



Analysis of long-term persistence of resistance mutations within the hepatitis C virus NS3 protease after treatment with telaprevir or boceprevir

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ABSTRACT

Background: Telaprevir and boceprevir are highly selective hepatitis C virus (HCV) NS3/4A protease-inhibitors in phase 3 development. Viral breakthrough during mono- and triple-therapies with PEG-interferon and ribavirin and relapse is associated with resistance.

Objectives: Potential persistence of resistance mutations during long-term follow-up should be analyzed. **Study design:** Clonal sequence analysis of the NS3-protease gene was performed at long-term follow-up in HCV genotype-1 infected patients who received telaprevir or boceprevir within phase-1b studies for comparison with resistant variants present directly after the end-of-treatment.

Results: After a median follow-up of 4.2 years in 28 of 82 patients HCV-RNA was still detectable. Resistance variants were detected in two of 14 telaprevir- and in four of 14 boceprevir-treated patients. For telaprevir patients two low-level (V36M, V36A) and one high-level (A156T) mutation associated with resistance were detected at low frequencies (4–9% of the clones). In five boceprevir-treated patients four low level mutations (V36A, T54A/S, V55A) were observed at low frequencies (1–10%) while in one patient additionally a combined variant (T54S + R155K) was detected at 94%. Presence of resistant variants at long-term follow-up was not predictable by variants detected at the end-of-treatment. In one patient a V55A variant which was dominant already at baseline was still detectable at long-term follow-up.

Conclusions: In the majority of patients after short-term treatment with telaprevir or boceprevir wild-type NS3-protease isolates are detectable by clonal sequencing at long-term follow-up. Detectable resistance mutations in single patients are not predictable by initial frequencies of variants.

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1. Background

Chronic hepatitis C virus (HCV) infection is a serious public health problem affecting an estimated 130 million people worldwide.¹ The current standard-of-care, pegylated interferon-alpha (PEG-IFN) plus ribavirin, is of limited efficacy with eradication of the virus in approx. 50% of patients.² A large number of directly acting antiviral agents (DAA) targeting the nonstructural-(NS)3-protease, the NS5A-protein, the RNA-dependent RNA-polymerase NS5B, as well as host cell proteins are currently in phase-1 to -3 clinical trials.^{3,4} Telaprevir and boceprevir, the most advanced agents,

have completed phase 3 studies. Results of clinical studies have shown a significant improvement of sustained virologic response (SVR) rates with triple therapies with telaprevir or boceprevir in combination with PEG-IFN and ribavirin in treatment-naïve and -experienced HCV genotype-1 infected patients.^{5–8}

For monotherapy with NS3-protease, NS5A-protein and non-nucleoside polymerase inhibitors rapid selection of resistant variants has been observed,^{9,10} which represents one of the major problems of DAAs. Combination therapy with PEG-IFN and ribavirin or with different DAAs has been shown to reduce the likelihood of virologic break-through due to resistance developing.^{7–14} However, even using triple therapy with pegylated interferon, ribavirin and NS3 proteaseinhibitors, still a significant number of patients experience viral breakthrough in association with selection of drug-resistant variants.^{7,8,11,12,14} Furthermore, variants associated with resistance have been detected in patients with relapse after the end-of-treatment.^{11,12,14} Finally, viral variants known to confer resistance to NS3-protease and NS5B non-nucleoside inhibitors have been detected in 0.2–2.8% of patients as the pre-existing dominant strain^{15–18} and preexisting R155K variants seem

Abbreviations: HCV, hepatitis C virus; NS, non-structural protein; RNA, ribonucleic acid; WHO, World Health Organization; DAA, directly acting antiviral agent; TID, every 8 h; BID, every 12 h; wk, week; QID, every 6 h; PEG-IFN α , pegylated interferon-alpha; PCR, polymerase chain reaction; GT, genotype; bp, base pairs; SOC, standard-of-care; IU, international units; HIV, human immunodeficiency virus; HBV, hepatitis B virus; SVR, sustained virologic response.

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Table 1
Results of clonal resistance analysis and characteristics of patients who received TVR therapy.

ID	Therapy	Resistance mutations at end of TVR treatment	SOC after TVR study	GT	Long-term follow-up				
					Years	VL [IU/ml]	Clones sequenced	Sensitivity of seq. analysis	Detected resistance mutations
1	3 × 750 mg TVR	V36A/M 3% R155K/T 37% A156 6% V36A/M + R155K/T 51% V36A/M + A156T 4%	Yes (outcome: BT)	1a	3.5	570.000	21	4.8%	None
2	3 × 450 mg TVR	Sequence data not available ^a	No	1b	4	12.000	20	5%	None
3	3 × 450 mg TVR	V36A 62% T54A 28% V36A + T54A 3% V36A + R155L 1%	Yes (outcome: BT)	1b	4	1.200.000	20	5%	None
4	3 × 450 mg TVR	Sequence data not available ^a	No	1b	4	3.700.000	27	3.7%	None
5	3 × 750 mg TVR	Sequence data not available ^a	No	1b	4	1.400.000	27	3.7%	None
6	2 × 1250 mg TVR	R155M/T/K/G 78% V36A/M + R155K/T 20% T54S + R155T 1%	No	1a	4	913.156	29	3.4%	None
7	3 × 750 mg TVR	A156T/V 98% R155T/K + A156T 2%	No	1a	4	1.100.000	22	4.5%	None
8	3 × 750 mg TVR	A156T/V/I 99% R155Q + A156T 1%	No	1b	4	2.200.000	21	4.8%	None
9	2 × 1250 mg TVR	V36A/M 38% T54A 5% R155K 13% V36M + T54A 1% V36A/M + R155G/K/T 24% T54A + R155K 1%	No	1a	4	1.400.000	27	3.7%	V36M 4%
10	3 × 750 mg TVR	Sequence data not available ^a	Yes (outcome: NR)	1b	4	380.000	25	4%	None
11	3 × 750 mg TVR	Sequence data not available ^a	No	1a	5	19.126	23	4.3%	None
12	2 × 1250 mg TVR	A156T/V 100%	No	1b	5	292.507	31	3.2%	None
13	3 × 750 mg TVR	Sequence data not available ^a	No	1b	5	7.012.950	29	3.4%	None
14	2 × 1250 mg TVR	V36M 5% T54S 1% R155M/T/K/G/S 64% V36M + T54S 2% V36M + R155T/K/G 9% T54S + R155T 1%	Unknown	1a	5	482.693	22	4.5%	V36A 9% A156T 5%

Patients were treated with telaprevir alone (450 mg TID, 750 mg TID, or 1250 mg BID).

^a HCV RNA negative at end of treatment; TVR, telaprevir; SOC, standard-of-care treatment; GT, genotype; VL, viral load; BT, breakthrough during SOC; NR, non-response to SOC.

to be associated with a reduced response to boceprevir and telaprevir.^{19,20}

2. Objectives

Preexisting or selected variants may affect virologic response to DAA. Here, we present long-term follow-up data on patients who were enrolled in phase-1 studies with telaprevir or boceprevir. After a median follow-up of 4.23 years clonal sequence analysis was performed in 28 patients with detectable HCV-RNA for analysis of potential persistence of viral variants (at amino acid (aa) positions 36, 54, 55, 155, 156, and 170) previously described to confer resistance to boceprevir or telaprevir.^{21–27}

3. Study design

3.1. Patient population

Altogether 82 patients with chronic HCV genotyp-1 infection were enrolled in phase-1 clinical trials with telaprevir and boceprevir at Saarland University Hospital. Up to 5.5 years after termination of study treatment 42/82 patients could be contacted for

a long-term follow-up visit. HCV-RNA was still detectable in 34/42 patients ($n = 6$ placebo; $n = 14$ telaprevir; $n = 14$ boceprevir).

3.2. Telaprevir

Thirty-three HCV genotyp-1 infected patients were enrolled into two randomized, double-blind, placebo-controlled phase-1b trials, which were described recently.^{28,29} The patients were treated with telaprevir alone (450 mg TID, 750 mg TID, or 1250 mg BID) or in combination with PEG-IFN- α -2a (180 μ g/wk) for 2 weeks (750 mg TID). Clonal sequence analysis of the NS3-protease gene could be performed in 14 available patients with ongoing HCV-infection during long-term follow-up. Clonal resistance analysis at end-of-treatment was obtained from a previous study.²¹

3.3. Boceprevir

Forty-nine HCV genotyp-1 infected patients were enrolled into randomized, double-blind, placebo-controlled phase-1b trials.^{30,31} The patients were treated with boceprevir alone (200 mg BID, 400 mg BID, or 400 mg TID) or in combination with PEG-IFN- α -2b (1.5 μ g/kg body weight/wk) for 2 weeks (400 mg

Table 2
Results of clonal resistance analysis and characteristics of patients who received BOC therapy.

ID	Therapy	Resistance mutations at end of BOC treatment	SOC after BOC study	GT	Long-term follow-up				
					Years	VL [IU/ml]	Clones sequenced	Sensitivity of seq. analysis	Detected resistance mutations
1	4 × 400 mg BOC + PEG	T54S 42% R155K 9%	Unknown	1b	2.25	1.422.930	27	3.7%	None
2	3 × 400 mg BOC + PEG	T54S 3% R155K 3% T54S + R155K 94%	No	1a	3	46.188	39	2.6%	T54S 3% T54S + R155K 94%
3	3 × 400 mg BOC	T54A 2% V170I 5%	No	1b	3.5	825.115	25	4%	None
4	4 × 600 mg BOC + PEG	V36M 15% V55A 13% V36M + R155K 72%	No	1a	3.75	5.379.862	69	1.4%	T54A 1% V55A 1%
5	4 × 600 mg BOC + PEG	V36A + V170A 2% T54A + V170A 2% A156V + V170A 2% V170A 94%	No	1b	3.75	693.217	72	1.4%	None
6	4 × 400 mg BOC + PEG	T54A 100%	No	1b	3.75	11.900.000	57	1.8%	None
7	3 × 400 mg BOC	None	No	1b	4.5	1.377.400	47	2.1%	None
8	2 × 200 mg BOC	None	No	1b	4.5	307.065	28	3.6%	None
9	3 × 400 mg BOCs	V55A 100%	No	1b	4.75	582.417	20	5%	T54S + V55A 5% V55A 95%
10	3 × 200 mg BOCs	A156T 3%	No	1b	4.75	545.735	20	5%	V36A 10%
11	3 × 200 mg BOCs	V36A 3%	No	1b	4.75	204.631	37	2.7%	None
12	2 × 400 mg BOC	None	No	1b	5	245.345	52	1.9%	None
13	3 × 400 mg BOC	None	No	1b	5.25	341.191	35	2.9%	V55A 3%
14	2 × 200 mg BOC	None	No	1a	5.5	1.254.100	20	5%	None

Patients were treated with boceprevir alone (200 mg BID, 400 mg BID, or 400 mg TID) or in combination with pegylated interferon alpha-2b (1.5 µg/kg body weight/wk) for 2 weeks (400 mg BOC TID, 400 mg BOC QID, or 600 mg BOC QID) or they were randomized to different sequences of three periods of treatment with BOC-mono (200 mg or 400 mg TID) for 7 days, PEG-IFN α -2b-mono (200 mg or 400 mg TID) for 14 days, PEG-IFN α -2b-mono (200 mg or 400 mg TID) for 14 days and combination of the two for 14 days. Between the different treatment schedules therapy was interrupted for 14 days. BOC, boceprevir; PEG, pegylated interferon alpha-2b; BOCs, sequential treatment regimen (BOC mono, PEG mono, BOC + PEG combination); SOC, standard-of-care treatment; GT, genotype; VL, viral load.

TID, 400 mg QID, or 600 mg QID) or they were randomized to different sequences of three periods of treatment with boceprevir-mono (200 mg or 400 mg TID) for 7 days, PEG-IFN α -2b-mono (200 mg or 400 mg TID) for 14 days and combination of the two for 14 days. Between the different treatment schedules therapy was interrupted for 14 days. Clonal sequence analysis of the NS3-protease gene could be performed in 14 available patients with ongoing HCV-infection at long-term follow-up. Clonal resistance analysis was performed in all patients at end-of-treatment in the present study or obtained from previous studies (patients #3, #9, #13).^{27,32}

Methods used in this and the former studies were the same, regarding RNA isolation, RT-PCR, amplification, sequencing, and analyzing sequences.

Written informed consent was obtained from each patient in accordance with the 1975 Declaration of Helsinki.

3.4. HCV-RNA extraction

Viral RNA was extracted from 140 µL serum using the QIAamp Viral RNA Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions. RNA quality was assessed by calculating the absorbance ratio OD_{260nm}/OD_{280nm} using NanoDrop model ND-1000 (PiqLab, Erlangen).

3.5. Amplification and sequencing of the HCV NS3-protease gene

The complete region encoding the NS3-protease was amplified by semi-nested RT-PCR by applying 8 µL of the isolated RNA. The appropriately sized PCR-products were purified using the QIAquick Gel Extraction Kit (Qiagen, Hilden, Germany) and subsequently

cloned using the StrataClone PCR Cloning Kit (Agilent Technologies, La Jolla, CA).

Isolated plasmid DNA from the molecular clones was subjected to sequence-PCR according to the manufacturer's instructions using the M13-forward or -reverse primers (BigDye Deoxy Terminators; Applied Biosystems, Foster City, CA). Sequencing was performed by the 3130xl Genetic Analyzer (Applied Biosystems, Foster City, CA).

Baseline consensus sequences were obtained in the context of the phase-1b studies and act as reference for analysis of the long-term follow-up samples. Baseline consensus sequences were checked for mutations associated with resistance at aa positions 36, 54, 55, 155, 156, and 170 as well.

3.6. Sequence analysis

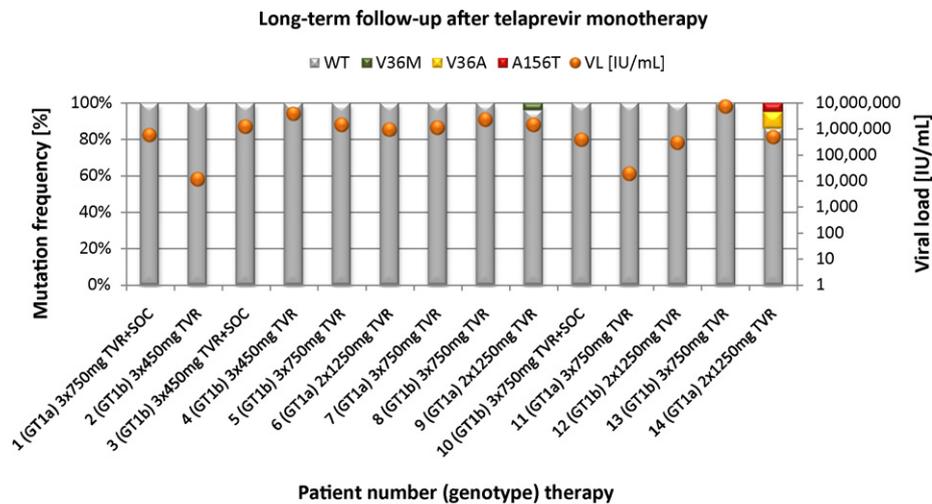
Sequences from the N-terminal 181 aa of the HCV NS3-protease were aligned and analyzed for mutations which were known to confer resistance against telaprevir and/or boceprevir. The sequences were analyzed using the software Mutational Surveyor (SoftGenetics, State College, PA). Phylogenetic analyses were performed with the ClustalW-Multiple Sequence Alignment tool (www.ebi.ac.uk/Tools/msa/clustalw2). The cut-off values for sensitivity of the sequence analysis represent the percentage corresponding to 1 clone out of the number of clones sequenced.

4. Results

4.1. Clinical outcome of patients

4.1.1. Telaprevir

Twenty-one patients presented for a long-term follow-up visit 3.5–5 (mean 4.25 ± 0.5) years after termination of telaprevir ther-



The bars represent the composition of the HCV quasispecies, found by clonal sequencing, in 14 patients, formerly treated with telaprevir, at long-term follow-up (primary y-axis). The corresponding viral loads are shown as orange dots within the bars (secondary y-axis).

WT: wild type; VL: viral load; GT: genotype; TVR: telaprevir; +SOC: standard-of-care treatment after telaprevir monotherapy

Fig. 1. Viral load and resistance mutations in the long-term follow-up of patients initially treated with telaprevir. The bars represent the composition of the HCV quasispecies, found by clonal sequencing, in 14 patients, formerly treated with telaprevir, at long-term follow-up (primary y-axis). The corresponding viral loads are shown as orange dots within the bars (secondary y-axis). WT, wild type; VL, viral load; GT, genotype; TVR, telaprevir; +SOC, standard-of-care treatment after telaprevir monotherapy. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

apy. Seven patients attained SVR after SOC in the meantime. Characteristics of the remaining patients are summarized in Table 1.

dosing. Thus, the HCV NS3-protease gene of 14 patients could be analyzed (Table 2).

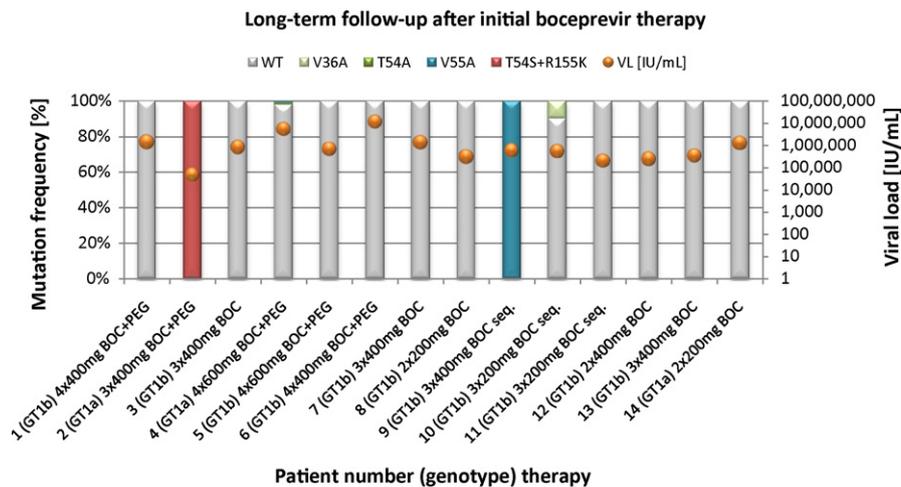
4.1.2. Boceprevir

4.2. Resistance analysis

Fifteen patients were available for a long-term follow-up visit 2.25–5.5 (mean 4.2 ± 0.9) years after direct antiviral therapy with boceprevir. One of them achieved SVR with SOC after boceprevir

4.2.1. Telaprevir

Clonal sequencing at long-term follow-up revealed only wild-type NS3-protease variants in 12 patients. One subtype 1a infected



The bars represent the composition of the HCV quasispecies, found by clonal sequencing, in 14 patients, formerly treated with boceprevir, at long-term follow-up (primary y-axis). The corresponding viral loads are shown as orange dots within the bars (secondary y-axis).

WT: wild type; VL: viral load; GT: genotype; BOC: boceprevir; PEG: pegylated interferon-alfa-2b; seq.: sequential treatment regimen (BOC mono, PEG mono, BOC+PEG combi)

Fig. 2. Viral load and resistance mutations in the long-term follow-up of patients initially treated with boceprevir. The bars represent the composition of the HCV quasispecies, found by clonal sequencing, in 14 patients, formerly treated with boceprevir, at long-term follow-up (primary y-axis). The corresponding viral loads are shown as orange dots within the bars (secondary y-axis). WT, wild type; VL, viral load; BOC, boceprevir; PEG, pegylated interferon-alpha-2b; seq., sequential treatment regimen (BOC mono, PEG mono, BOC + PEG combination). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

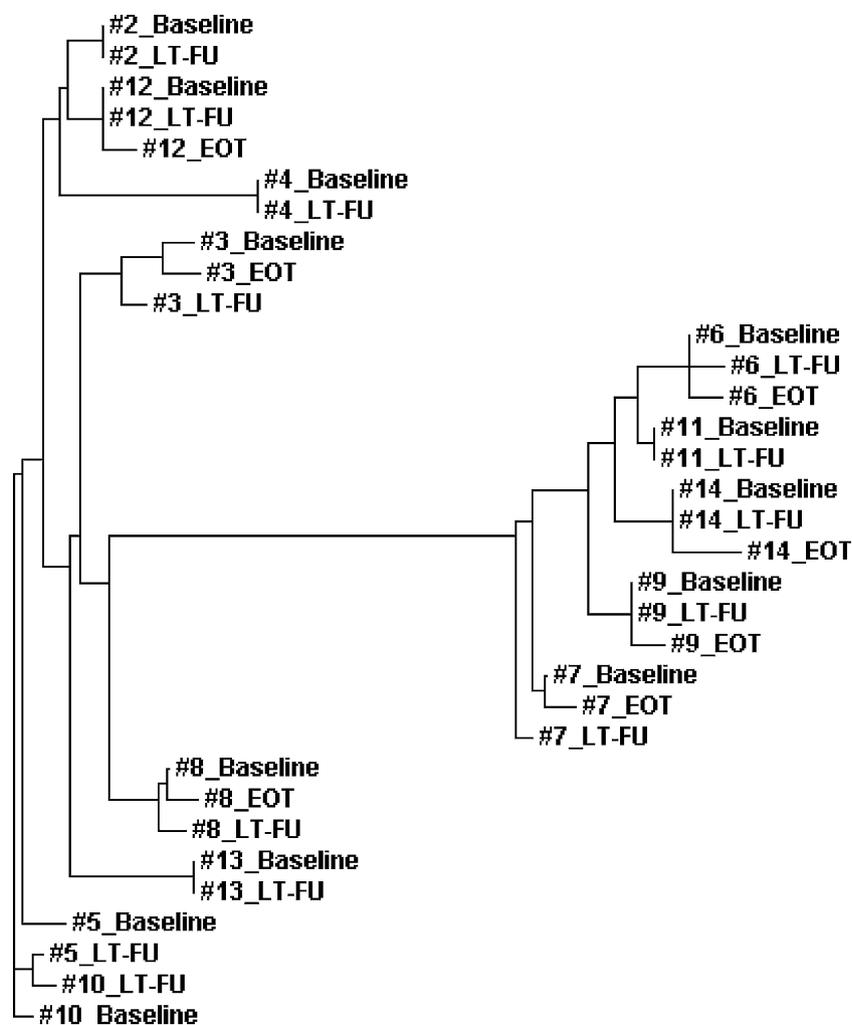


Fig. 3. Phylogenetic analysis of the consensus sequences from baseline, end-of-treatment, and long-term follow-up of each patient treated with telaprevir.

patient (#9, 2× 1250 mg) showed the V36M variant at 4% frequency, known to confer low level resistance to telaprevir. In another subtype 1a patient (#14, 2× 1250 mg) 9% of the analyzed clones showed V36A and even A156T, conferring the highest resistance against telaprevir, was present in 5% of the quasiespecies. In patient #9 V36M was already detectable at the end of telaprevir-monotherapy, while patient #14 did not exhibit V36A nor A156T after telaprevir treatment. Both patients with NS3 protease-inhibitor resistance variants were infected with HCV subtype 1a. Patient #9 did not receive SOC between completion of the telaprevir phase-1b study and long-term follow-up, for patient #14 this information is unknown (Table 1, Fig. 1).

4.2.2. Boceprevir

In patient #2 (GT1a; 3× 400 mg boceprevir+PEG) 94% of all clones contained the T54S+R155K double mutation and 3% contained T54S as single variant 3 years after proteaseinhibitor treatment. Directly after EOT T54S+R155K was present in 94% together with 3% T54S and 3% R155K as single mutations. In patient #4 (GT1a), 3.75 years after receiving 600 mg boceprevir QID plus PEG-IFN 1% T54A and 1% V55A were detected, each on separate clones (cut-off 1.4%). Contrary to V55A (13% frequency at EOT) T54A could not be detected after the end-of-treatment. V36M (EOT: 15%) and V36M+R155K (EOT: 72%) decreased completely to undetectable levels (<1.4%) during the 3.75 years. In patient #9 (GT1b), after 4.75 years we found 95% V55A and 5% T54S+V55A

and in patient #10 (GT1b) 10% V36A, both were treated sequentially with boceprevir mono for 7 days (patient #9: 3× 400 mg; patient #10: 3× 200 mg), PEG mono and boceprevir + PEG in combination for 14 days, respectively. V55A which was detected in 100% of the clones from patient #9, was already present at baseline and at the end-of-treatment as natural and only variant with a frequency of 100%.³² Directly after boceprevir dosing, patient #10 exhibited only A156T (3%), which was no longer detectable (cut-off: 5%) in the long-term follow-up analysis. Patient #13 (GT1b) showed 3% V55A (cut-off: 2.9%) 5.25 years after boceprevir therapy (3× 400 mg), while no variants were detected after EOT.

The remaining nine patients showed no resistant variants above the limit of detection, despite detectable resistance mutations at the end of DAA treatment with boceprevir alone or in combination with PEG-interferon-alpha (patients #1, #3, #5, #6, and #11; Table 2, Fig. 2).

4.3. Phylogenetic analysis

With phylogenetic analyses we confirmed that the long-term follow-up sequences are closely related to the baseline and end-of-treatment sequences. The consensus sequences within one patient at these time points show an accordance of at least 95%, whereas the consensus sequences of different patients show identities down to 81% (Figs. 3 and 4).

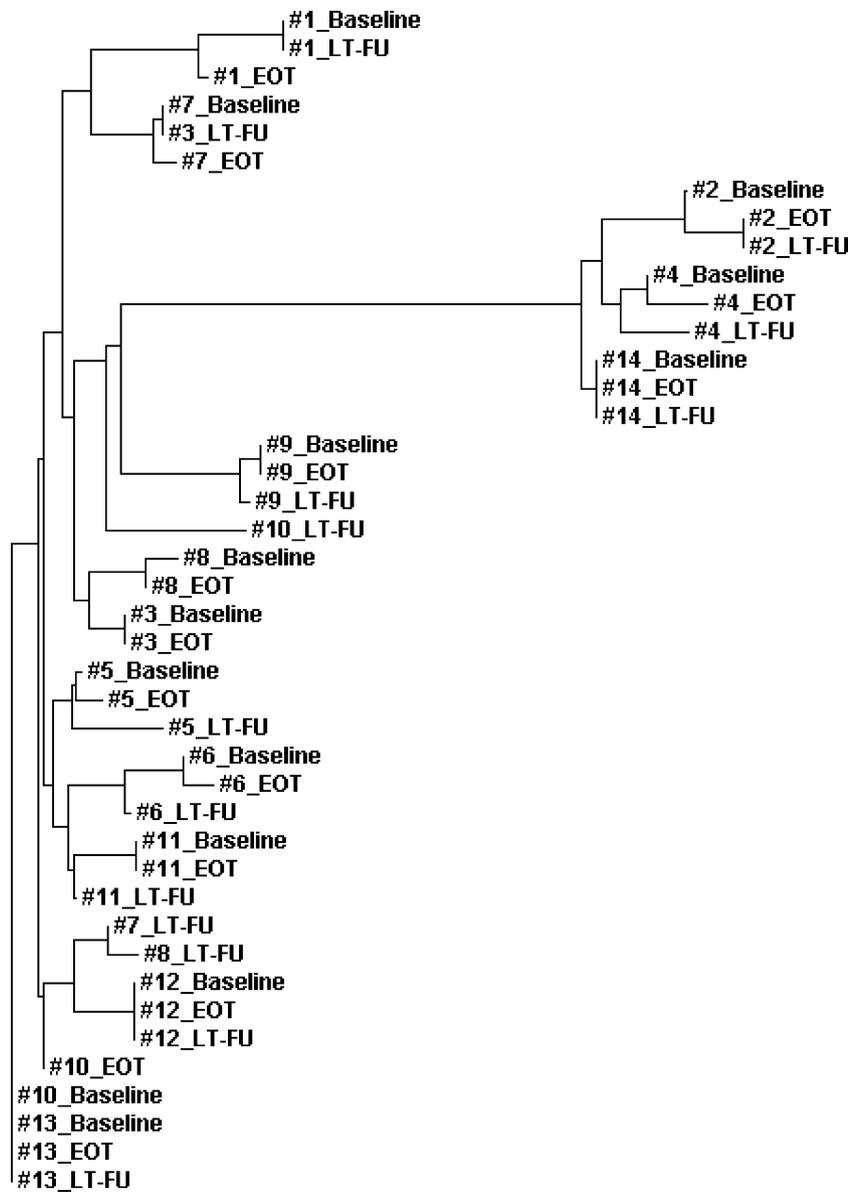


Fig. 4. Phylogenetic analysis of the consensus sequences from baseline, end-of-treatment, and long-term follow-up of each patient treated with boceprevir.

5. Discussion

HCV is a positive-stranded RNA virus with replication without a DNA intermediate and no mechanisms are known for HCV-RNA to be archived.^{33,34} Due to the short half-life, the rapid turnover and the low fidelity of the HCV NS5B RNA-dependent RNA-polymerase a high number of viral variants is generated with production of all possible single and double variants every day.^{35,36} Indeed, highly sensitive clonal sequence analyses showed a rapid selection of resistant variants during the first days of treatment with the HCV NS3 proteaseinhibitors telaprevir and boceprevir but also a rapid disappearance of resistant variants shortly after withdrawal of direct antiviral treatment.^{21,22,27} However, mutations within the NS3-protease conferring resistance to proteaseinhibitors do not impair infectivity of HCV virions and thus may persist at low levels also during long-term follow-up. In the present study, in the majority of patients only wild-type NS3-protease isolates were detectable 2.25–5.5 (mean 4.23 ± 0.7) years after the end of telaprevir or boceprevir treatment by clonal sequence analysis. Only in two patients with initial telaprevir treatment (1250 mg BID) and in five patients with initial

boceprevir treatment (600 mg QID, 400 mg and 200 mg TID, respectively) resistant variants have been detected by clonal sequencing at long-term follow-up. V36M in patient #9 seems to be persisting since its selection during telaprevir monotherapy where it was detectable at end-of-treatment already. Initial sequencing of patient #14 was obtained at EOT after a viral breakthrough during telaprevir-monotherapy and V36A/A156T were not detectable at this time point.²¹ However, it could be possible, that V36A and A156T, which were detected by clonal analysis 5 years later, were present at earlier timepoints during telaprevir treatment. A156T has been observed as natural variant in the liver of an untreated patient and thus so far unknown compensatory mechanisms explaining the relatively high replication capacity of this variant within the HCV quasispecies may exist.³⁷ Interestingly, for boceprevir and telaprevir resistance variants, the mutational pattern detected at long-term follow-up could not generally be predicted from variants present at the end-of-treatment which most likely is explained by the highly dynamic changes of HCV quasispecies.

Our study has several limitations: Boceprevir was dosed at lower levels than in phase-2/3 studies. Furthermore, during

phase-1b studies all patients received only short-term treatment with boceprevir or telaprevir. Longer treatment durations and especially a continuous dosing of the proteaseinhibitor after viral breakthrough or persistent HCV replication during triple therapies with PEG-interferon and ribavirin might increase the probability for selection of resistance variants. Such variants together with compensatory mutations may have a higher replicative fitness and thus a higher likelihood for long-term persistence at significant frequencies within the HCV-quasispecies. Finally, clonal sequence analysis performed in the initial phase-1 studies as well as in the present study have limited sensitivities to detect resistance mutations present at low frequencies. Further studies on potential persistence of resistant variants by deep-sequencing methods are required to complete our understanding of viral turnover of these variants.

Taken together, we have shown a rapid decline of the number of patients with NS3-protease resistance mutations as well as the frequency of resistant viral variants within the HCV quasispecies from the initial months after the end of short treatment intervals until long-term follow-up of up to 5.5 years. Future studies based on clonal resistance analysis or even deep-sequencing should be performed after failure to full-course telaprevir and boceprevir treatment regimens.

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Competing interests

SS, JV, NF, MWW, NG, CF, and DP have nothing to disclose. SZ and CS are consultants and received research support from MSD/Merck and Vertex/Tibotec.

Ethical approval

Not required.

References

- World Health Organisation (WHO). *Hepatitis C. Initiative for Vaccine Research*; 2009. http://www.who.int/vaccine_research/diseases/viral_cancers/en/index2.html [accessed January 21, 2009].
- Zeuzem S, Berg T, Moeller B, Hinrichsen H, Mauss S, Wedemeyer H, et al. Expert opinion on the treatment of patients with chronic hepatitis C. *J Viral Hepat* 2009;**16**(February):75–90.
- Asselah T, Benhamou Y, Marcellin P. Protease and polymerase inhibitors for the treatment of hepatitis C. *Liver Int* 2009;**29**(January (Suppl. 1)):57–67.
- Thompson AJ, McHutchison JG. Antiviral resistance and specifically targeted therapy for HCV (STAT-C). *J Viral Hepat* 2009;**16**(June):377–87.
- Zeuzem S, Andreone P, Pol S, Lawitz E, Diago M, Roberts S, et al. Telaprevir for retreatment of HCV infection. *N Engl J Med* 2011;**364**(June):2417–28.
- Jacobson IM, McHutchison JG, Dusheiko G, Di Bisceglie AM, Reddy KR, Bzowej NH, et al. Telaprevir for previously untreated chronic hepatitis C virus infection. *N Engl J Med* 2011;**364**(June):2405–16.
- Bacon BR, Gordon SC, Lawitz E, Marcellin P, Vierling JM, Zeuzem S, et al. Boceprevir for previously treated chronic HCV genotype 1 infection. *N Engl J Med* 2011;**364**(March):1207–17.
- Poordad F, McCone Jr J, Bacon BR, Bruno S, Manns MP, Sulkowski MS, et al. Boceprevir for untreated chronic HCV genotype 1 infection. *N Engl J Med* 2011;**364**(March):1195–206.
- Sarrazin C, Zeuzem S. Resistance to direct antiviral agents in patients with hepatitis C virus infection. *Gastroenterology* 2010;**138**(February):447–62.
- Kieffer TL, Kwong AD, Picchio GR. Viral resistance to specifically targeted antiviral therapies for hepatitis C (STAT-Cs). *J Antimicrob Chemother* 2010;**65**(February):202–12.
- McHutchison JG, Everson GT, Gordon SC, Jacobson IM, Sulkowski M, Kauffman R, et al. Telaprevir with peginterferon and ribavirin for chronic HCV genotype 1 infection. *N Engl J Med* 2009;**360**(April):1827–38.
- Hezode C, Forestier N, Dusheiko G, Ferenci P, Pol S, Goefer T, et al. Telaprevir and peginterferon with or without ribavirin for chronic HCV infection. *N Engl J Med* 2009;**360**(April):1839–50.
- Gane EJ, Roberts SK, Stedman CA, Angus PW, Ritchie B, Elston R, et al. Oral combination therapy with a nucleoside polymerase inhibitor (RG7128) and danoprevir for chronic hepatitis C genotype 1 infection (INFORM-1): a randomised, double-blind, placebo-controlled, dose-escalation trial. *Lancet* 2010;**376**(October):1467–75.
- McHutchison JG, Manns MP, Muir AJ, Terrault NA, Jacobson IM, Afdhal NH, et al. Telaprevir for previously treated chronic HCV infection. *N Engl J Med* 2010;**362**(April):1292–303.
- Kuntzen T, Timm J, Berical A, Lennon N, Berlin AM, Young SK, et al. Naturally occurring dominant resistance mutations to hepatitis C virus protease and polymerase inhibitors in treatment-naïve patients. *Hepatology* 2008;**48**(December):1769–78.
- Gaudieri S, Rauch A, Pfafferott K, Barnes E, Cheng W, McCaughan G, et al. Hepatitis C virus resistance and immune-driven adaptations: relevance to new antiviral therapy. *Hepatology* 2009;**49**(April):1069–82.
- Kim AY, Timm J, Nolan BE, Reyrol LL, Kane K, Berical AC, et al. Temporal dynamics of a predominant proteaseinhibitor-resistance mutation in a treatment-naïve, hepatitis C virus-infected individual. *J Infect Dis* 2009;**199**(March):737–41.
- Colson P, Brouk N, Lembo F, Castellani P, Tamalet C, Gerolami R. Natural presence of substitution R155K within hepatitis C virus NS3 protease from a treatment-naïve chronically infected patient. *Hepatology* 2008;**47**(February):766–7.
- Bartels DJ, Zhou Y, Zhang EZ, Marcial M, Byrn RA, Pfeiffer T, et al. Natural prevalence of hepatitis C virus variants with decreased sensitivity to NS3-4A proteaseinhibitors in treatment-naïve subjects. *J Infect Dis* 2008;**198**(September):800–7.
- Vierling J, Kwo P, Lawitz E, McCone J, Schiff E, Pound D, et al. Frequencies of resistance-associated amino acid variants following combination treatment with boceprevir plus PEGINTRON (peginterferon alfa-2b)/ribavirin in patients with chronic hepatitis C (CHC), genotype 1 (G1). *Hepatology* 2010;**52**:702A–3A.
- Sarrazin C, Kieffer TL, Bartels D, Hanzelka B, Muh U, Welker M, et al. Dynamic hepatitis C virus genotypic and phenotypic changes in patients treated with the proteaseinhibitor telaprevir. *Gastroenterology* 2007;**132**(May):1767–77.
- Kieffer TL, Sarrazin C, Miller JS, Welker MW, Forestier N, Reesink HW, et al. Telaprevir and pegylated interferon-alpha-2a inhibit wild-type and resistant genotype 1 hepatitis C virus replication in patients. *Hepatology* 2007;**46**(September):631–9.
- Tong X, Bogen S, Chase R, Girijavallabhan V, Guo Z, Njoroge RG, et al. Characterization of resistance mutations against HCV ketoamide proteaseinhibitors. *Antiviral Res* 2008;**77**(March):177–85.
- Flint M, Mullen S, Deatly AM, Chen W, Miller LZ, Ralston R, et al. Selection and characterization of hepatitis C virus replicons dually resistant to the polymerase and proteaseinhibitors HCV-796 and boceprevir (SCH 503034). *Antimicrob Agents Chemother* 2009;**53**(February):401–11.
- Qiu P, Sanfiorenzo V, Curry S, Guo Z, Liu S, Skelton A, et al. Identification of HCV proteaseinhibitor resistance mutations by selection pressure-based method. *Nucleic Acids Res* 2009;**37**(June):e74.
- Barbotte L, Ahmed-Belkacem A, Chevaliez S, Soulier A, Hezode C, Wajcman H, et al. Characterization of V36C, a novel amino acid substitution conferring hepatitis C virus (HCV) resistance to telaprevir, a potent peptidomimetic inhibitor of HCV protease. *Antimicrob Agents Chemother* 2010;**54**(June):2681–3.
- Susser S, Welsch C, Wang Y, Zettler M, Domingues FS, Karey U, et al. Characterization of resistance to the proteaseinhibitor boceprevir in hepatitis C virus-infected patients. *Hepatology* 2009;**50**(December):1709–18.
- Reesink HW, Zeuzem S, Weegink CJ, Forestier N, van Vliet A, van de Wetering de Rooij, et al. Rapid decline of viral RNA in hepatitis C patients treated with VX-950: a phase 1b, placebo-controlled, randomized study. *Gastroenterology* 2006;**131**(October):997–1002.
- Forestier N, Reesink HW, Weegink CJ, McNair L, Kieffer TL, Chu HM, et al. Antiviral activity of telaprevir (VX-950) and peginterferon alfa-2a in patients with hepatitis C. *Hepatology* 2007;**46**(September):640–8.
- Sarrazin C, Rouzier R, Wagner F, Forestier N, Larrey D, Gupta SK, et al. SCH 503034, a novel hepatitis C virus proteaseinhibitor, plus pegylated interferon alpha-2b for genotype 1 nonresponders. *Gastroenterology* 2007;**132**(April):1270–8.
- Zeuzem S, Sarrazin C, Rouzier R, Tarral A, Brion N, Gupta S, et al. Anti-viral activity of SCH 503034, a HCV proteaseinhibitor, administered as monotherapy in hepatitis C genotype-1 (HCV-1) patients refractory to pegylated interferon (PEG-IFN-alpha). *Hepatology* 2005;**42**(October (Suppl. 4)):233A–4A.
- Vermehren J, Susser S, Lange CM, Forestier N, Karey U, Hughes EA, et al. Mutations selected in the hepatitis C virus NS3 protease domain during sequential treatment with boceprevir with and without pegylated interferon alfa-2b. *J Viral Hepat* 2011. Epub.
- Rehermann B, Nascimbeni M. Immunology of hepatitis B virus and hepatitis C virus infection. *Nat Rev Immunol* 2005;**5**(March):215–29.
- Turner BG, Summers MF. Structural biology of HIV. *J Mol Biol* 1999;**285**(January):1–32.
- Duffy S, Shackleton LA, Holmes EC. Rates of evolutionary change in viruses: patterns and determinants. *Nat Rev Genet* 2008;**9**(April):267–76.
- Rong L, Dahari H, Ribeiro RM, Perelson AS. Rapid emergence of proteaseinhibitor resistance in hepatitis C virus. *Sci Transl Med* 2010;**2**(May):30ra32.
- Cubero M, Esteban JI, Otero T, Sauleda S, Bes M, Esteban R, et al. Naturally occurring NS3-protease-inhibitor resistant mutant A156T in the liver of an untreated chronic hepatitis C patient. *Virology* 2008;**370**(January):237–45.