

## Rational Design of Envelope Identifies Broadly Neutralizing Human Monoclonal Antibodies to HIV-1

Xueling Wu,<sup>1\*</sup> Zhi-Yong Yang,<sup>1\*</sup> Yuxing Li,<sup>1\*</sup> Carl-Magnus Hogerkorp,<sup>1,†</sup> William R. Schief,<sup>4</sup> Michael S. Seaman,<sup>5</sup> Tongqing Zhou,<sup>1</sup> Stephen D. Schmidt,<sup>1</sup> Lan Wu,<sup>1</sup> Ling Xu,<sup>1</sup> Nancy S. Longo,<sup>1</sup> Krisha McKee,<sup>1</sup> Sijy O'Dell,<sup>1</sup> Mark K. Louder,<sup>1</sup> Diane L. Wycuff,<sup>1</sup> Yu Feng,<sup>1,‡</sup> Martha Nason,<sup>2</sup> Nicole Doria-Rose,<sup>3</sup> Mark Connors,<sup>3</sup> Peter D. Kwong,<sup>1</sup> Mario Roederer,<sup>1</sup> Richard T. Wyatt,<sup>1,‡</sup> Gary J. Nabel,<sup>1,§</sup> John R. Mascola<sup>1§</sup>

<sup>1</sup>Vaccine Research Center, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, MD 20892, USA. <sup>2</sup>Biostatistics Research Branch, Division of Clinical Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, MD 20892, USA. <sup>3</sup>Laboratory of Immunoregulation, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, MD 20892, USA. <sup>4</sup>Department of Biochemistry, University of Washington, Seattle, WA 98195, USA. <sup>5</sup>Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA 02115, USA.

\*These authors contributed equally to this work.

†Present address: Department of Molecular Genetics, Novo Nordisk A/S, Novo Nordisk Park, 2760 Måløv, Denmark.

‡Present address: Department of Immunology and Microbial Science, and IAVI Center for Neutralizing Antibody at TSRI, The Scripps Research Institute, La Jolla, CA 92037, USA.

§To whom correspondence should be addressed. E-mail: gnabel@nih.gov (G.J.N.); jmascola@nih.gov (J.R.M.)

**Cross-reactive neutralizing antibodies (NAb) are found in the sera of many HIV-1-infected subjects, but the virologic basis of their neutralization remains poorly understood. We used knowledge of HIV-1 envelope (Env) structure to develop antigenically resurfaced glycoproteins specific for the structurally conserved site of CD4 receptor binding. These probes were used to identify sera with NAb to the CD4-binding site (CD4bs) and to isolate individual B cells from such an HIV-1-infected donor. By expressing immunoglobulin genes from individual cells, we identified three monoclonal antibodies, including a pair of somatic variants that neutralized over 90% of circulating HIV-1 isolates. Exceptionally broad HIV-1 neutralization can be achieved with individual antibodies targeted to the functionally conserved CD4bs of gp120, an important insight for future HIV-1 vaccine design.**

Having crossed from chimpanzees to humans in only the last century, HIV-1 has rapidly evolved a daunting degree of diversity posing a considerable challenge for vaccine development. The definition of naturally-occurring broadly neutralizing antibodies (NAb) has proven elusive, and the ability to target conserved determinants of the viral envelope (Env) has proven difficult (1-2). During HIV-1 infection, almost all individuals produce antibodies to Env, but only a small fraction can neutralize the virus (1-2). Recently, several groups have shown that the sera of 10 – 25% of infected

subjects contain broadly reactive NAb (3-6), including some sera that neutralize the majority of viruses from diverse genetic subtypes (5-7). NAb react with the HIV-1 Env spike, which is composed of three heavily glycosylated gp120 molecules, each noncovalently associated with a transmembrane gp41 molecule. To initiate viral entry into cells, the gp120 binds to the cell surface receptor CD4 (8). We previously reported that selected sera contain NAb directed against the CD4-binding site (CD4bs) of gp120 (7, 9), and we defined the structure of the CD4bs in complex with the neutralizing monoclonal antibody (mAb) b12 (10-11). Antibody b12 was isolated from a phage display library in 1992 and can neutralize approximately 40% of known HIV-1 isolates (12-14). The more recently isolated CD4bs mAb HJ16 also neutralizes about 40% of viral isolates (15). Attempts to isolate more broadly reactive CD4bs-directed NAb from human B cells have not met with success, in part because the gp120 or gp41 proteins used were reactive with many HIV-1 specific antibodies, including non-neutralizing antibody specificities (15-16). In this study, we used our knowledge of Env structure, together with computer-assisted protein design, to define recombinant forms of HIV-1 Env that specifically interact with NAb directed to the CD4bs. These Env probes were used to identify and sort individual B cells expressing CD4bs antibodies, enabling the selective isolation of CD4bs-directed mAbs with extensive neutralization breadth.

To generate a molecule that preserved the antigenic structure of the neutralizing surface of the CD4bs, but eliminated other antigenic regions of HIV-1, we designed proteins whose exposed surface residues were substituted with SIV homologues and other non-HIV-1 residues (17) (Fig. 1A and fig. S1). These changes were conferred on a core gp120 and a stabilized core gp120, both of which retained the major contact surface for CD4 located on its outer domain (10-11, 18). The gp120 core lacked variable regions 1-3 and part of the amino and carboxy termini of the full gp120 molecule, and the stabilized core contained cross-links between different subregions of the core protein. Eight resurfaced proteins were designed and expressed, together with CD4bs mutants that served as negative controls by eliminating binding to the neutralizing mAb b12. Three resurfaced core Env s retained reactivity with b12 and mAb 2G12, the latter of which recognizes a surface glycan epitope and served as a positive control for a conformationally intact protein. The resurfaced stabilized core 3 (RSC3) was chosen as the preferred candidate for further studies, because a greater percentage of its surface other than the outer domain CD4bs area was altered compared to the other variants (fig. S2). The conformational integrity and specificity of the RSC3 protein was confirmed using a panel of known mAbs (table S1). As expected, RSC3 displayed strong reactivity to mAb b12 and little or no reactivity to a CD4 fusion protein (Fig. 1B and 1C) (19). RSC3 also reacted with two weakly neutralizing CD4bs mAbs, b13 and m18, but it displayed no reactivity to four CD4bs mAbs that do not neutralize primary HIV-1 isolates, nor with mAbs directed to other regions of the HIV-1 Env, including the coreceptor-binding region of gp120 and the V3 and C5 regions of gp120 (table S1). ΔRSC3, which lacked a single amino acid at position 371 that eliminated b12 binding, served as a negative control. Together, these data confirmed the integrity of the antibody binding surface of this resurfaced protein, and it was used for analyses of sera and to identify B cells from an HIV-1 infected individual whose sera contained broadly reactive Nabs.

We screened a panel of broadly neutralizing sera for the presence of antibodies that could preferentially bind to RSC3 compared to ΔRSC3. CD4bs antibodies were detected in several sera, including serum from donor 45 which was previously reported to contain Nabs directed to the CD4bs of gp120 (fig. S3) (7). To determine whether antibodies that bind to RSC3 were responsible for the broad neutralization mediated by serum 45, we performed neutralization studies using RSC3 to compete cognate antibodies. The utility of this assay was confirmed with the CD4bs mAbs b12 and F105. mAb b12 neutralizes many primary HIV-1 strains, whereas F105 neutralizes mainly laboratory-adapted or other highly sensitive virus strains, such as the HXB2 strain used here.

The addition of RSC3, but not ΔRSC3, inhibited b12-mediated neutralization of HXB2. RSC3 had no effect on F105 neutralization, and neither RSC3 nor ΔRSC3 affected neutralization by the anti-V3 mAb 447-52D (Fig. 1D). To interrogate serum 45 neutralization, we performed similar RSC3 competition studies with this serum against a panel of diverse HIV-1 strains (Fig. 1E). This analysis suggested that serum 45 neutralization was principally directed against the CD4bs on functional viral spikes, and that the RSC3 faithfully mimicked this structure.

To isolate CD4bs-directed mAbs, we used a recently described method of antigen-specific memory B-cell sorting (16), together with single cell PCR, to amplify IgG heavy and light chain genes from the cDNA of individual B cells (16, 20). RSC3 and ΔRSC3 were expressed with a tagged amino acid sequence that allows biotin labeling. The two proteins could thus be distinguished by FACS analysis after labeling with streptavidin (SA) conjugated to the fluorochromes allophycocyanin (SA-APC) or phycoerythrin (SA-PE), respectively. Peripheral blood mononuclear cells (PBMC) from donor 45 were incubated with RSC3 SA-APC and ΔRSC3 SA-PE, and single antigen-specific memory B cells were sorted into wells of a microtiter plate after selecting for memory B cells ( $CD19^+$ ,  $CD20^+$ ,  $IgG^+$ ) that bound to the RSC3 but not ΔRSC3 probe (Fig. 2A). Out of approximately 25 million PBMC, 29 single RSC3-specific memory B cells were sorted and the matching heavy and light chain genes were successfully amplified from 12 cells. After cloning into IgG1 expression vectors that reconstituted the heavy and light chain constant regions, the full IgG mAbs were expressed. Three antibodies (VRC01, VRC02 and VRC03) bound strongly to RSC3 and weakly or not at all to ΔRSC3 (Fig. 2B, left panel, Fig. 2C and fig. S4). To confirm the specificity of these antibodies for the CD4bs, ELISA binding of each mAb was tested against a wild type gp120 and the CD4bs-defective D368R mutant (Fig. 2B, right panel). VRC01 and VRC02 bound with  $\geq$  100-fold lower relative affinity to the D368R mutant compared to wild type gp120, and VRC03 showed no detectable binding to the CD4bs knockout mutant. The ELISA binding profile to an extended panel of mutant Env proteins further confirmed the CD4bs specificity of VRC01, VRC02 and VRC03 (table S1).

Analysis of the heavy and light chain nucleotide sequences using JoinSolver software and the IMGT database revealed that VRC01 and VRC02 were somatic variants of the same IgG1 clone. The heavy-chain CDR3 region of both mAbs was composed of the same 14 amino acids (fig. S5) and both mAbs were highly somatically mutated, with 32% of VH and 17 - 19% of VK nucleotides divergent from putative germline gene sequences. VRC03 was a likely a unique IgG1 clone, but its heavy chain was derived from the same IGHV1-02\*02 andIGHJ1\*01 alleles as VRC01 and VRC02. VRC03 was

also highly somatically mutated, with an unusual 7 amino-acid insertion in heavy-chain framework 3, and 30% of VH and 20% of VK nucleotides divergent from germline gene sequences. The heavy-chain CDR3 of VRC03 contained 16 amino acids. All three mAbs share common sequence motifs in heavy chain CDR1, CDR2 and CDR3.

The binding characteristics of the mAbs were further analyzed by surface plasmon resonance (SPR), competition ELISA and isothermal titration calorimetry (ITC). SPR demonstrated that VRC01 ( $K_D = 3.88 \times 10^{-9}$  M), and VRC02 ( $K_D = 1.11 \times 10^{-8}$  M) bound gp120 with high affinity while VRC03 reacted with about 10-fold lower affinity ( $K_D = 7.31 \times 10^{-8}$  M) (fig. S4). To evaluate the epitope reactivity of these mAbs on gp120, competition ELISAs were performed with a panel of well characterized mAbs. As expected, binding by all three VRC mAbs was competed by CD4bs mAbs b12 and F105, and by CD4-Ig (Fig. 3A left panel and fig S6A). Unexpectedly, the binding of mAb 17b to its site in the coreceptor binding region of gp120 was markedly enhanced by the addition of VRC01 or VRC02 (Fig. 3A, right panel). This enhancing effect was similar, though not as profound, as the known effect of CD4-Ig. In contrast, mAb b12 inhibited mAb 17b binding (Fig. 3A, right panel), as previously shown (21). A similar enhancing result was observed for VRC01 in an assay that measures gp120 binding to its CCR5 coreceptor (fig. S6C). Thus, VRC01 and VRC02 act as partial CD4 agonists in their interaction with gp120, whereas VRC03 does not display this effect. Thermodynamic analysis by ITC provided data consistent with the ELISA results, and demonstrated a change in enthalpy ( $-\Delta H$ ) associated with the VRC01-gp120 interaction that was similar to the interaction of CD4-Ig and gp120 (Fig. 3B), further demonstrating that VRC01 binding induced conformational changes in gp120. In contrast to the data for gp120 binding, VRC01 did not enhance viral neutralization by mAb 17b (fig. S7). These data suggest that VRC01 and VRC02 partially mimic the interaction of CD4 with gp120. This may explain their broad reactivity, since essentially all HIV-1 isolates must engage CD4 for cell entry.

The potency and breadth of neutralization by VRC01, VRC02 and VRC03, compared to b12 and CD4-Ig, were assessed on a comprehensive panel of Env pseudoviruses (Fig. 4 and table S2). These 190 viral strains represented all major circulating HIV-1 genetic subtypes (clades) and included viruses derived from acute and chronic stages of HIV-1 infection (22-23). VRC01 neutralized 91% of these viruses with a geometric mean value of 0.33  $\mu\text{g/ml}$  (Fig. 4 and table S2). The data for VRC02 were very similar (table S2). Of note, these mAbs were derived from an HIV-1 clade B-infected donor yet displayed neutralization activity against all genetic subtypes of HIV-1. VRC03 was less broad than VRC01 and VRC02, neutralizing 57% of the viruses (table

S2) (24). In contrast, b12, also derived from a clade B donor, neutralized 41% of viruses tested. Since VRC01 was derived from a donor whose sera was also broadly neutralizing, we assessed the relationship between the neutralization breadth and potency of serum 45 IgG and mAb VRC01. Among 140 viruses tested, there was a significant association ( $P = 0.005$ ; Fisher's exact test) between the number of viruses neutralized by serum 45 IgG and the number neutralized by VRC01 (fig. S8A). Among the 122 viruses neutralized by both serum IgG and VRC01, there was a strong association ( $P < 0.0001$ ; Deming linear regression) between the neutralization potency of the serum IgG and the potency of VRC01 (fig. S8B). Therefore, although VRC01 did not account for all serum 45 IgG neutralization, the VRC01-like antibody specificity largely accounts for the extensive breadth and potency of serum 45. These findings demonstrate that a focused B-cell response can target a highly conserved region of the HIV-1 Env in humans (25).

Other mAbs are able to neutralize HIV-1, but none has a profile of potency and breadth similar to VRC01 and VRC02 (1, 13-14, 26-27). Antibody 4E10 requires relatively high concentrations to neutralize primary strains of HIV-1 (13) and it neutralizes only 12% of Env-pseudoviruses at a concentration of less than 1  $\mu\text{g/ml}$  (14). The well-characterized CD4bs mAb b12 (13-14) and the more recently described HJ16 (15) are informative with respect to antigen recognition, but each display restricted breadth (~ 40% of HIV-1 strains). Recently, two broadly neutralizing somatic variant mAbs, PG16 and PG9, were isolated by high-throughput neutralization screening of B-cell supernatants (14). The PG16 and PG9 neutralized 73% and 79% respectively of viruses tested, and recognize a glycosylated region of HIV-1 Env that is present on the native viral trimer, but this epitope is not well presented on gp120 or gp140.

VRC01 and VRC02 access the CD4bs region of gp120 in a manner that partially mimics the interaction of CD4 with gp120 (28). This observation may explain their impressive breadth of reactivity. The isolation of these mAbs from an HIV-1 infected subject and the demonstration that they neutralize the vast majority of HIV-1 strains by targeting the functionally conserved receptor binding region of Env, provides proof of concept that such antibodies can be elicited in humans. The discovery of these mAbs provides new insights into how the human immune system is able to effectively target a vulnerable site on the viral Env.

## References and Notes

1. R. Pantophlet, D. R. Burton, GP120: target for neutralizing HIV-1 antibodies. *Annu Rev Immunol* **24**, 739 (2006).
2. D. C. Montefiori, L. Morris, G. Ferrari, J. R. Mascola, Neutralizing and other antiviral antibodies in HIV-1 infection and vaccination. *Curr Opin HIV AIDS* **2**, 169 (2007).

3. L. Stamatatos, L. Morris, D. R. Burton, J. R. Mascola, Neutralizing antibodies generated during natural HIV-1 infection: good news for an HIV-1 vaccine? *Nat Med* **15**, 866 (2009).
4. D. N. Sather *et al.*, Factors associated with the development of cross-reactive neutralizing antibodies during human immunodeficiency virus type 1 infection. *J Virol* **83**, 757 (2009).
5. M. D. Simek *et al.*, Human immunodeficiency virus type 1 elite neutralizers: individuals with broad and potent neutralizing activity identified by using a high-throughput neutralization assay together with an analytical selection algorithm. *J Virol* **83**, 7337 (2009).
6. N. A. Doria-Rose *et al.*, Breadth of human immunodeficiency virus-specific neutralizing activity in sera: clustering analysis and association with clinical variables. *J Virol* **84**, 1631 (2010).
7. Y. Li *et al.*, Broad HIV-1 neutralization mediated by CD4-binding site antibodies. *Nat Med* **13**, 1032 (2007).
8. R. Wyatt *et al.*, The antigenic structure of the HIV gp120 envelope glycoprotein. *Nature* **393**, 705 (1998).
9. Y. Li *et al.*, Analysis of neutralization specificities in polyclonal sera derived from human immunodeficiency virus type 1-infected individuals. *J Virol* **83**, 1045 (2009).
10. P. D. Kwong *et al.*, Structure of an HIV gp120 envelope glycoprotein in complex with the CD4 receptor and a neutralizing human antibody. *Nature* **393**, 648 (1998).
11. T. Zhou *et al.*, Structural definition of a conserved neutralization epitope on HIV-1 gp120. *Nature* **445**, 732 (2007).
12. D. R. Burton *et al.*, Efficient neutralization of primary isolates of HIV-1 by a recombinant human monoclonal antibody. *Science* **266**, 1024 (1994).
13. J. M. Binley *et al.*, Comprehensive cross-clade neutralization analysis of a panel of anti-human immunodeficiency virus type 1 monoclonal antibodies. *J Virol* **78**, 13232 (2004).
14. L. M. Walker *et al.*, Broad and potent neutralizing antibodies from an African donor reveal a new HIV-1 vaccine target. *Science* **326**, 285 (2009).
15. D. Corti *et al.*, Analysis of memory B cell responses and isolation of novel monoclonal antibodies with neutralizing breadth from HIV-1-infected individuals. *PLoS One* **5**, e8805 (2010).
16. J. F. Scheid *et al.*, Broad diversity of neutralizing antibodies isolated from memory B cells in HIV-infected individuals. *Nature* **458**, 636 (2009).
17. Materials and Methods are available as supporting material on Science Online.
18. B. Dey *et al.*, Characterization of human immunodeficiency virus type 1 monomeric and trimeric gp120 glycoproteins stabilized in the CD4-bound state: antigenicity, biophysics, and immunogenicity. *J Virol* **81**, 5579 (2007).
19. The CD4-binding property of the core proteins would result in their binding to circulating CD4-expressing cells, such as CD4+ T cells in peripheral blood. Since this interaction would confound the selection of B cells expressing CD4bs IgG, we additionally sought to design proteins containing the majority of the CD4bs epitope surface that maintained b12 reactivity, but that no longer bound to CD4. Specifically, computer-assisted modeling was used to alter the β20/21 region of gp120 that is required for CD4 binding, but is not part of the b12-contact surface (fig. S1).
20. J. Wrammert *et al.*, Rapid cloning of high-affinity human monoclonal antibodies against influenza virus. *Nature* **453**, 667 (2008).
21. J. P. Moore, J. Sodroski, Antibody cross-competition analysis of the human immunodeficiency virus type 1 gp120 exterior envelope glycoprotein. *J Virol* **70**, 1863 (1996).
22. B. Korber, S. Gnanakaran, The implications of patterns in HIV diversity for neutralizing antibody induction and susceptibility. *Curr Opin HIV AIDS* **4**, 408 (2009).
23. M. S. Seaman *et al.*, Tiered categorization of a diverse panel of HIV-1 Env pseudoviruses for neutralizing antibody assessment. *J Virol* **84**, 1439 (2010).
24. Additional studies using primary PBMC-derived viral isolates and PBMC as target cells confirmed that VRC01, VRC02 and VRC03 were able to neutralize replication-competent uncloned HIV-1 isolates grown in primary T-cells (table S3). We also tested the impact of VRC01 on the functional viral spike because VRC01 and VRC02, like CD4, showed the ability to alter the conformation of gp120 and enhanced binding of CD4-induced antibodies like 17b. Unlike CD4 however, VRC01 did not promote the entry of primary HIV-1 isolates into CD4-negative cells, and it did not augment the neutralization potency of mAb 17b (fig. S7).
25. Our successful isolation of CD4bs mAbs with extensive neutralization breadth was likely due to several factors. Recent advances in high-throughput neutralization assays facilitated the identification of HIV-1 antisera containing broad Nabs (4-7, 29-32). Advances in serum epitope mapping technologies led us to understand that conserved regions of HIV-1 Env may be targeted by such Nabs (4, 7, 9, 30, 33-37). The atomic level structure of gp120 (10, 38) and advances in computational modeling allowed development of the RSC probe pair that was used to identify and sort CD4bs antibody-secreting B cells. Due to the large repertoire of circulating anti-gp120 antibodies, most of them non-neutralizing, the epitope selectivity conferred by the RSC3 protein, together with use of the

- knock-out mutant, was likely important for the identification of the relatively rare memory B cells secreting NAbs to the CD4bs. Recently, the gp140 form of Env was used to identify HIV-1-specific B cells and study the repertoire of the antibody responses to the Env glycoprotein (16). Numerous unique antibodies were derived from each of 6 HIV-1 infected donors, but no single mAb demonstrated broad and potent neutralization. The ability to identify epitope-specific B cells, and isolate the specific IgG clones, has broad potential application. A similar epitope-specific approach could be used to isolate mAbs to sites of known structure from infectious disease pathogens, including tuberculosis and malaria, and can be applied to the identification of targets of autoimmunity or cancer therapy.
26. S. Zolla-Pazner, Identifying epitopes of HIV-1 that induce protective antibodies. *Nat Rev Immunol* **4**, 199 (2004).
  27. A. Forsman *et al.*, Llama antibody fragments with cross-subtype human immunodeficiency virus type 1 (HIV-1)-neutralizing properties and high affinity for HIV-1 gp120. *J Virol* **82**, 12069 (2008).
  28. Unlike CD4, VRC01 and VRC02 do not appear to require interaction with the conformationally mobile β20/21 region of gp120, because this region was altered in the RSC3 protein.
  29. S. G. Deeks *et al.*, Neutralizing antibody responses against autologous and heterologous viruses in acute versus chronic human immunodeficiency virus (HIV) infection: evidence for a constraint on the ability of HIV to completely evade neutralizing antibody responses. *J Virol* **80**, 6155 (2006).
  30. E. S. Gray *et al.*, Antibody specificities associated with neutralization breadth in plasma from human immunodeficiency virus type 1 subtype C-infected blood donors. *J Virol* **83**, 8925 (2009).
  31. D. M. Smith *et al.*, Lack of neutralizing antibody response to HIV-1 predisposes to superinfection. *Virology* **355**, 1 (2006).
  32. D. D. Richman, T. Wrin, S. J. Little, C. J. Petropoulos, Rapid evolution of the neutralizing antibody response to HIV type 1 infection. *Proc Natl Acad Sci U S A* **100**, 4144 (2003).
  33. J. M. Binley *et al.*, Profiling the specificity of neutralizing antibodies in a large panel of plasmas from patients chronically infected with human immunodeficiency virus type 1 subtypes B and C. *J Virol* **82**, 11651 (2008).
  34. E. Yuste *et al.*, Simian immunodeficiency virus engrafted with human immunodeficiency virus type 1 (HIV-1)-specific epitopes: replication, neutralization, and survey of HIV-1-positive plasma. *J Virol* **80**, 3030 (2006).
  35. A. K. Dhillon *et al.*, Dissecting the neutralizing antibody specificities of broadly neutralizing sera from human immunodeficiency virus type 1-infected donors. *J Virol* **81**, 6548 (2007).
  36. J. M. Decker *et al.*, Antigenic conservation and immunogenicity of the HIV coreceptor binding site. *J Exp Med* **201**, 1407 (2005).
  37. X. Shen *et al.*, In vivo gp41 antibodies targeting the 2F5 monoclonal antibody epitope mediate human immunodeficiency virus type 1 neutralization breadth. *J Virol* **83**, 3617 (2009).
  38. B. Chen *et al.*, Structure of an unliganded simian immunodeficiency virus gp120 core. *Nature* **433**, 834 (2005).
  39. This research was supported by the Intramural Research Program of the Vaccine Research Center, the Division of Clinical Research and the Laboratory of Immunoregulation of NIAID, National Institutes of Health. MSS was supported in part by the Bill & Melinda Gates Foundation's Collaboration for AIDS Vaccine Discovery, Comprehensive Antibody-Vaccine Immune Monitoring Consortium, grant number 38619. WRS was supported by the Bill and Melinda Gates Foundation Collaboration for AIDS Vaccine Discovery through a grant to Seattle Biomedical Research Institute, and by the International AIDS Vaccine Initiative. Nucleotide sequences of VRC01, VRC02 and VRC03 variable regions are available under GenBank accession numbers GU980702 to GU980707. A provisional patent application on mAbs VRC01, VRC02 and VRC03 has been filed by the NIH (listed investors are: XW, ZYY, YL, CMH, WRS, MC, PDK, MR, RTW, GJN and JRM). We thank Michel Nussenzweig and Johannes Scheid (Laboratory of Molecular Immunology, Howard Hughes Medical Institute, The Rockefeller University, New York, NY) for sharing expertise and reagents for IgG cloning and expression, and Bart Haynes (Duke University Medical Center) for sharing reagents and PCR methods. We are grateful to Steve Perfetto, Richard Nguyen and David Ambrozak for assistance with cell sorting; Elizabeth Lybarger for neutralization assays; Jonathan Stuckey and Brenda Hartman for assistance with figures; Ati Tislerics for manuscript preparation; Dean Follman for statistical advice; Chris Carrico for discussions on resurfacing, Yih-En Andrew Ban and Leo Kong for glycan modeling; and Pratip Chattopadhyay and Joanne Yu for fluorescent antibody production and qualification. We also thank Carolyn Williamson (University of Capetown, Capetown, South Africa), Michael Thomson (Instituto de Salud Carlos III, Madrid, Spain), Julie Overbaugh (Fred Hutchinson Cancer Research Center, Seattle, WA) and Sodsai Tovanabutra and Eric Sanders-Buell (Henry M. Jackson Foundation and the U.S. Military HIV Research Program, Rockville, MD) for contributing HIV-1 Env

plasmids for pseudovirus production. A patent application has been filed by NIH pertaining to the resurfacing technology used to generate the RSC3 probe and the probe itself (authors G.J.N., W.R.S., Z-Y.Y., T.Z., and P.D.K.).

## Supporting Online Material

[www.sciencemag.org/cgi/content/full/science.1187659/DC1](http://www.sciencemag.org/cgi/content/full/science.1187659/DC1)

### Materials and Methods

Figs. S1 to S8

Tables S1 to S3

References

28 January 2010; accepted 04 May 2010

Published online 08 July 2010; 10.1126/science.1187659

Include this information when publishing this paper.

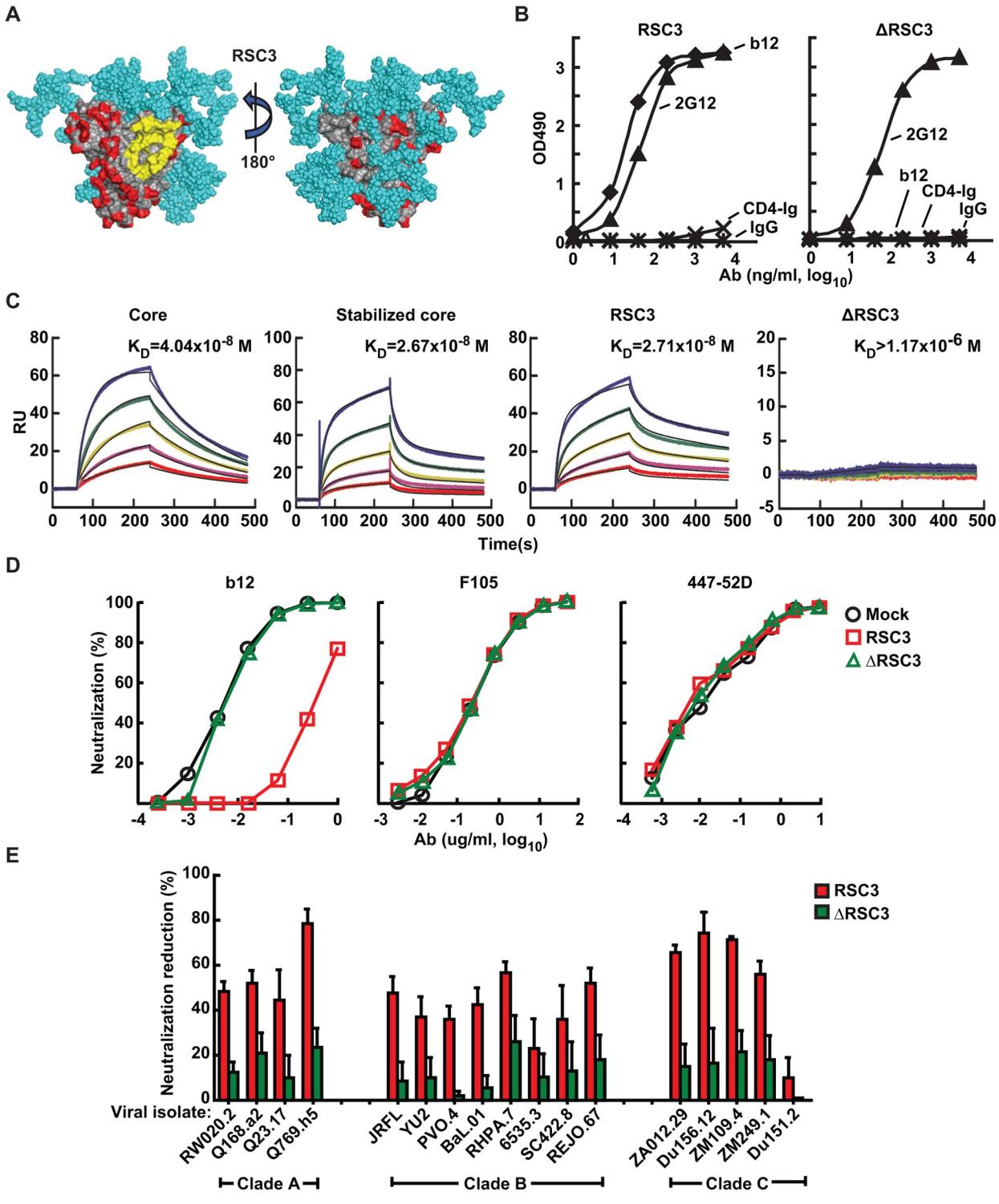
**Fig. 1.** Design and antigenic profile of RSC3 and analysis of epitope-specific neutralization. (A) Surface structure model of the RSC3. The outer domain contact site for CD4 is highlighted in yellow. Regions highlighted in red are antigenically resurfaced areas, shown both on the inner (left panel) and outer (right panel) faces of the core protein. Glycans are shown in light blue. (B) Antigenicity of the RSC3 protein based on enzyme-linked immunosorbent assay (ELISA) using the neutralizing CD4bs mAb b12 and CD4-Ig fusion protein. mAb 2G12 was used to confirm the structural integrity of the protein. (C) mAb b12 was immobilized on the sensor chip for surface plasmon resonance (SPR) kinetic binding analysis with the proteins shown. (D) RSC3 blockade of HIV-1 viral strain HXB2 neutralization by the broadly neutralizing CD4bs mAb b12, but not CD4bs mAb F105, which has limited neutralization breadth. The V3 neutralizing mAb 447-52D is shown as a control. (E) Analysis of serum 45 neutralization of a panel of 17 viruses, using RSC3 and ΔRSC3 to block neutralization activity. The percent reduction in the serum ID<sub>50</sub> caused by competition with RSC3 or ΔRSC3 is shown on the Y-axis (+/- SEM of three independent experiments). Viral strains and clades are shown on the X-axis. Values less than 20% are not considered significant in this assay.

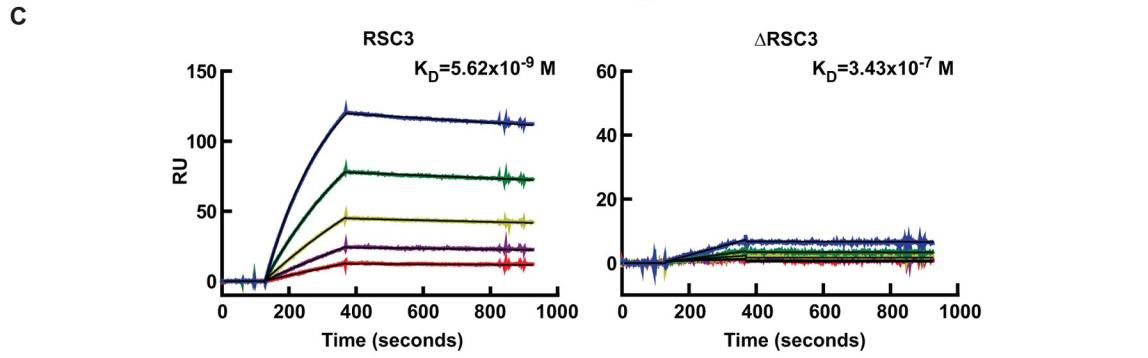
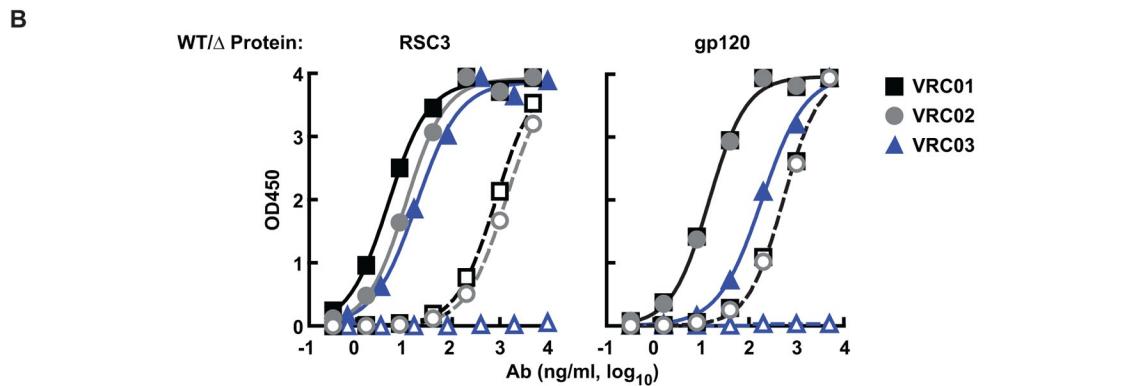
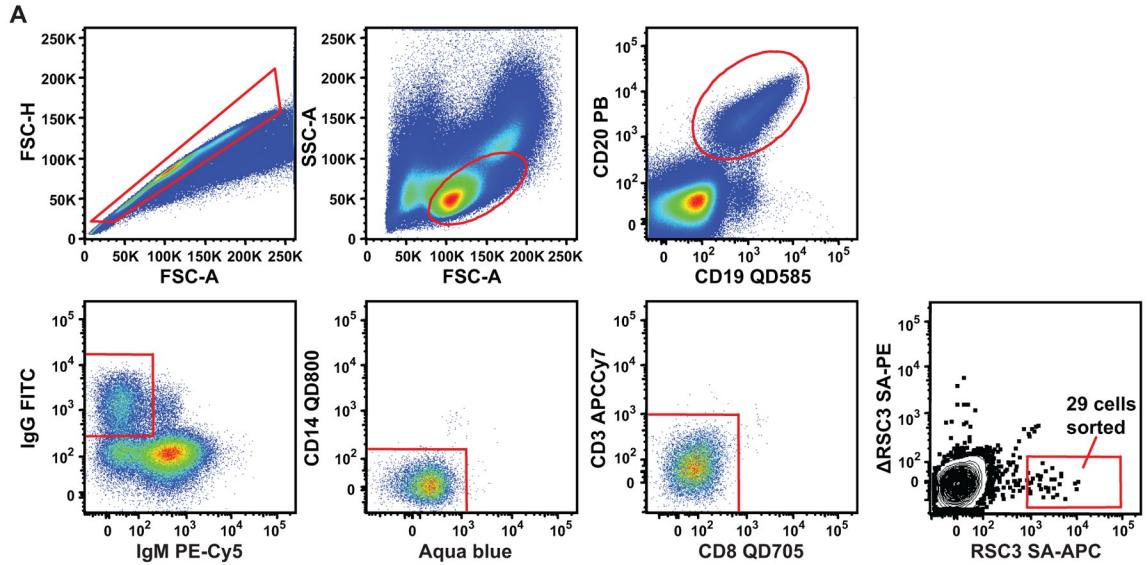
**Fig. 2.** Isolation of individual CD4bs-directed memory B cells by cell sorting, and binding characterization of isolated mAbs. (A) Twenty-five million PBMC from donor 45 were incubated with biotin-labeled RSC3 and ΔRSC3 that were complexed with SA-APC and SA-PE respectively, prior to addition to cells. Memory B cells were selected based on the presented gating strategy. Twenty-nine B cells that reacted with RSC3 and not ΔRSC3 (representing 0.05% of all memory B cells) were sorted into individual wells of a 96-well plate containing lysis buffer. (B) ELISA antigen binding profile of three isolated mAbs, VRC01, VRC02 and VRC03. Solid lines show mAb binding to RSC3 (left panel) and YU2

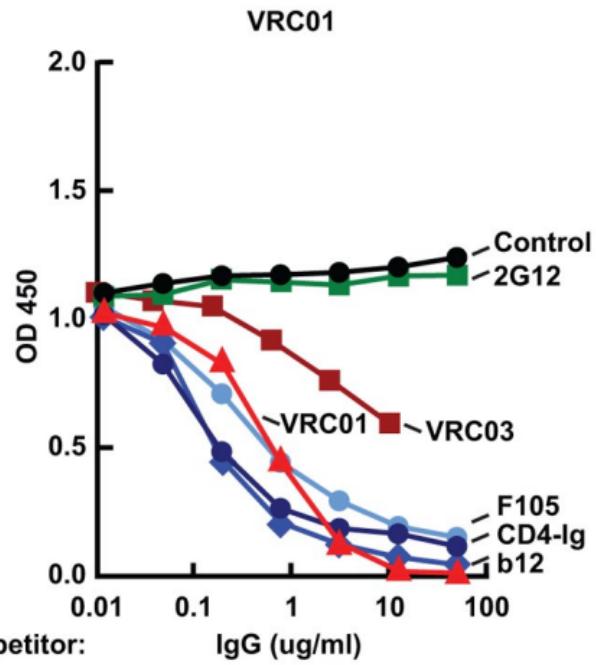
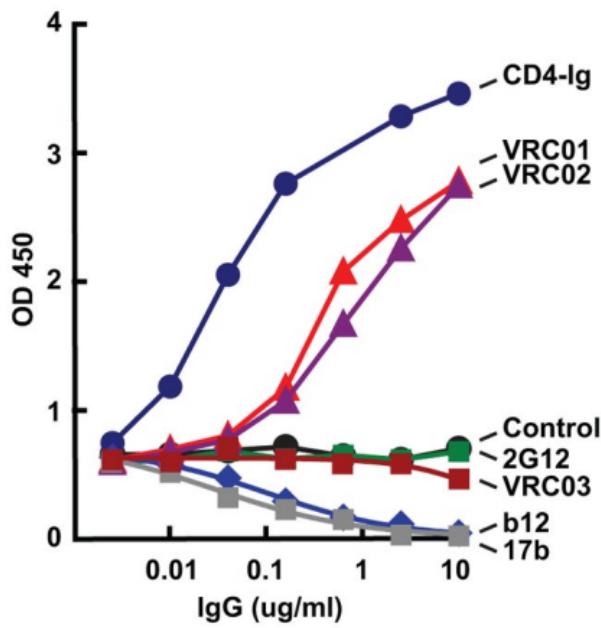
gp120 (right panel). Dashed lines indicate binding to ΔRSC3 (left) or to the CD4bs knockout mutant of gp120, D368R (right). (C) SPR binding analysis of VRC01 reacted with RSC3 and ΔRSC3. VRC01 was captured with an anti-human IgG-Fc antibody that was immobilized on the sensor chip. The SPR and ELISA data shown are from a representative experiment; several additional assays produced similar data.

**Fig. 3.** Antigenic and biophysical characterization of novel CD4bs-directed mAbs. (A) Competition ELISA performed with a single concentration of biotin-labeled VRC01 (left) or the co-receptor binding mAb 17b (right). The mAbs indicated near each line were titrated into the ELISA at increasing concentrations to evaluate the effect on VRC01 and 17b binding, respectively. The results shown are from a representative experiment; two additional assays produced similar data. (B) Isothermal titration calorimetry (ITC) to assess the change in enthalpy ( $\Delta H$ ) and entropy ( $-T\Delta S$ ) upon binding of mAbs to YU2 gp120. Each measured value is shown +/- SEM.

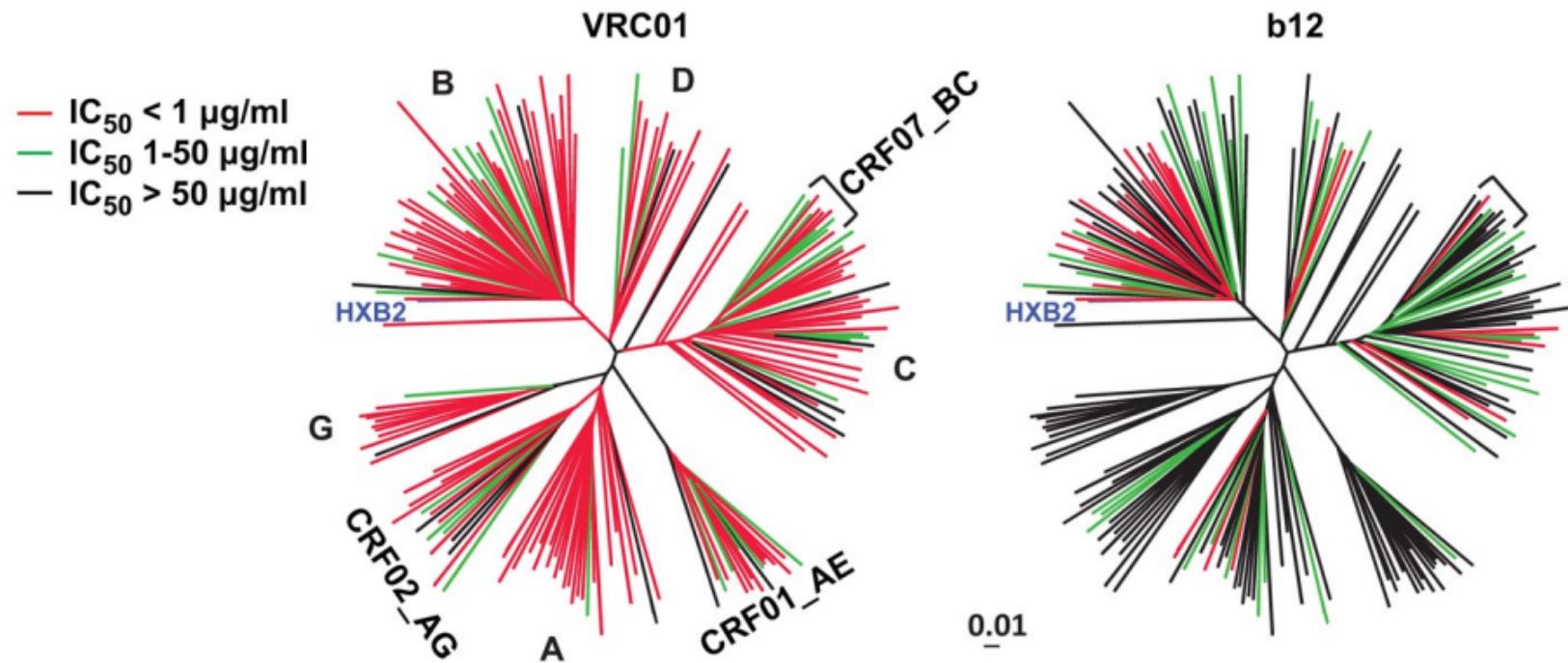
**Fig. 4.** Analysis of neutralization by mAbs VRC01 and b12 against a panel of 190 Env pseudoviruses representing all major circulating clades of HIV-1. Dendograms, made by the neighbor-joining method, show the protein distance of gp160 sequences from 190 HIV-1 primary isolates. The clade B reference strain HXB2 was used to root the tree, and the amino acid distance scale is indicated with a value of 1% distance as shown. The clades of HIV-1 main group, including circulating recombinant forms (CRFs), are indicated. Neutralization potency of VRC01 and b12 is indicated by the color of the branch for each virus. The data under the dendograms show the percent of viruses neutralized with an IC<sub>50</sub> < 50 µg/ml, and < 1 µg/ml, and the geometric mean IC<sub>50</sub> value for viruses neutralized with an IC<sub>50</sub> < 50 µg/ml.





**A****17b****B**

Antibody	$\Delta G$ (kcal/mol)	$\Delta H$ (kcal/mol)	$T\Delta S$ (kcal/mol)
VRC01	-9.87 +/- 0.18	-48.6 +/- 1.29	38.7 +/- 0.65
VRC03	-8.68 +/- 0.07	-8.47 +/- 0.23	-0.21 +/- 0.12
b12	-11.1 +/- 0.2	-17.0 +/- 1.7	5.9 +/- 1.7
CD4-Ig	-10.7 +/- 0.85	-50.7 +/- 1.38	40.0 +/- 0.81



Titer	VRC01	b12
IC <sub>50</sub> < 50 µg/ml	91%	41%
IC <sub>50</sub> < 1 µg/ml	72%	17%
Geometric mean IC <sub>50</sub> (µg/ml)	0.33	1.79