New perspectives for preventing hepatitis C virus liver graft infection



Daniel J Felmlee*, Audrey Coilly*, Raymond T Chung, Didier Samuel†, Thomas F Baumert†

Hepatitis C virus (HCV) infection is a leading cause of end-stage liver disease that necessitates liver transplantation. The incidence of virus-induced cirrhosis and hepatocellular carcinoma continues to increase, making liver transplantation increasingly common. Infection of the engrafted liver is universal and accelerates progression to advanced liver disease, with 20–30% of patients having cirrhosis within 5 years of transplantation. Treatments of chronic HCV infection have improved dramatically, albeit with remaining challenges of failure and access, and therapeutic options to prevent graft infection during liver transplantation are emerging. Developments in directed use of new direct-acting antiviral agents (DAAs) to eliminate circulating HCV before or after transplantation in the past 5 years provide renewed hope for prevention and treatment of liver graft infection. Identification of the ideal regimen and use of DAAs reveals new ways to treat this specific population of patients. Complementing DAAs, viral entry inhibitors have been shown to prevent liver graft infection in animal models and delay graft infection in clinical trials, which shows their potential for use concomitant to transplantation. We review the challenges and pathology associated with HCV liver graft infection, highlight current and future strategies of DAA treatment timing, and discuss the potential role of entry inhibitors that might be used synergistically with DAAs to prevent or treat graft infection.

Introduction

Hepatitis C virus (HCV) infection causes more than half of all liver transplantations in North America and Europe. 1-5 Furthermore, infection of engrafted livers is universal and accelerates progression to advanced liver disease, with 20-30% of patients having cirrhosis within 5 years of transplantation.⁶ The engrafted liver always becomes infected and undergoes rapid progression of fibrosis to serious liver disease. HCV infection is, therefore, associated with the poorest post-transplantation survival rates compared with other causes leading to liver transplantation.7 The accelerated natural history of allograft HCV infection in patients undergoing re-transplantation makes re-transplantation an ethically challenging proposition because organ shortages lead to selection of the best candidates in terms of survival, and patient and graft survival rates after re-transplantation are inferior to those after primary liver transplantation. Direct-acting antiviral agent (DAA) therapies, developed in the past 5 years, have proved effective in treating chronic HCV infection, and appear more effective in the liver transplant setting than conventional interferonbased treatments in patients with HCV genotype 1. However, treatment options are still limited for patients who need liver transplants as a result of HCV infection, as transplantation requires immunosuppressive drugs to avoid graft rejection with potential drug-drug interactions, the diminished health of this patient population, and the metabolic burden placed on the newly engrafted liver by co-administered pharmaceutical drugs.

The most straightforward means of avoiding the pathogenesis of liver graft infection would be to use preventive measures to avoid graft infection, but the strong efficacy of available DAAs could allow withholding antiviral treatment during the operative stage and addressing HCV infection postoperatively. Here, we

review the specific hurdles associated with HCV infection in liver transplantation, evidence supporting treatment strategies of patients needing transplantation, and the outlook for prophylactic measures against liver graft infection

Challenges of HCV liver graft infection Universal graft infection in HCV RNA positive patients

Because of the burden of HCV on transplants, new, potent DAAs are anticipated to decrease transplantation activity, thereby reducing the number of patients presenting with hepatocellular carcinoma and decompensated cirrhosis.⁸ To achieve this goal, however, comprehensive screening is necessary, because most patients with chronic HCV infection only seek medical care after liver-related complications.⁹ A positive outlook is warranted given that findings from a 2013 analysis showed that a decline of more than 90% in total infections could be achievable by 2030, though this result will require a 3–5-times increase in diagnosis and treatment.¹⁰ However, the public health strategy to address this widespread problem must hope for the best while planning for the worst.

HCV recurrence after liver transplantation remains universal in all patients with detectable serum HCV RNA before transplantation. 30% of patients who have undetectable levels of serum HCV RNA on therapy before transplantation relapse, excluding patients who have sustained virological response (SVR) to therapy for an extended period. HCV recurrence is an important medical problem and is responsible for an increased risk of death and graft failure. Detection of HCV RNA in transplant recipients before transplantation is associated with diminished 5-year patient survival (69.9% vs 76.6%, p<0.0001) and allograft survival (56.8% vs 67.7%, p<0.0001) compared with patients who had a transplant for an indication other than HCV.12

Lancet Infect Dis 2016; 16: 735-45

*Contributed equally as first authors

†Contributed equally as last

Inserm. U1110. Institut de Recherche sur les Maladies Virales et Hépatiques. Strasbourg, France (D J Felmlee PhD, Prof T F Baumert MD); Université de Strasbourg. Strasbourg, France (DJ Felmlee, Prof T F Baumert); Hepatology Research Group, Peninsula School of Medicine and Dentistry, University of Plymouth, Plymouth, UK (D | Felmlee): AP-HP Hôpital Paul Brousse, Centre Hépato-Biliaire, Villeiuif, France (A Coilly MD, Prof D Samuel MD): University Paris-Sud, UMR-S 1193, Villejuif, France (A Coilly, Prof D Samuel); Inserm Unit 1193. Villeiuif F-94800. France (A Coilly, Prof D Samuel); Gastrointestinal Unit Massachusetts General Hospital Harvard Medical School, Boston, MA, USA (Prof RT Chung MD, Prof T F Baumert): and Institut Hospitalo-Universitaire, Pôle Hépato-digestif, Hôpitaux Universitaires de Strasbourg, Strasbourg, France (ProfT F Baumert)

Correspondence to:
Prof Thomas F Baumert,
Inserm Unit 1110, Hôpitaux
Universitaires de Strasbourg,
3 Rue Koeberlé,
F-67000 Strasbourg, France
thomas.baumert@unistra.fr

or

Prof Didier Samuel, Inserm Unit 1193, Villejuif F-94800, France

didier.samuel@aphp.fr

Reinfection is not only a serious problem for the recipient, but also impacts negatively on the precious resource of suitable donated organs.

Rapid fibrosis progression after liver transplantation

The diminished 5-year survival rate in patients who had a transplant because of HCV infection is attributed to accelerated development of pathology due to the immunesuppressive drugs given to prevent graft rejection. Whereas the average time of progression from initial HCV infection to cirrhosis is about 30 years, 20-30% of transplant recipients develop cirrhosis within 5 years.6 Whereas only 30% of non-transplant patients with cirrhosis have liver decompensation after 10 years of cirrhosis, more than 40% of graft recipients decompensate within 12 months of liver transplantation, of whom less than 50% survive the following year. Progression to fibrosis in the context of HCV recurrence varies widely depending on individual patient characteristics, whereas the average time of progression to cirrhosis after liver transplantation is 10-12 years.13 Re-transplantation is the only therapeutic option to achieve long-term survival of patients with decompensated cirrhosis after transplantation. Because of poor patient and graft posttransplant survival rates and the paucity of suitable organ donations, re-transplantation is not a sustainable option in most countries.14

An important clinical challenge is to identify scenarios of early and rapid fibrosis development in which to use early intervention while minimising liver damage, which highlights the importance of the development of diagnostics. The previous consensus was that interferonbased antiviral therapy should be initiated after detection of chronic hepatitis of the liver graft, usually greater than F1 on the METAVIR fibrosis scoring system. The diagnosis of HCV recurrence is typically based on liver biopsy detection, because biopsies can reveal severity of disease progression and exclude other possible diagnoses. Substantial periportal sinusoidal fibrosis in early biopsies (<6 months) has been shown to be a good predictor of severe HCV recurrence.15 Use of serum markers that decisively indicate fibrosis progression and other non-invasive techniques that measure liver stiffness will contribute to future decision making in post-transplantation HCV treatment. Liver stiffness values of less than 8.7 kPa have a 90% negative predictive value and can be used as a threshold to define substantial fibrosis.16 Pressure levels exceeding 6 mm Hg of hepatic venous pressure likewise indicate fibrosis.17

Robust recurrence of HCV RNA levels soon after transplantation is associated with poor prognosis, so early monitoring of HCV levels is crucial. Robust recurrence occurs in 2–8% of patients and often results in fibrosing cholestatic hepatitis. Fibrosing cholestatic hepatitis is characterised by high levels of cholestatic enzymes and the presence of extensive dense portal fibrosis with immature fibrous bands extending into the

sinusoidal spaces, ductular proliferation, cholestasis, and moderate mononuclear inflammation detected in liver graft biopsies.¹⁸ If symptoms do not improve with antiviral drug treatment, fibrosing cholestatic hepatitis typically proceeds to complete liver failure.

Several risk factors that contribute to rapid and severe fibrosis progression have been identified. High HCV RNA levels in either serum or liver tissue are associated with increased progression to cirrhosis, graft loss, and death.¹⁹ Recipient and donor characteristics associated with poor outcome include female sex, donor age, and graft steatosis, while HLA matching and *IL28B* (also known as *IFNL3*) genotype negatively associate with poor outcome.^{4,20,21} Although some of these factors can be selected for before transplantation, others are unpredictable and only antiviral treatment can improve the prognosis of transplant recipients.

The strategic options of HCV treatment with liver transplantation can be divided based on timing of treatment, HCV clearance before transplantation, inhibition of graft infection concomitant with transplantation, or antiviral treatment after graft infection (figure 1).

DAA-based strategies for prevention and treatment of liver graft infection

HCV cure before transplantation

The optimum strategy is, clearly, to prevent reinfection early by eliminating HCV infection prior to transplantation. This strategy had been difficult to apply until approval of DAAs, because interferon-based therapies have limited effectiveness for those with advanced disease while on a transplantation waiting list with SVR being achieved in only 8-39% of cases. Tolerability of interferon treatment is generally poor in these patients and contraindicated in patients with decompensated cirrhosis who require either dose reduction (70%) or early discontinuation (30%) of treatment. The results of a phase 2 clinical study²⁸ of sofosbuvir with ribavirin in 61 patients on waiting lists for liver transplants show that this approach with DAAs is efficacious. Of 43 patients who had a viral response prior to transplantation, 70% had sustained viral clearance 12 weeks after transplantation. However, the efficacy of this strategy is genotype-dependent and managing DAA combinations in the pre-transplant period is challenging. The use of sofosbuvir and ribavirin in advanced cirrhosis could contribute to lactic acidosis in approximately 14% of patients.²⁹ In the SOLAR-1 trial,³⁰ Charlton and colleagues investigated the NS5A inhibitor ledipasvir in combination with sofosbuvir and ribavirin for patients with cirrhosis and moderate or severe hepatic impairment due to genotype 1 and 4 infections. SVR12 was achieved in 86-89% in this difficult-to-treat cohort. The inclusion of ledipasvir with sofosbuvir and ribavirin has been investigated in the SOLAR-2 study31,32 and inclusion of daclatasvir with sofosbuvir and ribavirin was studied in

the ALLY-133 trial, focusing on patients with advanced liver disease pre-transplantation or with recurrent HCV post-transplantation. In the SOLAR-2 study, patients with the sofosbuvir-sensitive HCV genotypes 1 or 4 were treated for 12 or 24 weeks with sofosbuvir, ledipasvir, and ribavirin. Preliminary results revealed high SVR rates of 85–88%, irrespective of treatment duration in genotype 1. Longer treatment duration was superior (SVR rate of 86% vs 57%) for patients with genotype 4 infection, and effective antiviral therapy was associated with improvement in liver histology, Model for End Stage Liver Disease (MELD), and Child-Pugh (CP) scores.³² For some patients with decompensated cirrhosis, however, MELD or CP scores increased. Searching for prognostic factors of clinical or biological response instead of only viral response is an ongoing and needed area of investigation.

The ALLY-1 phase 3 study³³ included 60 patients with advanced cirrhosis who were given daclatasvir, sofosbuvir, and ribavirin. Although overall the SVR12 was 83%, response depended on severity of liver disease (92% for CP A [lowest severity], 94% for CP B, and 56% for CP C [highest severity]). These findings suggest that further studies are needed to define the best therapy management for CP C patients.

Although the inclusion of sofosbuvir has had an effect on the management of genotype 1 infection, use of this drug did not substantially improve treatment for genotype 3 infection by comparison with genotype 1. Foster and colleagues³⁴ analysed the addition of NS5A inhibitors to sofosbuvir and ribavirin treatment in patients with decompensated cirrhosis caused by genotypes 1 or 3.32 The response rates varied between 44% and 88%, depending on genotype, NS5A inhibitor, and the use of ribavirin. Addition of ledipasvir to sofosbuvir and ribavirin was inferior to daclatasvir plus sofosbuvir in patients with genotype 3 infection. Over 40% of patients had improved liver function with a mean improvement of more than 2 points on the MELD score. Overall, these combinations showed good efficacy results and safety profiles, although some patients had worsening of their MELD scores. However, severity of cirrhosis remains an impediment to response, even with the new combinations of DAAs. Although the combination of sofosbuvir and NS5A inhibitor velpatasvir for 12 weeks provides an SVR in over 95% of patients without cirrhosis, 35,36 SVR rate was 83% (n=90) in patients with decompensated cirrhosis.³⁷ However, the addition of ribavirin to this combination improves the SVR rate to 94%, even in patients with cirrhosis.³⁷

The crucial argument for treatment before transplantation is the prospect of avoiding liver transplantation altogether for individuals with liver disease that has not progressed to hepatocellular carcinoma. About two-thirds of patients achieve clinical and biological improvement during treatment in studies that enrolled patients with decompensated

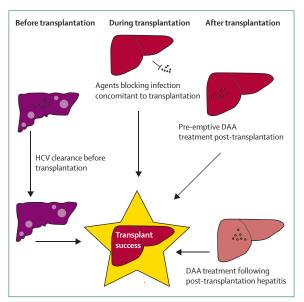


Figure 1: Timing of antiviral strategies for successful liver transplantation in HCV-infected patients

After a patient presents with a cirrhotic liver (upper left) with HCV infection (green dots), multiple strategies have been proposed for a successful, non-infected transplantation. Successful treatment that eliminates the virus before transplantation (left) has been successful in approximately 70% of patients given DAAs. ²¹ Evidence has shown that immunoprevention and HCV entry antagonists can impair infection concomitant to transplantation (centre, top). ²²⁻²⁵ Allowing infection and treating later with DAAs (right, top and bottom) has proven successful in 70–97% of cases depending on the study and drug used. ^{26,27} DAA=direct-acting antiviral. HCV=hepatitis C virus.

cirrhosis.30,33,37,38 However, our own analysis of the data revealed that a third of patients do not improve or worsen during treatment, regardless of antiviral response. For patients who do improve, the difference is often modest with variations of only 1 or 2 points in MELD score. In a meta-analysis³⁹ involving five studies and including 533 patients, 28% had an improvement of MELD score over 3. Some patients do improve to the point where liver transplantation is avoidable. A French cohort study³⁸ of 183 patients awaiting transplantation showed that of 53 patients with decompensated cirrhosis, 36% had a complete clinical and biological response, meaning a CP A at the end of treatment. The best improvement was in patients with the least disease progression—those with baseline CP with an area under the curve of 0.81; CP threshold of improvement 7.5. This raises further questions, because some patients keep improving over longer periods of follow-up. When considering longer followup times for patients while on waiting lists for liver transplantation, comorbidities need to be considered. Optimum conditions and thresholds have not been defined for removing patients from waiting lists. Although we could expect significant improvement for a third of patients, this improvement is more likely to be seen in patients with less severe disease. For individuals with more severe disease, caution should be taken

Cirrhosis (percentage of patients with cirrhosis, %)	SVR12	Reference, trial name	
1) Yes (40%)	70%	Charlton ⁵⁶	
1) Yes	94%	Poordad, ³³ ALLY-1	
(1) Yes (31%)	96%	Coilly, ⁵⁸ ANRS CO23 CUPILT	
No	97%	Kwo, ⁵⁹ CORAL-I	
and G4 Yes (about 50%)	92%	Charlton, ³⁰ SOLAR I; Manns, ³¹ SOLAR II	
F3-F4 (29%)	90%	Pungpapong ⁶⁰	
(1) Yes (56%)	90% (SVR4)	Brown, ⁶¹ HCV-TARGET	
r=boosted with ritonavir. SVR12=sustained virological response at 12 weeks. G1=genotype 1. F3–F4=METAVIR fibrosis score 3–4. SVR4=sustained virological response at 4 weeks.			
r=boosted with ritonavir. SVR12=sustained virological response at 12 weeks. G1=genotype 1. F3-F4=METAVIR fibrosis score 3-4. SVR4=sustained virological response 4 weeks. Table: Available results of DAA-based regimens to treat HCV recurrence after liver transplantation			

because treating HCV in patients only to achieve partial biological improvement might be deleterious.

Patients on transplantation waiting lists with hepatocellular carcinoma, who normally present with compensated cirrhosis, have several approved regimens available, albeit regimens with limited efficacy. In fact, treatment can be unsuccessful in about 30% of patients considering the rate of dropout before transplantation and early hepatocellular carcinoma recurrence after liver transplantation. The severe disease state of patients on the transplantation waiting list can limit treatment options and the 12 weeks needed to confirm SVR status is not always afforded before transplantation. Patients with severe end-stage liver disease prior to liver transplantation or who require complicated post-operative treatment are frequently ineligible for preemptive interferon therapy.

DAA treatment after HCV graft infection

At present, two therapeutic approaches can be considered after transplantation: the pre-emptive strategy involving treatment in the first month following transplantation, or not to start treatment until chronic hepatitis is observed. Despite the clear benefits of early treatment, the pre-emptive strategy is historically not used because of safety and efficacy limitations of initiating interferonbased antiviral therapy during the postoperative period. Heavy DAA, interferon-free combination therapy could revive this strategy, although the evidence regarding efficacy is scarce because of these therapies are new. Factors affecting the future use of the pre-emptive strategy will depend on safety, cost, and tolerability of next-generation DAAs in relation to typical liver graft damage incurred before assessment of HCV recurrence.

Conversely, treating HCV recurrence has been the standard therapy and, until 2011, involved 48 weeks of PEG-interferon and ribavirin treatment. Three systematic reviews^{26,27,46} have reported an SVR rate in these conditions of only approximately 30% with limitations of tolerability including bacterial infections, haematological toxicity, and graft rejection. Early virological response (EVR) is a major predictive factor associated with SVR.47,48 However, effective antiviral treatment after transplantation has clear benefits in preventing disease progression. 49-54 Firstgeneration protease inhibitors, telaprevir or boceprevir, were the first drugs to be tested in the treatment of recurrent HCV after transplantation. Their inclusion with PEG-interferon and ribavirin improved SVR rates by 50-65% in recipients infected with genotype 1 HCV, but with a worse safety profile and potent drug-drug interactions.55,56 Although feasible, these regimens required close monitoring and expertise of caregivers leading to their withdrawal. The development of sofosbuvir added to ribavirin further changed the treatment landscape in post-transplantation antiviral administration, which is shown in two studies^{28,56} where initiation of treatment took place a year after liver engraftment. Sofosbuvir plus ribavirin treatment has a 70% efficacy rate in yielding SVR that is roughly equal to the virological response seen when clearing the virus before transplantation.²⁸ Although this response efficacy is not optimum, it shows efficacy and tolerability even in the most severe patients.⁵⁷ However, a more complex combination of DAAs can be more efficacious, and several studies describing this finding have already been published (table). Data come from both open-label studies and real-life cohorts (HCV-TARGET⁶¹ and CUPILT^{58,62} studies). The SVR12 rates are usually greater than 90%,

better than SVR12 rate treating patients with decompensated cirrhosis prior to liver transplantation, and well tolerated. In the SOLAR-1 study³⁰ assessing post-transplantation treatment, progressive liver disease was associated with lowered response; however, all six individuals who had fibrosing cholestatic hepatitis achieved SVR 12 weeks after the end of treatment. Although the treatment of HCV in transplant patients has been substantially improved and simplified, several issues remain to be clarified. The efficacy of these studies shows the promise of DAAs and combinatorial therapy; multiple targets and mechanisms of action synergise to eliminate the virus.

The optimum duration of therapy remains to be defined. Although some risk factors of treatment failure were identified for interferon-based regimens, no risk factors have been identified for new DAAs, except in HCV genotype 3 infection. In the non-transplant setting, most studies comparing different treatment durations did not show any benefit of longer treatment, and better adherence, fewer side-effects, and lower cost were associated with a shorter duration. In the transplant setting, robust data are scarce and many studies conservatively use 24 weeks of treatment in this special population until more evidence is collected.

Benefits of the use of ribavirin in future regimens are not yet established and use of ribavirin could be abandoned once next-generation DAAs with higher efficacy are added. There remains a great benefit of ribavirin in patients with severe liver disease and recurrent HCV post-transplantation.^{34,63,64}

In liver transplant patients, renal impairment is common and should be properly assessed before initiating antiviral therapy, especially sofosbuvir-based regimens.65 The metabolism of sofosbuvir takes place in the kidney and its use is not recommended in patients with creatinine clearance below 30 mL/min until an appropriate dose is determined. A phase 2b, open-label study of 200 mg or 400 mg sofosbuvir and ribavirin for 24 weeks in patients infected with HCV genotype 1 or 3, and ledipasvir or sofosbuvir in individuals with genotype 1 and 4 infection with renal insufficiency is ongoing (ClinicalTrials.gov, number NCT01958281). For other available DAAs, the metabolism takes place in the liver. Although no detrimental effect is expected in ribavirin-free combination, the ANRS C023 CUPILT group reported a slight, but significant, reduction in creatinine clearance during treatment 72.7 [SD 29.0] to 66.3 [25.7] mL/min between baseline and end of treatment, p<0.0001) using the combination of sofosbuvir and daclatasvir.58 Further studies are needed to clarify if this decrease in clearance is seen in all patients or one subgroup that should be identified and monitored closely. In a multicentre trial⁶⁶ of liver transplant recipients with recurrent HCV infection treated with sofosbuvir based regimens, renal improvement was observed in 58% of patients. Patients with SVR at 12 weeks post-treatment were more likely to have renal improvement, indicating that HCV affects renal health.⁶⁶

Drug-drug interactions between DAAs and immunosuppressive drugs, mainly calcineurin inhibitors, remain a concern with these regimens. Simeprevir, a secondgeneration protease inhibitor, is a partial cytochrome P450 (CYP) 3A inhibitor. Because the immunosuppressant drug ciclosporin is likewise a partial CYP3A inhibitor, combination results in accumulation of both drugs in the blood, and coadministration is discouraged. Ombitasvir, paritaprevir, ritonavir, and dasabuvir require dosing modifications for calcineurin inhibitors tacrolimus and ciclosporin. 59,68 Conversely, sofosbuvir, ledipasvir, and daclatasvir do not seem to interact with calcineurin inhibitors.68 However, close monitoring before, during, and after DAA therapy is essential. In the ANRS C023 CUPILT study,58 59% of 130 patients treated with sofosbuvir and daclatasvir after liver transplantation had to change dose of one immunosuppressive drug during therapy.

Finally, the optimum timing for initiation of therapy post-transplantation remains to be determined. Antiviral therapy is usually initiated only when histologically proven recurrent HCV occurs (fibrosis stage 2 or higher on the METAVIR score or severe and rapid progression of fibrosis as observed in cholestatic hepatitis). This decision was based on the tolerability of the classic interferonbased regimen, which required post-transplantation recovery time to restore health. Development of interferon-free therapy allows treatment of patients earlier after liver transplantation without waiting for disease markers indicating HCV recurrence. This strategy is reasonable, but scientific evidence of its efficacy is not available. Nonetheless, earlier treatment seems safe and effective and the potential risk of allowing fibrosis progression on liver graft could raise ethical issues. Treatment of patients early after liver transplantation could help to overcome the issue of differentiating HCV recurrence and rejection and it could also prevent graft rejection induced by viral clearance while immunosuppressive drug concentrations are still high at early stages after liver transplantation. Immunosuppressive drug concentrations have been reported to decrease substantially in patients responding to antiviral therapy as viral clearance improves hepatic microsomal function and elevated regulatory T-cell levels could decline.69

Prevention of graft infection concomitant with transplantation: HCV entry inhibitors

Viral entry has been shown to play an essential role during reinfection of the graft after liver transplantation. Thus, concomitant treatment of safe and effective entry inhibitors, including virus-targeting neutralising antibodies (nAbs), during and immediately after transplantation could prove an effective means of preventing graft infection without allowing allograft

damage.72 This strategy is supported by the experience in prevention of hepatitis B virus (HBV) graft infection where hepatitis B immune globulin in combination with nucleoside or nucleotide analogues can reduce HBV recurrence in liver transplant patients to 4%.71,73-75 Entry inhibitors have been shown to effectively inhibit HCV infection, work synergistically with DAAs, and have proved to be safe and effective in humanised mice.76 Although most of these drugs are in preclinical development, the results of the first clinical trials with anti-envelope antibodies^{22,23,77} and a small molecule hosttargeting inhibitor²⁴ suggest that they could be future instruments in the antiviral arsenal during transplantation. Strategies for blocking viral entry during liver graft infection can target either the virus or host entry factors.

Anti-envelope antibodies

Viral diversity, high rate of glycosylation of HCV glycoproteins, and association with apolipoproteins helps the escape of HCV from nAbs. 78-84 During HCV infection, nAbs develop that mostly target regions of E2 that

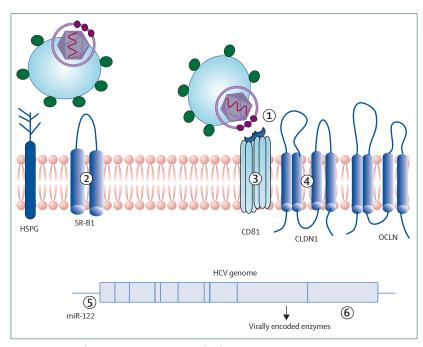


Figure 2: HCV entry factors as targets to prevent graft infection

Several points of HCV entry can be effective targets to prevent initial or ongoing liver graft infection. (1) The HCV glycoprotein E1/E2 is crucial for HCV entry, and nAbs binding to E1/E2 have proved effective in animal models and clinically. ^{22,23,90,91,94} (2) Early steps of HCV entry probably involve initial attachment of apolipoprotein E to HSPG and utilisation of SR-B1. SR-B1 inhibitors have been effective in animal models and in the clinic in the context of liver transplantation. ^{24,25,91,90} (3) HCV E2 directly binds to host entry factor CD81, and antibodies binding CD81 prevent HCV infection in animal models. ^{201,209} (4) Antibodies recognising CLDN1 have proved effective in preventing and curing HCV infection in animal models. ^{201,209} Furthermore, small molecules such as erlotinib targeting epidermal growth factor receptor, ²⁰⁶ a kinase promoting CD81-claudin-1 co-receptor formation, and ezetimibe targeting cholesterol transporter NP1CL1 (not shown) have been shown to inhibit HCV infection in humanised mouse models. ²⁰⁷ (5) Downstream of cell entry, miR-122 antagonists (antagomirs) have been shown to be effective and safe for use in animals and patients. ^{208,109} (6) DAAs targeting virally encoded enzymes have revolutionised HCV treatment. nAbs=neutralising antibodies. HSPG=heparan sulfate proteoglycans. SR-B1=scavenger receptor B1. CLDN1=claudin 1. OCLN=occludin. miR=microRNA. HCV=hepatitis C virus.

interact with the host receptor CD81.79,85-87 The crystal structure of the core of glycoprotein E288,89 defined the area of the protein to which most nAbs bind. Polyclonal and monoclonal nAbs isolated from patients with chronic HCV infection or given by gene therapeutic approaches are capable of inhibiting infection of human liver chimeric mice. 90-95 In patients, nAbs targeting the HCV envelope glycoprotein (MBL-HCV1) effectively delayed viral rebound, proving the principle that immunotherapy will be an effective addition to the synergistic antiviral arsenal.²² Studies are underway that combine MBL-HCV1 with DAAs to optimise therapeutic efficacy of this approach with the latest tools. Clinical trials of human HCV immune globulin in combination with DAAs show that administration of the immune globulin is safe and more efficacious than DAAs alone.23 However, a potential challenge in the use of nAbs for prevention of infection is identical to the problem during chronic infection—ie, genetic adaptation enabling viral escape.²² Complementing anti-envelope antibodies, small molecules have been identified to interfere with viral entry. 96-98

Host-targeting entry inhibitors

One solution that could feasibly prevent viral escape from antiviral antibodies would be to target host entry factors (figure 2).72,110 Infection with HCV variants that escape host anti-envelope antibodies or are resistant to DAAs are effectively blocked by host-targeting entry inhibitors,70,76,111-113 Host-targeting agents have been investigated for several steps of viral entry. HCV virions circulate in dynamic complex with lipoproteins and apolipoproteins. 114,115 The first step of HCV attachment is mediated by apolipoprotein E binding to heparan sulfates on the basolateral surface of the hepatocyte.116 Inhibitors of heparan sulfate attachment, such as the green tea polyphenol epigallocatechin-3-gallate, are generally safe and can impair infection in cell culture systems, 117,118 although in HCV mouse models the addition of epigallocatechin-3-gallate adds no observable advantage compared with anti-envelope antibodies alone.119 The next step of the HCV entry process is interaction of the virion with scavenger receptor B1 (SR-B1). Antibodies against SR-B1 substantially inhibit HCV infection in small animal models 99,100 and prevent antiviral resistance to DAAs.¹²⁰ Inhibition of the lipid transfer activity of SR-B1 is sufficient to inhibit infection.121 A small chemical inhibitor of SR-B1, ITX5061, has been tested in patients undergoing transplantation with HCV infection.24 Patients with HCV genotype 1 infection on treatment had sustained reduction of viral load, and the genetic variation of the quasispecies was limited.24 After this initial attachment, a sequence of events takes place including the triggering of signalling pathways involving host kinases such as epidermal growth factor receptor to cluster essential entry factor claudin 1 (CLDN1) and CD81. 106,122 Erlotinib, a small molecule inhibitor of epidermal growth factor

receptor, has been shown to inhibit HCV infection in both cell culture and animal models. 106 Antibodies that recognise CD81 have also been shown in small animal models to inhibit HCV infection. 101,102 Anti-CLDN1-specific antibodies are not only effective in preventing HCV infection, but can also prevent and cure HCV infection in humanised mice in monotherapy without resistance and observable side-effects. 103-105 The anti-CLDN1 antibody has been shown to be highly synergistic with DAAs,76 and to prevent antiviral resistance by impairing viral spread. 112 There are several other host entry factors such as occludin. 123,124 Niemann-Pick C1-like 1,107 transferrin receptor 1,125 and serum response factor binding protein 1.126 A clinically approved small chemical inhibitor of Niemann-Pick C1-like 1 has likewise proved effective in small animal models and to synergise with DAA. 107,127 Future research will enable the discovery and development of host-targeting entry inhibitors of optimum safety in administration and efficiency in synergising with DAAs to play a key role in increasing the cure rates in liver transplantation.

Advantages and disadvantages

The success of next-generation DAAs in treatment raises the debate of whether HCV entry inhibitors have a place in future clinical practice. 127,128 Advantages and disadvantages of using entry inhibitors exist. 72,129 Strengths include their capacity to be used in targeting intervention around transplantation with a short duration of treatment. The barrier for resistance, inhibiting the potential adaptation of viral mutant strains resistant to treatment, appears to be higher for host-targeting entry inhibitors than for DAA when used in monotherapy. 103,112 Given their complementary mechanism of action to DAAs¹¹⁰ and efficacy against DAA-resistant viruses, ^{112,130} entry inhibitors could be used in patients who do not respond to pre-emptive therapy, while preventing costly post-transplant therapy. The high level of synergy of entry inhibitors with DAAs seen in cell culture and animal models suggests that these agents could shorten treatment time and circumvent the development of antiviral-resistant variants. 76,103,120 One drawback is that a number of entry inhibitors are only now reaching clinical development stages. There are more DAAs in the drug development pipeline that might not be limited by the current safety issues of resistance and complications of renal failure. Targeting host factors will require careful surveillance of side-effects110 and viral escape has been described. 22,130-133 Furthermore, the numbers of individuals with serious HCV-related liver disease will decline, as will prices for the DAA regimens. Other approaches targeting host factors downstream of HCV cell entry^{110,134} include microRNA (miRNA) antagonists (antagomirs) or cyclophilin inhibitors. 135,136 A key miRNA that boosts HCV replication, miR-122, acts by shifting HCV genome activity away from translation and toward replication.¹³⁷ Antisense agents targeting

Search strategy and selection criteria

We searched PubMed for articles published from January, 1995, to January, 2016, with the terms "hepatitis C", "transplantation", "HCV", "liver graft", and "cirrhosis". Relevant presentations of upcoming publications were identified at the European Association for the Study of the Liver International Liver Congresses (2012–15) and the American Association for the Study of Liver Diseases Liver Meetings (2012–16). Articles resulting from these searches and relevant references cited in those articles were reviewed. Articles published in English, French, and German were included.

miR-122 have been shown to be safe and efficient in primate models and patients. ^{108,109,138} Cyclophilin, a host factor required for viral replication, is efficiently inhibited by alisporivir, a host-targeting agent in clinical development. ¹³⁹ Whether these strategies are feasible in liver transplantation remains to be shown.

Summary and conclusion

Treatment of patients in need of liver transplant as a result of HCV-associated advanced disease is a sensitive and complicated tree of decision making. The successful use of DAAs as prophylactic and therapeutic agents against HCV infection, both before and after transplantation, promises to assist increasing numbers of individuals who find themselves in this historically difficult-to-treat population. Randomised clinical trials are ongoing to define the role of entry inhibitors in the prevention of graft infection.

Contributors

All authors contributed to the writing and editing of the manuscript. DJF and AC contributed equally. DS and TFB contributed equally.

Declaration of interests

AC has served as a consultant and has received speaker fees from Astellas, Novartis, AbbVie, Bristol-Myers Squibb, Gilead, and Janssen-Cilag. RTC reports grants from from Gilead, Merck, Bristol-Myers Squibb, Mass Biologics, and Janssen; grants and personal fees from AbbVie; and personal fees from Idenix. DS has served as a consultant for Astellas, Bristol-Myers Squibb, Gilead, LFB, MSD, Novartis, Roche, Biotest, and AbbVie. TFB reports personal fees from Gilead and Vironexx; grants and personal fees from Biotest; and has a patent WO2010034812, US 8,518,408 issued for anti-host cell factor antibodies and small molecules for prevention and treatment of HCV infection. DJF declares no competing interests.

Acknowledgments

We thank Mirjam Zeisel and Che Colpitts (Inserm U1110, University of Strasbourg, France) for critical reading of the manuscript. TFB acknowledges grant support from the European Union (ERC-2008-AdG-HEPCENT, ERC-AdG-2014-671231-HEPCIR, FP7 HepaMab, and Interreg IV FEDERHepato-Regio-Net 2012, H2020-2015-667273-HEP-CAR), the National Institutes of Heatlth NIAID U19A1123862, the Agence Nationale de Recherche sur le SIDA et les hépatities virales (ANRS), and the Direction Générale de l'Offre de Soins (A12027MS). RTC receives grant support from NIH DK078772. This work has been published under the framework of the LABEX ANR-10-LABX-0028_HEPSYS and benefits from funding from the state managed by the French National Research Agency as part of the Investments for the Future Programme.

References

- Martin P, DiMartini A, Feng S, Brown R Jr, Fallon M. Evaluation for liver transplantation in adults: 2013 practice guideline by the American Association for the Study of Liver Diseases and the American Society of Transplantation. Hepatology 2014; 59: 1144–65.
- Watt KD, Pedersen RA, Kremers WK, Heimbach JK, Charlton MR. Evolution of causes and risk factors for mortality post-liver transplant: results of the NIDDK long-term follow-up study. Am J Transplant 2010; 10: 1420–27.
- 3 Charlton M, Ruppert K, Belle SH, et al. Long-term results and modeling to predict outcomes in recipients with HCV infection: results of the NIDDK liver transplantation database. *Liver Transpl* 2004; 10: 1120–30.
- 4 Gane EJ. The natural history of recurrent hepatitis C and what influences this. Liver Transpl 2008; 14 (suppl 2): S36–44.
- 5 Kim WR. The burden of hepatitis C in the United States. Hepatology 2002; 36 (suppl 1): S30–34.
- 6 Berenguer M, Prieto M, Rayon JM, et al. Natural history of clinically compensated hepatitis C virus-related graft cirrhosis after liver transplantation. *Hepatology* 2000; 32: 852–58.
- 7 Adam R, Karam V, Delvart V, et al. Evolution of indications and results of liver transplantation in Europe. A report from the European Liver Transplant Registry (ELTR). J Hepatol 2012; 57: 675–88.
- 8 Deuffic-Burban S, Mathurin P, Rosa I, et al. Impact of emerging hepatitis C virus treatments on future needs for liver transplantation in France: a modelling approach. *Dig Liver Dis* 2014; 46: 157–63.
- 9 Denniston MM, Klevens RM, McQuillan GM, Jiles RB. Awareness of infection, knowledge of hepatitis C, and medical follow-up among individuals testing positive for hepatitis C: National Health and Nutrition Examination Survey 2001–2008. Hepatology 2012; 55: 1652–61.
- 10 Wedemeyer H, Duberg AS, Buti M, et al. Strategies to manage hepatitis C virus (HCV) disease burden. J Viral Hepat 2014; 21 (suppl 1): S60–89.
- Forns X, Garcia-Retortillo M, Serrano T, et al. Antiviral therapy of patients with decompensated cirrhosis to prevent recurrence of hepatitis C after liver transplantation. J Hepatol 2003; 39: 389–96.
- 12 Forman LM, Lewis JD, Berlin JA, Feldman HI, Lucey MR. The association between hepatitis C infection and survival after orthotopic liver transplantation. *Gastroenterology* 2002; 127: 889–96
- 13 Prieto M, Berenguer M, Rayon JM, et al. High incidence of allograft cirrhosis in hepatitis C virus genotype 1b infection following transplantation: relationship with rejection episodes. *Hepatology* 1999; 29: 250–56.
- 14 Carrion JA, Navasa M, Forns X. Retransplantation in patients with hepatitis C recurrence after liver transplantation. J Hepatol 2010; 53: 962-70
- Marino Z, Mensa L, Crespo G, et al. Early periportal sinusoidal fibrosis is an accurate marker of accelerated HCV recurrence after liver transplantation. J Hepatol 2014; 61: 270–77.
- 16 Carrion JA, Torres F, Crespo G, et al. Liver stiffness identifies two different patterns of fibrosis progression in patients with hepatitis C virus recurrence after liver transplantation. *Hepatology* 2010: 51: 23–34.
- Blasco A, Forns X, Carrion JA, et al. Hepatic venous pressure gradient identifies patients at risk of severe hepatitis C recurrence after liver transplantation. *Hepatology* 2006; 43: 492–99.
- 18 Berenguer M, McCaughan G. Hepatitis C virus-associated cholestatic hepatitis: we cannot seem to agree on diagnostic criteria. *Liver Transpl* 2013; 19: 115–17.
- 19 Charlton M, Seaberg E, Wiesner R, et al. Predictors of patient and graft survival following liver transplantation for hepatitis C. Hepatology 1998; 28: 823–30.
- 20 Fukuhara T, Taketomi A, Motomura T, et al. Variants in IL28B in liver recipients and donors correlate with response to peg-interferon and ribavirin therapy for recurrent hepatitis C. Gastroenterology 2010: 139: 1577–85.
- Veldt BJ, Duarte-Rojo A, Thompson AJ, et al. Recipient IL28B polymorphism is an important independent predictor of posttransplant diabetes mellitus in liver transplant patients with chronic hepatitis C. Am J Transplant 2012; 12: 737–44.

- 22 Chung RT, Gordon FD, Curry MP, et al. Human monoclonal antibody MBL-HCV1 delays HCV viral rebound following liver transplantation: a randomized controlled study. Am J Transplant 2013; 13: 1047–54.
- 23 Terrault N, Satapathy SK, Therapondos G, et al. Prevention of hepatitis C virus (HCV) recurrence with peri-transplant hepatitis C immune globulin combined with pre-transplant (Pre-LT) antiviral therapy (AVT). Hepatology 2014; 60 (suppl 1): 206A.
- 24 Rowe IA, Tully DC, Armstrong MJ, et al. Effect of scavenger receptor class B type I antagonist ITX5061 in patients with hepatitis C virus infection undergoing liver transplantation. *Liver Transpl* 2016; 22: 287-97
- 25 Rowe I, Armstrong M, Parker R, et al. Effect of scavenger receptor B-I antagonist, ITX5061, on early hepatitis C virus kinetics in liver transplantation: results of a phase 1b clinical trial. *Lancet* 2014; 383: S90 (abstr).
- 26 Berenguer M. Systematic review of the treatment of established recurrent hepatitis C with pegylated interferon in combination with ribavirin. J Hepatol 2008; 49: 274–87.
- Wang CS, Ko HH, Yoshida EM, Marra CA, Richardson K. Interferon-based combination anti-viral therapy for hepatitis C virus after liver transplantation: a review and quantitative analysis. Am J Transplant 2006; 6: 1586–99.
- 28 Curry MP, Forns X, Chung RT, et al. Sofosbuvir and ribavirin prevent recurrence of HCV infection after liver transplantation: an open-label study. *Gastroenterology* 2015; 148: 100–07.
- 29 Welker MW, Luhne S, Lange CM, et al. Lactic acidosis in patients with hepatitis C virus cirrhosis and combined ribavirin/sofosbuvir treatment. J Hepatol 2016; 64: 790–99.
- 30 Charlton M, Everson GT, Flamm SL, et al. Ledipasvir and sofosbuvir plus ribavirin for treatment of HCV infection in patients with advanced liver disease. *Gastroenterology* 2015; 149: 649–59.
- 31 Manns M, Forns X, Samuel D, et al. Ledipasvir/sofosbuvir with ribavirin is safe and efficacious in decompensated and post liver transplantation patients with HCV infection: preliminary results of the prospective SOLAR 2 trial. J Hepatol 2015; 62 (suppl 2): S187.
- 32 Gambato M, Villanueva A, Abraldes JG, Altamirano J, Forns X. Clinical trial watch: reports from the EASL International Liver Congress (ILC), Vienna, April 2015. J Hepatol 2015; 63: 753–62.
- 33 Poordad F, Schiff ER, Vierling JM, et al. Daclatasvir with sofosbuvir and ribavirin for HCV infection with advanced cirrhosis or post-liver transplant recurrence. *Hepatology* 2016; published online Jan 11. DOI:10.1002/hep.28446.
- 34 Foster GR, Irving WL, Cheung MC, et al. Impact of direct acting antiviral therapy in patients with chronic hepatitis C and decompensated cirrhosis. *J Hepatol* 2016; published online Jan 30. DOI: 10.1016/j.jhep.2016.01.029.
- 35 Foster GR, Afdhal N, Roberts SK, et al. Sofosbuvir and velpatasvir for HCV genotype 2 and 3 infection. N Engl J Med 2015; 373: 2608–17.
- 36 Feld JJ, Jacobson IM, Hezode C, et al. Sofosbuvir and velpatasvir for HCV genotype 1, 2, 4, 5, and 6 infection. N Engl J Med 2015; 373: 2599–607.
- 37 Curry MP, O'Leary JG, Bzowej N, et al. Sofosbuvir and velpatasvir for HCV in patients with decompensated cirrhosis. N Engl J Med 2015; 373: 2618–28.
- 38 Coilly A, Pageaux G, Houssel-Debry P, et al. Improving liver function and delisting of patients awaiting liver transplantation for HCV cirrhosis: do we ask too much to DAA? *Hepatology* 2015; 62 (suppl 1): 257A.
- 39 Munoz SJ, Reich DJ, Rothstein KD, et al. Curing decompensated wait listed HCV patients with the new DAAs: the potential significant impact on liver transplant wait list and organ allocation. Hepatology 2015; 62 (suppl 1): 311A.
- 40 Toso C, Dupuis-Lozeron E, Majno P, et al. A model for dropout assessment of candidates with or without hepatocellular carcinoma on a common liver transplant waiting list. *Hepatology* 2012; 56: 149–56.
- 41 Bzowej N, Nelson DR, Terrault NA, et al. PHOENIX: a randomized controlled trial of peginterferon alfa-2a plus ribavirin as a prophylactic treatment after liver transplantation for hepatitis C virus. Liver Transpl 2011; 17: 528–38.
- 42 Chalasani N, Manzarbeitia C, Ferenci P, et al. Peginterferon alfa-2a for hepatitis C after liver transplantation: two randomized, controlled trials. *Hepatology* 2005; 41: 289–98.

- 43 Mazzaferro V, Tagger A, Schiavo M, et al. Prevention of recurrent hepatitis C after liver transplantation with early interferon and ribavirin treatment. *Transplant Proc* 2001; 33: 1355–57.
- 44 Shergill AK, Khalili M, Straley S, et al. Applicability, tolerability and efficacy of preemptive antiviral therapy in hepatitis C-infected patients undergoing liver transplantation. Am J Transplant 2005; 5: 118–24.
- 45 Sugawara Y, Makuuchi M, Matsui Y, et al. Preemptive therapy for hepatitis C virus after living-donor liver transplantation. *Transplantation* 2004; 78: 1308–11.
- 46 Xirouchakis E, Triantos C, Manousou P, et al. Pegylated-interferon and ribavirin in liver transplant candidates and recipients with HCV cirrhosis: systematic review and meta-analysis of prospective controlled studies. J Viral Hepat 2008; 15: 699–709.
- 47 Berenguer M, Palau A, Fernandez A, et al. Efficacy, predictors of response, and potential risks associated with antiviral therapy in liver transplant recipients with recurrent hepatitis C. *Liver Transpl* 2006; 12: 1067–76.
- 48 Roche B, Sebagh M, Canfora ML, et al. Hepatitis C virus therapy in liver transplant recipients: response predictors, effect on fibrosis progression, and importance of the initial stage of fibrosis. Liver Transpl 2008; 14: 1766–77.
- 49 Abdelmalek MF, Firpi RJ, Soldevila-Pico C, et al. Sustained viral response to interferon and ribavirin in liver transplant recipients with recurrent hepatitis C. Liver Transpl 2004; 10: 199–207.
- 50 Bizollon T, Ahmed SN, Radenne S, et al. Long term histological improvement and clearance of intrahepatic hepatitis C virus RNA following sustained response to interferon-ribavirin combination therapy in liver transplanted patients with hepatitis C virus recurrence. Gut 2003; 52: 283–87.
- 51 Firpi RJ, Abdelmalek MF, Soldevila-Pico C, et al. Combination of interferon alfa-2b and ribavirin in liver transplant recipients with histological recurrent hepatitis C. Liver Transpl 2002; 8: 1000–06.
- 52 Picciotto FP, Tritto G, Lanza AG, et al. Sustained virological response to antiviral therapy reduces mortality in HCV reinfection after liver transplantation. J Hepatol 2007; 46: 459–65.
- 53 Walter T, Scoazec JY, Guillaud O, et al. Long-term antiviral therapy for recurrent hepatitis C after liver transplantation in nonresponders: biochemical, virological, and histological impact. *Liver Transpl* 2009; 15: 54–63.
- 54 Carrion JA, Navasa M, Garcia-Retortillo M, et al. Efficacy of antiviral therapy on hepatitis C recurrence after liver transplantation: a randomized controlled study. Gastroenterology 2007; 132: 1746–56.
- 55 Coilly A, Dumortier J, Botta-Fridlund D, et al. Multicenter experience with boceprevir or telaprevir to treat hepatitis C recurrence after liver transplantation: when present becomes past, what lessons for future? PLoS One 2015; 10: e0138091.
- 56 Charlton M, Gane E, Manns MP, et al. Sofosbuvir and ribavirin for treatment of compensated recurrent hepatitis C virus infection after liver transplantation. Gastroenterology 2015; 148: 108–17.
- 57 Forns X, Charlton M, Denning J, et al. Sofosbuvir compassionate use program for patients with severe recurrent hepatitis C after liver transplantation. *Hepatology* 2015; 61: 1485–94.
- 58 Coilly A, Fougerou C, De Ledinghen V, et al. The association of sofosbuvir and daclatasvir for treating severe recurrence of HCV infection after liver translplantation; results from a large French prospective multicentric ANRS CO23 CUPILT cohort. J Hepatol 2015; 62 (suppl 2): S235.
- 59 Kwo PY, Mantry PS, Coakley E, et al. An interferon-free antiviral regimen for HCV after liver transplantation. N Engl J Med 2014; 371: 2375–82.
- 60 Pungpapong S, Werner KT, Aqel B, et al. Multicenter experience using sofosbuvir and simeprevir with/without rivavirin to treat HCV genotype 1 after liver transplantation. *Hepatology* 2014; 60 (suppl 1): 201A.
- 61 Brown RS Jr, O'Leary JG, Reddy KR, et al. Interferon-free therapy for genotype 1 hepatitis C in liver transplant recipients: real-world experience from the hepatitis C therapeutic registry and research network. Liver Transpl 2016; 22: 24–33.
- 62 Leroy V, Dumortier J, Coilly A, et al. Efficacy of sofosbuvir and daclatasvir in patients with fibrosing cholestatic hepatitis C after liver transplantation. Clin Gastroenterol Hepatol 2015; 13: 1993–2001.

- 63 Sulkowski MS, Eron JJ, Wyles D, et al. Ombitasvir, paritaprevir co-dosed with ritonavir, dasabuvir, and ribavirin for hepatitis C in patients co-infected with HIV-1: a randomized trial. *JAMA* 2015; 313: 1223–31.
- 64 Reddy KR, Bourliere M, Sulkowski M, et al. Ledipasvir and sofosbuvir in patients with genotype 1 hepatitis C virus infection and compensated cirrhosis: an integrated safety and efficacy analysis. *Hepatology* 2015; 62: 79–86.
- 65 Ojo AO, Held PJ, Port FK, et al. Chronic renal failure after transplantation of a nonrenal organ. N Engl J Med 2003; 349: 931–40.
- 66 Faisal N, Renner EL, Bilodeau M, et al. Impact of sofosbuvir-based regimens on renal function in liver transplant recipients: results of a multicenter study. *Hepatology* 2015; 62 (suppl 1): 312A.
- 67 Ouwerkerk-Mahadevan S, Snoeys J, Peeters M, Beumont-Mauviel M, Simion A. Drug-drug interactions with the NS3/4A protease inhibitor simeprevir. *Clin Pharmacokinet* 2015; 55: 197–208.
- 68 Kwo PY, Badshah MB. New hepatitis C virus therapies: drug classes and metabolism, drug interactions relevant in the transplant settings, drug options in decompensated cirrhosis, and drug options in end-stage renal disease. Curr Opin Organ Transplant 2015; 20: 235–41.
- 69 Kugelmas M, Osgood MJ, Trotter JF, et al. Hepatitis C virus therapy, hepatocyte drug metabolism, and risk for acute cellular rejection. *Liver Transpl* 2003; 9: 1159–65.
- 70 Fafi-Kremer S, Fofana I, Soulier E, et al. Viral entry and escape from antibody-mediated neutralization influence hepatitis C virus reinfection in liver transplantation. J Exp Med 2010; 207: 2019–31.
- 71 Fofana I, Fafi-Kremer S, Carolla P, et al. Mutations that alter use of hepatitis C virus cell entry factors mediate escape from neutralizing antibodies. Gastroenterology 2012; 143: 223–33.
- 72 Fofana I, Jilg N, Chung RT, Baumert TF. Entry inhibitors and future treatment of hepatitis C. Antiviral Res 2014; 104: 136–42.
- 73 Gane EJ, Angus PW, Strasser S, et al. Lamivudine plus low-dose hepatitis B immunoglobulin to prevent recurrent hepatitis B following liver transplantation. *Gastroenterology* 2007; **132**: 931–37.
- 74 Crespo G, Marino Z, Navasa M, Forns X. Viral hepatitis in liver transplantation. Gastroenterology 2012; 142: 1373–83.
- 75 Takaki A, Yasunaka T, Yagi T. Molecular mechanisms to control posttransplantation hepatitis B recurrence. Int J Mol Sci 2015; 16: 17494–513.
- 76 Xiao F, Fofana I, Thumann C, et al. Synergy of entry inhibitors with direct-acting antivirals uncovers novel combinations for prevention and treatment of hepatitis C. Gut 2015; 64: 483–94.
- 77 Davis GL, Nelson DR, Terrault N, et al. A randomized, open-label study to evaluate the safety and pharmacokinetics of human hepatitis C immune globulin (Civacir) in liver transplant recipients. Liver Transpl 2005; 11: 941–49.
- 78 Helle F, Vieyres G, Elkrief L, et al. Role of N-linked glycans in the functions of hepatitis C virus envelope proteins incorporated into infectious virions. J Virol 2010; 84: 11905–15.
- 79 von Hahn T, Yoon JC, Alter H, et al. Hepatitis C virus continuously escapes from neutralizing antibody and T-cell responses during chronic infection in vivo. *Gastroenterology* 2007; 132: 667–78.
- 80 Zhang P, Zhong L, Struble EB, et al. Depletion of interfering antibodies in chronic hepatitis C patients and vaccinated chimpanzees reveals broad cross-genotype neutralizing activity. Proc Natl Acad Sci USA 2009; 106: 7537–41.
- 81 Baumert TF, Fauvelle C, Chen DY, Lauer GM. A prophylactic hepatitis C virus vaccine: a distant peak still worth climbing. *J Hepatol* 2014; 61 (suppl 1): S34–44.
- 82 Fauvelle C, Felmlee DJ, Crouchet E, et al. Apolipoprotein E mediates evasion from hepatitis C virus neutralizing antibodies. Gastroenterology 2016; 150: 206–17.
- 83 Bankwitz D, Steinmann E, Bitzegeio J, et al. Hepatitis C virus hypervariable region 1 modulates receptor interactions, conceals the CD81 binding site, and protects conserved neutralizing epitopes. J Virol 2010; 84: 5751–63.
- 84 Keck ZY, Girard-Blanc C, Wang W, et al. Antibody response to hypervariable region 1 interferes with broadly neutralizing antibodies to hepatitis C virus. J Virol 2016; 90: 3112–22.
- 85 Owsianka AM, Tarr AW, Keck ZY, et al. Broadly neutralizing human monoclonal antibodies to the hepatitis C virus E2 glycoprotein. J Gen Virol 2008; 89: 653–59.

- 86 Zhang J, Randall G, Higginbottom A, Monk P, Rice CM, McKeating JA. CD81 is required for hepatitis C virus glycoprotein-mediated viral infection. J Virol 2004; 78: 1448–55.
- 87 Haberstroh A, Schnober EK, Zeisel MB, et al. Neutralizing host responses in hepatitis C virus infection target viral entry at postbinding steps and membrane fusion. *Gastroenterology* 2008; 135: 1719–28.
- 88 Kong L, Giang E, Nieusma T, et al. Hepatitis C virus E2 envelope glycoprotein core structure. Science 2013; 342: 1090–94.
- 89 Khan AG, Whidby J, Miller MT, et al. Structure of the core ectodomain of the hepatitis C virus envelope glycoprotein 2. Nature 2014; 509: 381–84.
- 90 Law M, Maruyama T, Lewis J, et al. Broadly neutralizing antibodies protect against hepatitis C virus quasispecies challenge. Nat Med 2008: 14: 25–27.
- 91 Vanwolleghem T, Bukh J, Meuleman P, et al. Polyclonal immunoglobulins from a chronic hepatitis C virus patient protect human liver-chimeric mice from infection with a homologous hepatitis C virus strain. Hepatology 2008; 47: 1846–55.
- 92 Akazawa D, Moriyama M, Yokokawa H, et al. Neutralizing antibodies induced by cell culture-derived hepatitis C virus protect against infection in mice. Gastroenterology 2013; 145: 447–55.
- 93 Meuleman P, Bukh J, Verhoye L, et al. In vivo evaluation of the cross-genotype neutralizing activity of polyclonal antibodies against hepatitis C virus. *Hepatology* 2011; 53: 755–62.
- 94 de Jong YP, Dorner M, Mommersteeg MC, et al. Broadly neutralizing antibodies abrogate established hepatitis C virus infection. Sci Transl Med 2014: 6: 254ra129.
- 95 Desombere I, Fafi-Kremer S, Van Houtte F, et al. Monoclonal anti-envelope antibody AP33 protects humanized mice against a patient-derived hepatitis C virus challenge. *Hepatology* 2016: 63: 1120–34
- 96 Perin PM, Haid S, Brown RJ, et al. Flunarizine prevents hepatitis C virus membrane fusion in a genotype-dependent manner by targeting the potential fusion peptide within E1. Hepatology 2016; 63: 49–62.
- 97 St Vincent MR, Colpitts CC, Ustinov AV, et al. Rigid amphipathic fusion inhibitors, small molecule antiviral compounds against enveloped viruses. Proc Natl Acad Sci USA 2010; 107: 17339–44.
- 98 He S, Lin B, Chu V, et al. Repurposing of the antihistamine chlorcyclizine and related compounds for treatment of hepatitis C virus infection. Sci Transl Med 2015; 7: 282ra49.
- 99 Meuleman P, Catanese MT, Verhoye L, et al. A human monoclonal antibody targeting scavenger receptor class B type I precludes hepatitis C virus infection and viral spread in vitro and in vivo. Hepatology 2012; 55: 364–72.
- 100 Vercauteren K, Van Den Eede N, Mesalam AA, et al. Successful anti-scavenger receptor class B type I (SR-BI) monoclonal antibody therapy in humanized mice after challenge with HCV variants with in vitro resistance to SR-BI-targeting agents. Hepatology 2014; 60: 1508–18.
- 101 Meuleman P, Hesselgesser J, Paulson M, et al. Anti-CD81 antibodies can prevent a hepatitis C virus infection in vivo. Hepatology 2008; 48: 1761–68.
- 102 Ji C, Liu Y, Pamulapati C, et al. Prevention of hepatitis C virus infection and spread in human liver chimeric mice by an anti-CD81 monoclonal antibody. *Hepatology* 2014; 61: 1136–44.
- 103 Mailly L, Xiao F, Lupberger J, et al. Clearance of persistent hepatitis C virus infection in humanized mice using a claudin-1-targeting monoclonal antibody. Nat Biotechnol 2015; 33: 549–54.
- 104 Yamashita M, Iida M, Tada M, et al. Discovery of anti-claudin-1 antibodies as candidate therapeutics against hepatitis C virus. J Pharmacol Exp Ther 2015; 353: 112–18.
- Fukasawa M, Nagase S, Shirasago Y, et al. Monoclonal antibodies against extracellular domains of claudin-1 block hepatitis C virus infection in a mouse model. J Virol 2015; 89: 4866–79.
- 106 Lupberger J, Zeisel MB, Xiao F, et al. EGFR and EphA2 are host factors for hepatitis C virus entry and possible targets for antiviral therapy. Nat Med 2011; 17: 589–95.
- 107 Sainz B, Jr, Barretto N, Martin DN, et al. Identification of the Niemann-Pick C1-like 1 cholesterol absorption receptor as a new hepatitis C virus entry factor. Nat Med 2012; 18: 281–85.
- 108 Lanford RE, Hildebrandt-Eriksen ES, Petri A, et al. Therapeutic silencing of microRNA-122 in primates with chronic hepatitis C virus infection. Science 2010; 327: 198–201.

- 109 Janssen HL, Reesink HW, Lawitz EJ, et al. Treatment of HCV infection by targeting microRNA. N Engl J Med 2013; 368: 1685–94.
- 110 Zeisel MB, Lupberger J, Fofana I, Baumert TF. Host-targeting agents for prevention and treatment of chronic hepatitis C perspectives and challenges. J Hepatol 2013; 58: 375–84.
- 111 Fofana I, Krieger SE, Grunert F, et al. Monoclonal anti-claudin antibodies prevent hepatitis C virus infection of primary human hepatocytes. *Gastroenterology* 2010; **139**: 953–64.
- 112 Xiao F, Fofana I, Heydmann L, et al. Hepatitis C virus cell-cell transmission and resistance to direct-acting antiviral agents. PLoS Pathog 2014; 10: e1004128.
- 113 Zeisel MB, Felmlee DJ, Baumert TF. Hepatitis C virus entry. Curr Top Microbiol Immunol 2013; 369: 87–112.
- .14 Andre P, Komurian-Pradel F, Deforges S, et al. Characterization of low- and very-low-density hepatitis C virus RNA-containing particles. J Virol 2002; 76: 6919–28.
- 115 Felmlee DJ, Sheridan DA, Bridge SH, et al. Intravascular transfer contributes to postprandial increase in numbers of very-low-density hepatitis C virus particles. *Gastroenterology* 2010; 139: 1774–83.
- 116 Jiang J, Cun W, Wu X, Shi Q, Tang H, Luo G. Hepatitis C virus attachment mediated by apolipoprotein E binding to cell surface heparan sulfate. J Virol 2012; 86: 7256–67.
- 117 Ciesek S, von Hahn T, Colpitts CC, et al. The green tea polyphenol, epigallocatechin-3-gallate, inhibits hepatitis C virus entry. *Hepatology* 2011; 54: 1947–55.
- 118 Colpitts CC, Schang LM. A small molecule inhibits virion attachment to heparan sulfate- or sialic acid-containing glycans. J Virol 2014; 88: 7806–17.
- 119 O'Shea D, Law J, Egli A, et al. Prevention of hepatitis C virus infection using a broad cross-neutralizing monoclonal antibody (AR4A) and epigallocatechin gallate. *Liver Transpl* 2016; 22: 324–32.
- 120 Vercauteren K, Brown RJ, Mesalam AA, et al. Targeting a host-cell entry factor barricades antiviral-resistant HCV variants from on-therapy breakthrough in human-liver mice. *Gut* 2015; published online Aug 25. DOI: 10.1136/gutjnl-2014-309045.
- 121 Zahid MN, Turek M, Xiao F, et al. The postbinding activity of scavenger receptor class B type I mediates initiation of hepatitis C virus infection and viral dissemination. *Hepatology* 2013; 57: 492–504.
- 122 Zona L, Lupberger J, Sidahmed-Adrar N, et al. HRas signal transduction promotes hepatitis C virus cell entry by triggering assembly of the host tetraspanin receptor complex. *Cell Host Microbe* 2013; **13**: 302–13.
- 123 Ploss A, Evans MJ, Gaysinskaya VA, et al. Human occludin is a hepatitis C virus entry factor required for infection of mouse cells. Nature 2009; 457: 882–86.
- 124 Benedicto I, Molina-Jimenez F, Bartosch B, et al. The tight junction-associated protein occludin is required for a postbinding step in hepatitis C virus entry and infection. J Virol 2009; 83: 8012–20.
- 125 Martin DN, Uprichard SL. Identification of transferrin receptor 1 as a hepatitis C virus entry factor. *Proc Natl Acad Sci USA* 2013; 110: 10777–82.
- 126 Gerold G, Meissner F, Bruening J, et al. Quantitative proteomics identifies serum response factor binding protein 1 as a host factor for hepatitis C virus entry. *Cell Rep* 2015; 12: 864–78.
- 127 Uprichard SL, Sainz B, Jr. Inhibition of hepatitis C entry: too soon to dismiss while many are still being denied treatment. Gut 2015; 64: 690–91.
- 128 Pawlotsky JM. Viral entry inhibition: too late for hepatitis C, but promising for other viral infections. Gut 2015; 64: 362–64.
- 129 Ploss A, Dubuisson J. New advances in the molecular biology of hepatitis C virus infection: towards the identification of new treatment targets. Gut 2012; 61 (suppl 1): i25–35.
- 130 Zhu H, Wong-Staal F, Lee H, et al. Evaluation of ITX 5061, a scavenger receptor B1 antagonist: resistance selection and activity in combination with other hepatitis C virus antivirals. J Infect Dis 2012: 205: 656–62.
- 131 Haid S, Grethe C, Dill MT, Heim M, Kaderali L, Pietschmann T. Isolate-dependent use of claudins for cell entry by hepatitis C virus. Hepatology 2014; 59: 24–34.
- 132 Hopcraft SE, Evans MJ. Selection of a hepatitis C virus with altered entry factor requirements reveals a genetic interaction between the E1 glycoprotein and claudins. *Hepatology* 2015; 62: 1059–69.

- 133 Colpitts CC, Zeisel MB, Baumert TF. When one receptor closes, another opens: Claudins and the hepatitis C virus E1 glycoprotein. Hepatology 2015; 62: 991–93.
- 134 Manns MP, von Hahn T. Novel therapies for hepatitis C—one pill fits all? Nat Rev Drug Discov 2013; 12: 595–610.
- 135 Bandiera S, Pfeffer S, Baumert TF, Zeisel MB. miR-122—a key factor and therapeutic target in liver disease. *J Hepatol* 2015;
- 136 Lin K, Gallay P. Curing a viral infection by targeting the host: the example of cyclophilin inhibitors. *Antiviral Res* 2013; **99**: 68–77.
- 137 Masaki T, Arend KC, Li Y, et al. miR-122 stimulates hepatitis C virus RNA synthesis by altering the balance of viral RNAs engaged in replication versus translation. *Cell Host Microbe* 2015; 17: 217–28.
- 138 Van Der Ree M, de Vree ML, Stelma F, et al. A single subcutaneous dose of 2mg/kg or 4mg/kg of RG-101, a GalNAc-conjugated oligonucleotide with antagonist activity against MIR-122, results in significant viral load reductions in chronic hepatitis C patients. *J Hepatol* 2015; 62 (suppl 2): S261.
- 139 Guedj J, Yu J, Levi M, et al. Modeling viral kinetics and treatment outcome during alisporivir interferon-free treatment in hepatitis C virus genotype 2 and 3 patients. *Hepatology* 2014; **59**: 1706–14.