritonavir (RTV) combinations in healthy volunteers. In: Program and abstracts of the 6th Conference on Retroviruses and Opportunistic Infections, Chicago, January 31– February 4, 1999:136. abstract.

87. Khaliq Y, Gallicano K, Sahai J, et al. Effect of nelfinavir (NFV) on

short and long term plasma exposure of saquinavir in hard gel capsule (SQV-HGC) during TID and BID dosing regimens. AIDS 1998;12:Suppl 4:S28. abstract.

**88.** van Heeswijk RPG, Veldkamp AI, Hoetelmans RMW, et al. The steady-state plasma pharmacokinetics of indinavir alone and in combination with a low dose of ritonavir in twice daily dosing regimens in HIV-1-infected individuals. AIDS 1999;13:F95-F99.

**89.** Workman C, Whittaker W, Dyer W, Sullivan J. Combining ritonavir and indinavir decreases IDV associated nephrolithiasis. In: Program and abstracts of the 6th Conference on Retroviruses and Opportunistic Infections, Chicago, January 31–February 4, 1999:195. abstract.

**90**. Piscitelli S, Bechtel C, Sadler B, Falloon J, Intramural AIDS Program. The addition of a second protease inhibitor eliminates amprenavir-efavirenz drug interactions and increases plasma amprenavir concentrations. In: Program and abstracts of the 7th Conference on Retroviruses and Opportun-

istic Infections, San Francisco, January 30–February 2, 2000:90. abstract. **91.** Hendrix CW, Fiske WD, Fuchs EJ, et al. Pharmacokinetics of the triple combination of saquinavir, ritonavir, and efavirenz in HIV-positive patients. In: Program and abstracts of the 7th Conference on Retroviruses and Opportunistic Infections, San Francisco, January 30–February 2, 2000:90. abstract.

**92.** Lal R, Hsu A, Granneman GR, et al. Multiple dose safety, tolerability and pharmacokinetics of ABT-378 in combination with ritonavir. In: Program and abstracts of the 5th Conference on Retroviruses and Opportunistic Infections, Chicago, February 1–5, 1998:201. abstract.

**93.** Baldwin JR, Borin MT, Ferry JJ, et al. Pharmacokinetic (PK) interaction between the HIV protease inhibitors tipranavir and ritonavir. In: Program and abstracts of the 39th Interscience Conference on Antimicrobial Agents and Chemotherapy, San Francisco, September 26–29, 1999. Washington, D.C.: American Society for Microbiology, 1999:24. abstract.

**94.** Ferry JJ, Herman BD, Carel BJ, Carlson GF, Batts DH. Pharamcokinetic drug-drug interaction study of delavirdine and indinavir in healthy

volunteers. J Acquir Immune Defic Syndr Hum Retrovirol 1998;18:252-9. **95.** Cox SR, Ferry JJ, Batts DH, et al. Delavirdine (D) and marketed protease inhibitors (PIs): pharmacokinetic (PK) interaction studies in healthy volunteers. In: Program and abstracts of the 4th Conference on Retroviruses and Opportunistic Infections, Washington, D.C., January 22–26, 1997:133. abstract.

96. Fortovase (saquinavir). Nutley, N.J.: Roche Laboratories, 1999 (pack-age insert).

**97.** Varhe A, Olkkola KT, Neuvonen PJ. Effect of fluconazole dose on the extent of fluconazole-triazolam interaction. Br J Clin Pharmacol 1996;42: 465-70.

**98.** Sahai J, Gallicano K, Pakuts A, Cameron DW. Effect of fluconazole on zidovudine pharmacokinetics in patients infected with human immuno-deficiency virus. J Infect Dis 1994;169:1103-7.

**99.** Crussell-Porter LL, Rindone JP, Ford MA, Jaskar DW. Low-dose fluconazole therapy potentiates the hypoprothrombinemic response of warfarin sodium. Arch Intern Med 1993;153:102-4.

**100.** Hafner R, Bethel J, Power M, et al. Tolerance and pharmacokinetic interactions of rifabutin and clarithromycin in human immunodeficiency

virus-infected volunteers. Antimicrob Ágents Chemother 1998;42:631-9. **101.** Wallace RJ Jr, Brown BA, Griffith DE, Girard W, Tanaka K. Reduced serum levels of clarithromycin in patients treated with multidrug regimens including rifampin or rifabutin for *Mycobacterium avium-M. intracellulare* infection. J Infect Dis 1995;171:747-50.

**102.** Trapnell CB, Narang PK, Li R, Lavelle JP. Increased plasma rifabutin levels with concomitant fluconazole therapy in HIV-infected patients. Ann Intern Med 1996;124:573-6.

**103.** Narang PK, Trapnell CB, Schoenfelder JR, Lavelle JP, Biachine JR. Fluconazole and enhanced effect of rifabutin prophylaxis. N Engl J Med 1994;330:1316-7.

**104.** Piscitelli SC, Burstein AH, Chaitt D, Alfaro RM, Falloon J. Indinavir concentrations and St John's wort. Lancet 2000;355:547-8.

**105.** Roby CA, Anderson GD, Kantor E, Dryer DA, Burstein AH. St John's wort: effect on CYP3A4 activity. Clin Pharmacol Ther 2000;67: 451-7.

**106.** Johne A, Brockmoller J, Bauer S, Maurer A, Langheinrich M, Roots I. Pharmacokinetic interaction of digoxin with an herbal extract from St John's wort (Hypericum perforatum). Clin Pharmacol Ther 1999;66:338-45.

**107.** Foster BC, Gallicano K, Cameron W, Choudhri SH. Constituents of garlic inhibit cytochrome P450 3A4 mediated drug metabolism. Can J Infect Dis 1998;9:Suppl A:472P. abstract.

**108.** Dalvi RR. Alterations in hepatic phase I and phase II biotransformation enzymes by garlic oil in rats. Toxicol Lett 1992;60:299-305.

**109.** Laroche M, Choudhri S, Gallicano K, Foster B. Severe gastrointestinal toxicity with concomitant ingestion of ritonavir and garlic. Can J Infect Dis 1998;9:Suppl A:471P. abstract.

**110.** Piscitelli SC. Use of complementary medicines by patients with HIV infection: full sail into uncharted waters. May 2000. (See http://www.medscape.com/medscape/HIV/journal/2000/v06.n03/

mha0605.pisc/mha0605.pisc-01.html.) (See NAPS document no. 05584 for 6 pages, c/o Microfiche Publications, 248 Hempstead Tpke., West Hempstead, NY 11552.)

**111.** Renton KW, Knickle LC. Regulation of hepatic cytochrome P-450 during infectious disease. Can J Physiol Pharmacol 1990;68:777-81.

**112.** Reiss WG, Piscitelli SC. Drug-cytokine interactions: mechanisms and clinical implications. Biodrugs 1998;9:389-95.

**113.** Schwartz DH, Merigan TC. Interleukin-2 in the treatment of HIV disease. Biotherapy 1990;2:119-36.

**114.** Whittington R, Faulds D. Interleukin-2: a review of its pharmacological properties and therapeutic use in patients with cancer. Drugs 1993;46: 446-514.

**115.** Piscitelli SC, Vogel S, Figg WD, et al. Alteration in indinavir clearance during interleukin-2 infusions in patients infected with the human immunodeficiency virus. Pharmacotherapy 1998;18:1212-6.

**116.** Molla A, Korneyeva M, Gao Q, et al. Ordered accumulation of mutations in HIV protease confers resistance to ritonavir. Nat Med 1996;2: 760-6.

**117.** Acosta EP, Henry K, Baken L, Page LM, Fletcher CV. Indinavir concentrations and antiviral effect. Pharmacotherapy 1999;19:708-12.

**118.** Burger DM, Hoetelmans RMW, Mulder JW, et al. Low plasma levels of indinavir (IDV) are highly predictive of virological treatment failure in patients using IDV-containing triple therapy. In: Conference record of the 12th World AIDS Conference, Geneva, June 28–July 3, 1998:828. abstract.

**119.** Acosta EP. The promise of therapeutic drug monitoring in HIV infection. August 1999. (See http://www.medscape.com/medscape/HIV/journal/1999/v05.n04/mha0803/mha0803.acos/mha0803.acos-01.html.) (See NAPS document no. 05584 for 5 pages, c/o Microfiche Publications, 248 Hempstead Tpke., West Hempstead, NY 11552.)

**120.** Piscitelli SC. The limited value of therapeutic drug monitoring in HIV infection. August 1999. (See http://www.medscape.com/medscape/HIV/journal/1999/v05.n04/mha0803/mha0803.pisc/

mha0803.pisc-01.html.) (See NAPS document no. 05584 for 4 pages, c/o Microfiche Publications, 248 Hempstead Tpke., West Hempstead, NY 11552.)

**121.** Back DJ, Gibbons SE, Khoo SH, Merry C, Barry MG, Mulcahy FM. Therapeutic drug monitoring of antiretrovirals: ready for the clinic? J Int Assoc Physicians AIDS Care 2000;6:34-7.

Copyright © 2001 Massachusetts Medical Society.