Higher tenofovir exposure is associated with longitudinal declines in kidney function in women living with HIV

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Objective: Tenofovir disoproxil fumarate is a commonly used antiretroviral drug, but risk factors for tenofovir (TFV)-associated kidney disease are not fully understood. We used intensive pharmacokinetic studies in a cohort of HIV-infected women on TFV-based therapy to study the relationship between TFV exposure and subsequent kidney function.

Design: This is a nested study within the Women's Interagency HIV Study, a multicenter, prospective cohort of HIV-infected women. Participants on TFV-based therapy underwent 24-h intensive pharmacokinetic sampling after witnessed dose. Kidney function was measured over the succeeding 7 years by serum creatinine [estimated glomerular filtration rate calculated by serum creatinine (eGFRcr)].

Methods: Multivariable linear mixed models evaluated the relationship of baseline TFV area under the-time concentration curves (AUCs) with subsequent changes in kidney function. Covariates included age, diabetes, hypertension, race, BMI, ritonavir use, duration of TFV exposure, current CD4⁺ cell count, and HIV viral load.

Results: Of the 105 participants, persons within the highest baseline TFV AUC tertile had significantly lower eGFRcr compared with those in the lowest tertile (mean \pm stanstandard error: 80 ± 4.3 vs. 104 ± 2.5 ml/min per 1.73 m², P<0.0001). By year 7, this difference widened (72 ± 4.9 vs. 105 ± 2.9 , P<0.0001). After multivariable adjustment, TFV AUC in the highest tertile remained associated with lower eGFRcr relative to values in the lowest tertile at both baseline (-15 ml/min per 1.73 m², P=0.0047) and year 7 (-23 ml/min per 1.73 m², P=0.0002).

Conclusion: Through intensive TFV pharmacokinetic sampling, we found a strong association between greater TFV exposure and subsequent decline in kidney function. Variations in TFV drug exposure may partially account for subsequent nephrotoxicity in persons infected with HIV. Copyright © 2016 Wolters Kluwer Health, Inc. All rights reserved.

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Introduction

Tenofovir disoproxil fumarate (TDF) is one of the most commonly used antiretroviral drugs for the treatment and prevention of HIV infection. Currently, TDF is a component in several one-pill once-a-day combination antiretroviral therapy (ART) formulations and is recommended for first line ART regimens by international guidelines [1]. As TDF formulations become increasingly available in global settings where the burden of HIV is highest, and as the HIV-infected population ages, it is vital to gain a better understanding of the factors that contribute to TDF toxicity.

Systemic exposure to drug is optimally characterized by pharmacokinetic methods, but prior pharmacokinetic studies of TDF have been limited to populations that differ in race, sex, and comorbidity from large populations of HIV patients [2]. Additionally, these prior pharmacokinetic studies have not examined the relationship between variations in TDF exposure and subsequent kidney function. Determination of kidney function following intensive pharmacokinetic analyses is an important means to determine whether TDF exposure is predictive of subsequent kidney impairment.

Although TDF is highly effective and well tolerated in most recipients, its use has been associated with reduced kidney function and proteinuria [3-8], adverse consequences of treatment that can lead to discontinuation of the drug, and which may not be reversible after treatment discontinuation [4,7,9]. Although formulated as TDF to promote oral absorption and tissue delivery, pharmacokinetic studies often measure tenofovir (TFV), the active form of the drug in plasma, or its' phosphorylated metabolites. For many drugs, the occurrence of adverse effects is directly related to systemic exposure to the drug [defined as areas under the time concentration curve (AUC)]. TFV, like the majority of drugs, demonstrates significant interindividual variability in plasma drug and trough levels that can be affected by a variety of clinical factors [10–14]. Understanding the pharmacokinetics of TFV is important for providing insight into whether the degree of drug exposure is associated with toxicity. Interindividual variability in TFV exposure has been hypothesized to be a determinate of kidney injury, but studies to date examining this relationship have been limited by small sample sizes and short duration of observations, by study populations with limited generalizability, and use of surrogate assessments of TFV pharmacokinetics [15,16].

The primary objective of this study was to determine if TFV exposure, as measured by TFV AUCs is associated with kidney function over a long period of observation. Our hypothesis was that TFV exposure would be independently associated with a decline in kidney function over time.

Methods

Study design and population

The Women's Interagency HIV Study (WIHS) is a large, multicenter, prospective cohort study of HIV-infected women and at-risk HIV-uninfected women in the United States [17], ongoing since 1993. The WIHS is representative of US women living with HIV in terms of demographic and clinical parameters. We previously described the 'WIHS Intensive PK Study' [18,19], which enrolled 480 HIV-infected women on varying ART regimens. This intensive pharmacokinetic assessment, conducted from 2004 to 2008, included 12-24-h sampling of antiretroviral plasma levels after administration of a dose witnessed by study team members. Participant data were included in this analysis for women who reported steady-state use of TDF (i.e. TDF use for at least 6 months prior to pharmacokinetic evaluation), underwent 24-h intensive pharmacokinetic sampling for plasma TFV levels, and had at least one estimated glomerular filtration rate calculated by serum creatinine (eGFRcr) from a WIHS visit after the pharmacokinetic study. Participants were not excluded for ongoing recreational drug use. Laboratory studies, physical examinations, demographic information, self-reported ARV adherence, and other characteristics were obtained every 6 months on WIHS participants as part of participation in the overall prospective cohort study. Institutional review boards at all participating institutions approved the study and informed consent materials. Written informed consent for the pharmacokinetic study was obtained from each study participant. Participants were continued to be followed (and included in this study) even if they discontinued TDF, and they were only censored at loss-to-follow-up.

Intensive pharmacokinetic protocol methods

Pharmacokinetic protocols were conducted in clinical research centers or other facilities associated with collaborating WIHS sites. The TFV measurement procedure has been previously described [12], but important details are included here. Participant's typical diet was determined via phone interview 2-3 days prior to the pharmacokinetic study. That diet was then provided during the pharmacokinetic protocol. Patients used their routine medications during the protocol, and these were recorded by study staff. Plasma samples were drawn at timed intervals over 24 h for drug levels under conditions of actual use, including providing the participants' home medications, and processed within 30 min of sample collection. All participants received their standard dose of TDF and TFV levels were measured in plasma specimens collected at 0, 4, 8, 15, 18, and 24 h after witnessed dose. Participants were seen for the pharmacokinetic visit within 6 weeks of a core WIHS cohort visit, thus temporally linking measures between the WIHS data and pharmacokinetic data. Weight and medication use were collected at both the intensive pharmacokinetic and core WIHS visit, whereas data on comorbidities, HIV RNA level, CD4⁺ cell counts, and serum creatinine were collected during the core WIHS visits.

Laboratory procedures

Plasma levels of TFV were determined by liquid chromatography/tandem mass spectrometry with TDF-d6 as the internal standard [20]. The plasma sample was pretreated with trifluoroacetic acid for protein precipitation before injecting into the Micromass Quattro Ultima (Waters Corporation, Milford Massachusetts, USA) liquid chromatography/tandem mass spectrometry system as previously described [12,20]. The assay has been validated from 10 to 1000 ng/ml of TFV with a coefficient of variation less than 15% for quality control samples at low, medium, and high concentrations.

Covariates used for adjustment

As in our prior analysis [12], the following baseline covariates were included in all multivariable models: age, race/ethnicity, BMI, diagnosis of diabetes or hypertension, self-reported prior duration of TDF usage, concurrent ritonavir use, current CD4⁺ cell count (log transformed), and concurrent HIV viral load (detectable vs. nondetectable, defined as above or below assay thresholds of 120, 80 or 50 copies/mL, depending on assay used by WIHS at the time of the relevant visit).

Primary predictor

AUCs were used to estimate TDF exposure over the 24-h dosing interval (once at baseline); these were calculated for each individual using the trapezoidal rule [21]. For calculation of the AUC, any observations with TFV concentrations below the lower limit of quantification (10 ng/ml) were replaced by 0 ng/ml; 10 individuals had undetectable TFV at baseline at the beginning of the intensive pharmacokinetic study. One individual had a single value that was undetectable at a time other than baseline.

Outcome

The Chronic Kidney Disease Epidemiology Collaboration equation using serum creatinine was used to estimate glomerular filtration rate (eGFRcr in ml/min per 1.73 m²) [12,22]. We analyzed changes in eGFRcr as a continuous outcome, expressed as annual differences in ml/min per 1.73 m²/year over approximately 7 years of WIHS follow-up.

Statistical analysis

We stratified participants into three categories based on tertile of TFV AUC, and compared demographic and baseline clinical characteristics using the χ^2 and Kruskal–Wallis tests for categorical and continuous variables, respectively. Kernel density estimates were used to construct smoothed density curves to examine

distributions of baseline eGFRcr levels, and were compared by tertile of TFV AUC using Levene's test for homogeneity of variance. Multivariable linear mixed effect models were used to evaluate the relationship of baseline TFV AUC with subsequent change in kidney function, with random intercepts and slopes using an unstructured variance-covariance matrix for modeling. Graphical examination of the trajectories found curvilinear changes in eGFRcr. Therefore, we modeled eGFRcr using linear splines, with potentially different slopes during the earlier and later follow-up intervals (0-3.5 and 3.5-7 years). We analyzed tertiles of TFV AUC with tertile 1 (lowest AUC) as the reference category for all analyses. We also evaluated levels of TFV AUC (above vs. below median) and baseline eGFRcr (<90 vs. >90 ml/min/1.73 m²) jointly, to determine whether the association of TFV AUC with subsequent change in kidney function varied by level of baseline eGFRcr.

We used relative risk regression with a modified Poisson approach to examine associations of TFV AUC with incident chronic kidney disease (CKD) [23]. We analyzed TFV AUC both continuously (per 10% increase) and categorically (using tertiles). We defined incident CKD using eGFR by creatinine, calculated using the Chronic Kidney Disease Epidemiology Collaboration equation as eGFR less than 50, 60, or 70 at any two consecutive visits over approximately 7 years after baseline. All analyses were conducted using SAS (version 9.4, Cary, North Carolina, USA).

Results

The baseline demographic and clinical characteristics of participants in the study, stratified by TFV AUC tertile, are summarized in Table 1. A total of 117 women underwent intensive pharmacokinetic sampling for TFV, but 12 women were excluded from analysis because of missing data (including no eGFRcr) or lack of subsequent measures. The recorded dose for TDF was 300 mg once daily in all of the participants' ART regimens. Among the 105 women who contributed data to this analysis, the mean age was 43 years (range 22-65), 68% were African-American, 13% white, and 20% of other race; 25 (24%) self-identified as Hispanic ethnicity. Women with TFV AUC in the highest tertile were older and had lower median BMI relative to those with TFV AUC in the lower tertiles. Concurrent CD4⁺ cell counts were lower and detectable HIV RNA was more prevalent among women with TFV AUC in the highest tertile than those with lower AUCs, but these differences were not statistically significant.

We examined the distribution of eGFRcr at baseline, stratified by TFV AUC tertile (tertile 1 has the lowest AUC, tertile 2 had intermediate AUC values, and tertile 3

| Parameter | TFV AUC tertile 1 ($n = 35$) | TFV AUC tertile 2 $(n=35)$ | TFV AUC tertile 3 ($n = 35$) | P value |
|---|--------------------------------|----------------------------|--------------------------------|----------|
| TFV AUC range (ng*h/ml) | 1031-2640 | 2646-3922 | 4009-13 911 | |
| Age (years) | 41 (31, 47) | 42 (34, 49) | 47 (44, 51) | 0.0004 |
| Race | . , , | , , , | , , , | 0.32 |
| African-American | 22 (63%) | 27 (77%) | 22 (63%) | |
| White | 7 (20%) | 3 (9%) | 3 (9%) | |
| Other | 6 (17%) | 5 (14%) | 10 (29%) | |
| Hispanic | 11 (31%) | 3 (9%) | 11 (31%) | 0.029 |
| $BMI (kg/m^2)$ | 28 (25, 36) | 29 (26, 33) | 25 (22, 30) | 0.036 |
| Diabetes | 5 (14%) | 11 (31%) | 11 (31%) | 0.20 |
| Hypertension | 12 (34%) | 12 (34%) | 14 (40%) | 0.90 |
| Ritonavir use | 13 (37%) | 26 (74%) | 24 (69%) | 0.0040 |
| Duration of prior TDF exposure (years) | 1.5 (1.0, 2.0) | 1.0 (0.5, 1.5) | 1.5 (0.5, 2.0) | 0.071 |
| Current CD4 ⁺ cell count (cells/µl) | 412 (240, 593) | 374 (263, 644) | 306 (200, 502) | 0.097 |
| Nadir CD4 ⁺ cell count (cells/µl) | 188 (119, 318) | 214 (108, 297) | 159 (25, 211) | 0.099 |
| Detectable HIV viral load | 12 (34%) | 9 (26%) | 16 (46%) | 0.24 |
| Baseline eGFRcr (ml/min per 1.73 m ²) | 104 (93, 122) | 97 (85, 108) | 78 (61, 104) | < 0.0001 |
| Years of eGERcr follow-up | 7.3 (6.4. 9.1) | 7.7 (7.3, 8.0) | 7.0 (4.0, 7.8) | 0.014 |

Table 1. Baseline characteristics of study participants by tertile of tenofovir disoproxil fumarate area under the time concentration curve (n = 105 total).

Continuous parameters are presented as median (IQR) and categorical parameters as n (%). AUC, area under the time concentration curve; eGFRcr, estimated glomerular filtration rate by creatinine; IQR, interquartile range; TDF, tenofovir disoproxil fumarate; TFV, tenofovir.

had the highest TFV AUC values) (Fig. 1). The median eGFR cr at baseline was $104 \,\mathrm{ml/min}$ per $1.73 \,\mathrm{m}^2$ in tertile 1, 97 ml/min per $1.73 \,\mathrm{m}^2$ in tertile 2, and 78 ml/min per $1.73 \,\mathrm{m}^2$ in tertile 3 of TFV AUC. The spread of eGFR cr levels was considerably wider among participants with exposures in tertile 3 compared with those in tertiles 1 and 2 (P = 0.0021 by test for homogeneity of variance).

We next examined trajectories of change in eGFR.cr over time by TFV AUC tertile (Fig. 2). Mean eGFR.cr was lowest among those with highest TFV AUC at baseline (mean ml/min/1.73 m² \pm standard error: 80 \pm 4.3 vs. 104 \pm 2.5 for highest vs. lowest tertile, P < 0.0001), and these differences widened over time (year 7: 72 \pm 4.9 vs.

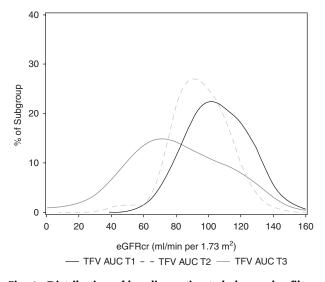


Fig. 1. Distribution of baseline estimated glomerular filtration rate calculated by serum creatinine (ml/min per 1.73 m²) in HIV-infected women, stratified by baseline tenofovir area-under-the-time-concentration-curve tertile.

 105 ± 2.9 , P < 0.0001). The trajectories of eGFRcr among women in tertiles 1 and 2 showed similar curvilinear changes, with increases in eGFRcr apparent during years 0-3.5, followed by decreases in years 3.5-7. By contrast, women with TFV AUC in tertile 3 showed decreases in eGFRcr throughout the entire follow-up period. Analyses confirmed that tertile 3 (during years 0-3.5) had significantly larger declines in eGFRcr than tertile 1 (-1.87 ml/min per 1.73 m²/year, P = 0.029; Table 2), whereas there was no statistically significant difference between tertiles 1 and 2. This more pronounced decline in eGFRcr for women whose TFV AUC were in tertile 3 remained statistically significant after multivariable adjustment for all cofactors. During the later period (years 3.5–7) significant decreases in eGFR cr occurred for women with TFV AUC values in all three tertiles, ranging from 1.4 to 2.4 ml/min per 1.73 $m^2/year$ (all P < 0.01), and no statistically significant differences were detected across tertiles. Supplemental Table 1, http://links.lww.com/QAD/A829 shows eGFR cr estimates from multivariate linear mixed models using piecewise linear spline, separated yearly.

We then restricted the analysis to include only those participants who had detectable TFV at the start of the study (N=95), which resulted in nine individuals being removed from tertile 1 and one individual being removed from tertile 3. Results were similar after excluding those with undetectable TFV, in that tertile 1 showed a 2.1 ml/min per 1.73 m² [95% confidence interval (CI) 0.92-3.4 ml/min per 1.73 m²] average annual increase in eGFR over years 0-3.5, and a 1.5 ml/min per 1.73 m²) average annual decline in eGFR over years 3.5-7. In multivariable adjusted analysis, over years 0-3.5, both tertiles 2 and 3 had smaller gains on average relative to tertile 1. Being in tertile 2 was associated with

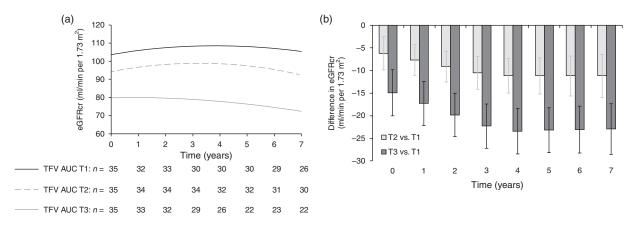


Fig. 2. (a) Trajectory of eGFRcr (ml/min per 1.73 m²) in HIV-infected women over time (unadjusted linear mixed model), showing mean. (b) This panel shows adjusted trajectory of eGFRcr by tertile of TFV AUC with estimates from a linear mixed model with piecewise linear spline, where T1, TFV AUC tertile 1; T2, TFV AUC tertile 2, and T3, TFV AUC tertile 3. AUC, area-under-the-time-concentration-curve; eGFRcr, estimated glomerular filtration rate calculated by serum creatinine; TFV, tenofovir.

a $2.3 \,\mathrm{ml/min}$ per $1.73 \,\mathrm{m}^2$ difference (95% CI -4.2 to -0.45 ml/min per 1.73 m²) and being in tertile 3 was associated with a 3.2 ml/min/1.73 m² difference (95% CI -5.0 to -1.38 ml/min per 1.73 m²), relative to tertile 1. Over years 3.5-7, differences in annual change narrowed and were no longer statistically different between tertiles. We also evaluated the association of TFV AUC with incident CKD using various eGFRcr $(50, 60, \text{ or } 70 \text{ ml/min}/1.73 \text{ m}^2)$ thresholds (Supplemental Table 2, http://links.lww.com/QAD/A829). In unadjusted analysis, each 10% increase in the TFV AUC was associated with a 37% increased risk of eGFR less than 50 ml/min per 1.73 m², a 30% increased risk of eGFR less than 60 ml/min per 1.73 m², and an 18% increased risk of eGFR less than 70 ml/min per 1.73 m² (all P < .05). Although we had reduced power because of the small numbers of incident cases, we found that greater TFV AUC remained associated with a 22% increased risk of eGFR less than 70 ml/min per 1.73 m² after multivariable adjustment. When defined as eGFR less than 78 ml/min per 1.73 m², rates of CKD ranged

from 5.7% in tertile 1 to 29% in tertile 3, resulting in a five-fold increased risk of CKD for those in tertile 3 even after multivariable adjustment. Finally, we examined the effect of stratifying by both TFV AUC and baseline eGFRcr, comparing trajectories of eGFRcr change over time (Fig. 3). We first considered persons with eGFRcr less than 90 ml/min per 1.73 m² at baseline (Fig. 3a); the eGFRcr trajectory appeared parallel in the lower (n = 15) and higher (n = 31) TFV AUC groups although the absolute level of eGFRcr was consistently lower in the higher TFV AUC group. In persons with eGFR cr more than 90 ml/min per 1.73 m² at baseline (Fig. 3b), the initial eGFRcr was similar in those with lower (n = 37) and higher (n = 22) TFV AUC, but those with higher TFV AUC experienced a greater decline in eGFR cr (P = 0.025 for years 0 - 3.5and P = 0.71 for years 3.5-7). Supplemental Table 3, http://links.lww.com/QAD/A829 shows eGFRcr estimates from multivariate linear mixed models using piecewise linear spline, separated yearly and stratified by both baseline eGFRcr and TFV AUC.

Table 2. Association of tenofovir area under the time concentration curve with annual change in estimated glomerular filtration rate by creatinine (ml/min/1.73 m²) stratified by study follow-up interval (first 3.5 and second 3.5 years).

| Parameter time period | TFV AUC tertile 1 | TFV AUC tertile 2 | TFV AUC tertile 3 | | |
|---|---------------------------------|--------------------------------|---------------------------------|--|--|
| Annual change in eGFR (95% CI): | | | | | |
| Year 0-3.5 | 1.63 (0.51, 2.7), P = 0.0044 | 1.68 (0.34, 3.0), P = 0.014 | -0.24 (-1.49, 1.02), P = 0.71 | | |
| Year 3.5-7.0 | -1.40 (-2.4, -0.45), P = 0.0044 | -2.4 (-3.7, -1.12), P = 0.0003 | -1.92 (-3.2, -0.60), P = 0.0051 | | |
| Unadjusted difference in annual change vs. tertile 1 (95% CI): | | | | | |
| Year 0-3.5 | Reference | 0.048 (-1.69, 1.78), P = 0.96 | -1.87 (-3.5, -0.20), P = 0.029 | | |
| Year 3.5-7.0 | Reference | -0.99 (-2.6, 0.58), P = 0.22 | -0.52 (-2.1, 1.10), P = 0.53 | | |
| Adjusted ^a difference in annual change vs. tertile 1 (95% CI): | | | | | |
| Year 0-3.5 | Reference | -1.41 (-3.2, 0.37), P = 0.12 | -2.5 (-4.2, -0.76), P = 0.0046 | | |
| Year 3.5–7.0 | Reference | 0.0079 (-1.61, 1.63), P = 0.99 | 0.17 (-1.56, 1.89), P = 0.85 | | |

Estimates from linear mixed models. AUC, area-under-the-time-concentration-curve; T1, TFV AUC tertile 1; T2, TFV AUC tertile 2; T3, TFV AUC tertile 3; TFV, tenofovir.

^aCovariates in adjusted model include: age, race, BMI, diabetes mellitus, hypertension, ritonavir use, duration of prior tenofovir exposure, CD4⁺ cell count and HIV viral load.

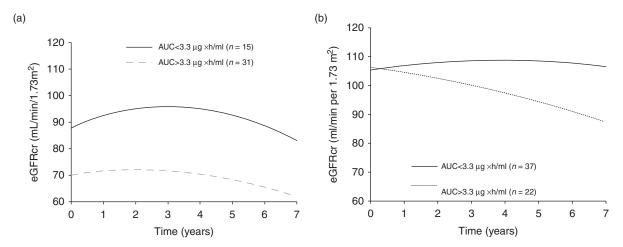


Fig. 3. (a) Trajectory of eGFRcr (ml/min per 1.73 m²) over time in patients with baseline GFRcr less than 90 ml/min per 1.73 m². (b) Baseline eGFRcr more than 90 ml/min per 1.73 m² stratified into groups of low TFV AUC (<3.3 μ g \times h/ml) vs. high TFV AUC (>3.3 μ g \times h/ml). AUC, area-under-the-time-concentration-curve; eGFRcr, estimated glomerular filtration rate calculated by serum creatinine; TFV, tenofovir.

Discussion

This is the first report to demonstrate that greater TFV exposure, measured by intensive 24-h pharmacokinetic assessment, is strongly associated with more rapid declines in kidney function over time. We observed a significant decline in kidney function over the first 3.5 years of follow-up, which did not normalize by year 7. Higher TFV exposure remained independently associated with more impaired kidney function longitudinally, even after controlling for duration of prior TFV exposure, baseline kidney function, and other relevant risk factors that may influence kidney function. The association between eGFRcr estimated at the time of intensive pharmacokinetic and TFV exposure as represented by AUCs in this study could be attributable to the effect of reduced kidney function on TFV clearance or the effect of higher TFV concentrations on kidney function. However, the longitudinal association of higher TFV AUC with a more rapid subsequent decline in kidney function provides strong evidence that greater exposure to TFV can injure the kidney over time.

These data indicate that the extent of TFV exposure may influence the occurrence of kidney injury, which may have implications for dosing and prescribing strategies. Our group and others have identified clinical factors that increase TFV AUC in intensive pharmacokinetic studies, including concomitant ritonavir use, pre-TDF impairment of eGFRcr, lower body weight, and increasing age [12]. Some have advocated for modifications in TDF dosing, particularly with the use of specific protease inhibitors [24]; given the results of our study, such alternative dosing regimens should be further explored to understand whether they may alleviate subsequent kidney toxicity for some individuals on this commonly used antiretroviral agent. We need a

better understanding of the factors that determine the pharmacokinetics and pharmacodynamics of TFV and the extent to which these are modified by changes in dosing. Modification to dosing must be evaluated while also ensuring that control of HIV replication is maintained. Our study has shown that there is tremendous between-person variability in exposure to TFV, despite fixed dose regimens, a fact that is important in understanding TDF's potential toxicity. Determining TFV AUC in the clinical setting may be impractical, but identifying other strategies to predict toxicity should be explored in future studies. One possibility is the use of single trough levels that have been correlated with total daily TFV exposure.

In persons at risk for progression of kidney disease in the setting of TDF use, other agents can be considered as part of the backbone ART regimen in lieu of TDF. For example, abacavir may lead to greater improvements in albuminuria and proteinuria in persons taking ART compared with those on TDF-based regimens [25]. Other therapeutic approaches may lead to lower rates of kidney injury because of lower systemic exposure to the TDF moiety. One such opportunity could be tenofovir alafenamide fumarate (TAF), a novel prodrug of TFV that has been reported as having the potential benefits of lower kidney and bone toxicity without a compromise in treatment efficacy [26–30]. Pharmacokinetic studies demonstrated that a TAF dose of 25 mg (compared with 300 mg of TDF) resulted in an approximate 86% reduction in systemic exposure and a seven-fold increase in the mean concentration of intracellular peripheral blood mononuclear cell TFV diphosphate [31]. In addition, TFV from TDF is renally secreted via organic anion transporters (OAT1 and OAT3), whereas TAF has not been shown to significantly interact with OAT or result in OAT-

dependent cytotoxicity *in vitro* [29]. However, it is unknown whether TAF treatment will lead to lower kidney tubular cell levels of TFV than TDF since TFV is a substrate for OAT. This idea has been suggested given the lower systemic exposure to TFV seen with TAF and clinical trials demonstrating lower levels of kidney injury markers for patients using TAF relative to TDF [26–28,31]. If it is felt that the mechanism of kidney tubular cell injury is attributable to mitochondrial injury, as supported by in-vitro studies [32–36], then intracellular concentrations of drug would then seem to be highly relevant. Longitudinal data in real-world settings will be required to determine whether TAF will be a reasonable therapeutic option for persons on TDF who develop kidney or bone toxicity.

There were several strengths to this study. First, this is the largest study to perform intensive pharmacokinetic measurements in patients on TDF-based regimens. We were able to follow these participants for an average of more than 7 years after completion of exposure determination, allowing a far more comprehensive assessment of the potential kidney complications of higher levels of TFV exposure. These study results were obtained from an ethnically and clinically diverse cohort comprised mostly of racial/ethnic minority women – a group that has been traditionally underrepresented in HIV research [37–39].

There were also some limitations to the interpretation of these data. An important assumption underlying our analysis was that TFV AUC does not change over time. This may not be true, and in the setting of declining eGFRcr, it is possible that TFV AUC may be gradually increasing; this could contribute to further reduction of kidney function in a vicious self-perpetuating cycle. In fact, our data support this, given the cross-sectional associations of lower eGFRcr with higher TFV exposure at baseline. It is, therefore, possible that our calculations of the relationship between eGFRcr and TFV AUC are underestimated. Also, we only measured glomerular function, and not tubular function. TFV-associated kidney disease may be a result of tubular injury [40,41] but the impact on eGFRcr over longer periods of exposure is not clear. This specific cohort does not yet have serial measurements of biomarkers of tubular injury such as IL-18, kidney injury molecule-1, or α -1 microglobulin. More specific and early markers of kidney toxicity such as these would enable better determination of the exact mechanisms underlying TFV-induced injury. We were not able to control for genetic factors that may play a role in determining TFV exposure and may be either confounding or modifying the relationship between TFV AUC and subsequent eGFRcr. A number of polymorphisms have been identified that result in increased intracellular TFV concentration or increased proximal tubular injury [42,43], but whether these factors or different ones are

relevant to the diverse population in this study is not well characterized. The study was limited to women, and therefore the results may pertain to women only. Nearly all the women in this study received coadministration of emtricitabine as part of their antiretroviral regimens and an impact of emtricitabine on the outcome cannot be definitely ruled out. Notably, a higher proportion of individuals in tertile 3 had less than 7 years of follow-up in the study; if they dropped out of the cohort because they were sicker or had worse kidney function, then our estimates for the decline in eGFRcr would be underestimated in this group. However, when we evaluated the association of TFV AUC with incident CKD using various eGFRcr (50, 60, or 70 ml/min per 1.73 m²) binary thresholds (Supplemental Table 2, http://links.lww.com/QAD/A829), higher TFV AUC remained associated with an increased risk of CKD.

In summary, we present compelling evidence suggesting that higher levels of exposure to TFV may partially account for subsequent declines in kidney function over time in a cohort of diverse women living with HIV. There is evidence for more rapid progression of kidney function decline in individuals with higher TFV exposure. There is also likely an effect of reduced kidney function leading to higher degrees of TFV exposure, which may operate in a cyclic feed-forward mechanism. These data support further studies exploring the factors that determine TFV pharmacokinetics as well as investigations regarding dosing reduction and novel treatment strategies to optimize drug exposure while minimizing drug toxicity in select high-risk populations.

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Conflicts of interest

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References

- World Health Organization. Consolidated guidelines on the use of antiretroviral drugs for treating and preventing HIV infection. Geneva, Switzerland: WHO; 30 June 2013. pp. 1–272.
- Barditch-Crovo P, Deeks SG, Collier A, Safrin S, Coakley DF, Miller M, et al. Phase i/ii trial of the pharmacokinetics, safety, and antiretroviral activity of tenofovir disoproxil fumarate in human immunodeficiency virus-infected adults. Antimicrob Agents Chemother 2001; 45:2733–2739.
- Calza L, Trapani F, Tedeschi S, Piergentili B, Manfredi R, Colangeli V, Viale P. Tenofovir-induced renal toxicity in 324 HIV-infected, antiretroviral-naïve patients. Scand J Infect Dis 2011; 43:656–660.
- Wever K, van Agtmael MA, Carr A. Incomplete reversibility of tenofovir-related renal toxicity in HIV-infected men. J Acquir Immune Defic Syndr 2010; 55:78–81.
- Monteagudo-Chu MO, Chang MH, Fung HB, Bräu N. Renal toxicity of long-term therapy with tenofovir in HIV-infected patients. J Pharm Pract 2012; 25:552–559.
- Hall AM, Hendry BM, Nitsch D, Connolly JO. Tenofovir-associated kidney toxicity in HIV-infected patients: a review of the evidence. Am J Kidney Dis 2011; 57:773–780.
- Scherzer R, Estrella M, Li Y, Choi AI, Deeks SG, Grunfeld C, et al. Association of tenofovir exposure with kidney disease risk in HIV infection. AIDS 2012; 26:867–875.
- Ryom L, Mocroft A, Kirk O, Worm SW, Kamara DA, Reiss P, et al.
 Association between antiretroviral exposure and renal impairment among HIV-positive persons with normal baseline renal function: the D:A:D study. J Infect Dis 2013; 207:1359–1369.
- Jose S, Hamzah L, Campbell LJ, Hill T, Fisher M, Leen C, et al. Incomplete reversibility of estimated glomerular filtration rate decline following tenofovir disoproxil fumarate exposure. J Infect Dis 2014; 210:363–373.
- Gagnieu MC, Barkil ME, Livrozet JM, Cotte L, Miailhes P, Boibieux A, et al. Population pharmacokinetics of tenofovir in AIDS patients. J Clin Pharmacol 2008; 48:1282–1288.
- Baheti G, Kiser JJ, Havens PL, Fletcher CV. Plasma and intracellular population pharmacokinetic analysis of tenofovir in HIV-1-infected patients. Antimicrob Agents Chemother 2011; 55:5294–5299.
- Baxi SM, Greenblatt RM, Bacchetti P, Scherzer R, Minkoff H, Huang Y, et al. Common clinical conditions – age, low BMI, ritonavir use, mild renal impairment – affect tenofovir pharmacokinetics in a large cohort of HIV-infected women. AIDS 2014; 28:59–66.
- 13. Gervasoni C, Meraviglia P, Landonio S, Baldelli S, Fucile S, Castagnoli L, et al. Low body weight in females is a risk factor for increased tenofovir exposure and drug-related adverse events. PLoS One 2013; 8:e80242.
- Le MP, Landman R, Koulla-Shiro S, Charpentier C, Sow PS, Diallo MB, et al. Tenofovir plasma concentrations related to estimated glomerular filtration rate changes in first-line regimens in African HIV-infected patients: ANRS 12115 DAYANA substudy. J Antimicrob Chemother 2015; 70:1517–1521.
- Rodriguez-Novoa S, Labarga P, D'Avolio A, Barreiro P, Albalate M, Vispo E, et al. Impairment in kidney tubular function in patients receiving tenofovir is associated with higher tenofovir plasma concentrations. AIDS 2010; 24:1064–1066.
- Poizot-Martin I, Solas C, Allemand J, Obry-Roguet V, Pradel V, Bregigeon S, et al. Renal impairment in patients receiving a tenofovir-cART regimen: impact of tenofovir trough concentration. J Acquir Immune Defic Syndr 2013; 62:375–380.
- Barkan SE, Melnick SL, Preston-Martin S, Weber K, Kalish LA, Miotti P, et al. The Women's Interagency HIV Study. WIHS Collaborative Study Group. Epidemiology 1998; 9:117–125.
- Gandhi M, Benet LZ, Bacchetti P, Kalinowski A, Anastos K, Wolfe AR, et al. Nonnucleoside reverse transcriptase inhibitor pharmacokinetics in a large unselected cohort of HIV-infected women. J Acquir Immune Defic Syndr 2009; 50:482–491.
- Gandhi M, Greenblatt RM, Bacchetti P, Jin C, Huang Y, Anastos K, et al. A single-nucleotide polymorphism in CYP2B6 leads to >3-fold increases in efavirenz concentrations in plasma and hair among HIV-infected women. J Infect Dis 2012; 206:1453–1461.
- Delahunty T, Bushman L, Robbins B, Fletcher CV. The simultaneous assay of tenofovir and emtricitabine in plasma using LC/MS/MS and isotopically labeled internal standards. J Chromatogr B Analyt Technol Biomed Life Sci 2009; 877:1907–1914.

- 21. Yeh KC, Kwan KC. A comparison of numerical integrating algorithms by trapezoidal, Lagrange, and spline approximation. J Pharmacokinet Biopharm 1978; 6:79–98.
- Gagneux-Brunon A, Delanaye P, Maillard N, Fresard A, Basset T, Alamartine E, et al. Performance of creatinine and cystatin C-based glomerular filtration rate estimating equations in a European HIV-positive cohort. AIDS 2013; 27:1573–1581.
- Zou G. A modified poisson regression approach to prospective studies with binary data. Am J Epidemiol 2004; 159:702–706.
- Hill A, Khoo S, Back D, Pozniak Á, Boffito M. Should the dose of tenofovir be reduced to 200-250 mg/day, when combined with protease inhibitors? J Int AIDS Soc 2014; 17:19583.
- Wyatt CM, Kitch D, Gupta SK, Tierney C, Daar ES, Sax PE, et al. Changes in proteinuria and albuminuria with initiation of antiretroviral therapy: data from a randomized trial comparing tenofovir disoproxil fumarate/emtricitabine versus abacavir/ lamivudine. J Acquir Immune Defic Syndr 2014; 67:36–44.
- Sax PE, Wohl D, Yin MT, Post F, DeJesus E, Saag M, et al. Tenofovir alafenamide versus tenofovir disoproxil fumarate, coformulated with elvitegravir, cobicistat, and emtricitabine, for initial treatment of HIV-1 infection: two randomised, double-blind, phase 3, noninferiority trials. Lancet 2015; 385:2606-2615.
- Sax PE, Zolopa A, Brar I, Elion R, Ortiz R, Post F, et al. Tenofovir alafenamide vs. tenofovir disoproxil fumarate in single tablet regimens for initial HIV-1 therapy: a randomized phase 2 study. J Acquir Immune Defic Syndr 2014; 67:52–58.
- Markowitz M, Zolopa A, Squires K, Ruane P, Coakley D, Kearney B, et al. Phase I/II study of the pharmacokinetics, safety and antiretroviral activity of tenofovir alafenamide, a new prodrug of the HIV reverse transcriptase inhibitor tenofovir, in HIV-infected adults. J Antimicrob Chemother 2014; 69:1362–1369.
- Bam RA, Yant SR, Cihlar T. Tenofovir alafenamide is not a substrate for renal organic anion transporters (OATs) and does not exhibit OAT-dependent cytotoxicity. Antivir Ther 2014; 19:687–692.
- Mills A, Crofoot G Jr, McDonald C, Shalit P, Flamm JA, Gathe J, et al. Tenofovir alafenamide versus tenofovir disoproxil fumarate in the first protease inhibitor-based single tablet regimen for initial HIV-1 therapy: a randomized phase 2 study. J Acquir Immune Defic Syndr 2015; 69:439–445.
- Immune Defic Syndr 2015; 69:439–445.
 Ruane PJ, DeJesus E, Berger D, Markowitz M, Bredeek UF, Callebaut C, et al. Antiviral activity, safety, and pharmacokinetics/pharmacodynamics of tenofovir alafenamide as 10-day monotherapy in HIV-1-positive adults. J Acquir Immune Defic Syndr 2013; 63:449–455.

- Abraham P, Ramamoorthy H, Isaac B. Depletion of the cellular antioxidant system contributes to tenofovir disoproxil fumarate-induced mitochondrial damage and increased oxidonitrosative stress in the kidney. J Biomed Sci 2013; 20:61.
- Kohler JJ, Hosseini SH, Hoying-Brandt A, Green E, Johnson DM, Russ R, et al. Tenofovir renal toxicity targets mitochondria of renal proximal tubules. Lab Invest 2009; 89:513–519.
- Kohler JJ, Hosseini SH. Subcellular renal proximal tubular mitochondrial toxicity with tenofovir treatment. Methods Mol Biol 2011; 755:267–277.
- Lebrecht D, Venhoff AC, Kirschner J, Wiech T, Venhoff N, Walker UA. Mitochondrial tubulopathy in tenofovir disoproxil fumarate-treated rats. J Acquir Immune Defic Syndr 2009; 51:258–263.
- Ramamoorthy H, Abraham P, Isaac B. Mitochondrial dysfunction and electron transport chain complex defect in a rat model of tenofovir disoproxil fumarate nephrotoxicity. J Biochem Mol Toxicol 2014; 28:246–255.
- Falcon R, Bridge DA, Currier J, Squires K, Hagins D, Schaible D, et al. Recruitment and retention of diverse populations in antiretroviral clinical trials: practical applications from the gender, race and clinical experience study. J Womens Health (Larchmt) 2011; 20:1043–1050.
- 38. Loutfy MR, Sherr L, Sonnenberg-Schwan U, Walmsley SL, Johnson M, d'Arminio Monforte A, et al. Caring for women living with HIV: gaps in the evidence. J Int AIDS Soc 2013; 16:18509
- d'Arminio Monforte A, Gonzalez L, Haberl A, Sherr L, Ssanyu-Sseruma W, Walmsley SL, et al. Better mind the gap: addressing the shortage of HIV-positive women in clinical trials. AIDS 2010; 24:1091–1094.
- Herlitz LC, Mohan S, Stokes MB, Radhakrishnan J, D'Agati VD, Markowitz GS. Tenofovir nephrotoxicity: acute tubular necrosis with distinctive clinical, pathological, and mitochondrial abnormalities. Kidney Int 2010; 78:1171–1177.
- Zaidan M, Lescure FX, Brocheriou I, Dettwiler S, Guiard-Schmid JB, Pacanowski J, et al. Tubulointerstitial nephropathies in HIV-infected patients over the past 15 years: a clinico-pathological study. Clin J Am Soc Nephrol 2013; 8:930–938.
- Rodriguez-Novoa SLP, Soriano V. Pharmacogenetics of tenofovir treatment. Pharmacogenomics 2009; 10:1675–1685.
- 43. Rodríguez-Nóvoa SLP, Soriano V, Egan D, Albalater M, Morello J, Cuenca L, et al. Predictors of kidney tubular dysfunction in HIV-infected patients treated with tenofovir: a pharmacogenetic study. Clin Infect Dis 2009; 48:e108–116.